

## Article

# Three Years After Soybean-Cover-Crop Rotation in Conventional and No-Till Practices: What Are the Consequences on Soil Nitrous Oxide Emissions?

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**Abstract:** Nitrous oxide is a potent greenhouse gas due to its long atmospheric lifespan (121 years) that results in a high global warming potential (GWP). Research has shown that no-tillage may be implemented as a mitigation strategy to reduce N<sub>2</sub>O emissions. The objective of the was to evaluate how conventional tillage (CT) and no-tillage (NT) can potential influence N<sub>2</sub>O emissions in soybean rotation in a semi-arid region of the central Free State of South Africa. The effect of conventional and no-till tillage practices on N<sub>2</sub>O emissions under soybean rotation was evaluated in the 3rd year of a 5-year rotation system, in a semi-arid region of the Free State of South Africa, from December 2022 to December 2023. The experimental area was divided into three blocks and there were two plots in each block: in total there were six plots. The treatments were planted in a soybean rotation system under no-tillage and conventional tillage. The monthly averages of N<sub>2</sub>O emissions were significantly different from each other during the soybean growing season; the highest emissions were recorded in August/September 2023 from both the NT and CT treatments after harvest. During this time, there were crop residues in the soil that increased soil carbon. There was a positive correlation between N<sub>2</sub>O emissions and soil carbon content ( $p = 0.21$ ) and between N<sub>2</sub>O emissions and soil organic matter ( $p = 0.43$ ). Emissions were significantly higher in CT (LSD = 0.3) than in NT. The lowest N<sub>2</sub>O emissions were recorded in December 2023 (LSD = 0.05) and were significantly reduced in the no-till plots compared to those of the conventional tillage plots. Furthermore, the lowest cumulative N<sub>2</sub>O emissions of  $0.26 \pm 0.22$  kg N<sub>2</sub>O-N ha<sup>-1</sup> were recorded during NT in the winter season and were significantly different from CT (LSD = 0.19). The results from our study indicate that the no-till practices in soybean rotation can decrease N<sub>2</sub>O emissions.

**Keywords:** nitrous oxide emissions; soybean; no-tillage; conventional tillage



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## 1. Introduction

The production of nitrous oxide (N<sub>2</sub>O) from agricultural soils is a significant environmental concern due to its role as a potent greenhouse gas (GHG) in the 21st century [1] and its contribution to the depletion of the ozone layer [2]. Crop production accounts for approximately 50% of N<sub>2</sub>O emissions from agriculture [1]. The production of N<sub>2</sub>O from

agricultural soils is mainly modulated through two microbial processes: denitrification and nitrification [3].

Nitrification plays an critical role in the nitrogen (N) cycle, influencing soil fertility and impacting environmental quality [4]. Nitrification is increased at 50–60% WFPS levels [5,6]. The process increases at temperatures between 20 °C and 30 °C and, below 10 °C and above 35 °C, it slows significantly [7,8]. Denitrification plays an important role in the N cycle, affecting soil fertility and contributing to environmental issues, such as GHG emissions, mainly of N<sub>2</sub>O [1,9–11]. High water-filled pore spaces (WFPS) (>60%), are conducive to denitrification [12] and temperatures between 25 °C and 30 °C [3,13–15]. High levels of nitrate (NO<sub>3</sub><sup>−</sup>) [16,17], NH<sub>4</sub><sup>+</sup> and carbon in the soil improve the denitrification rate [16,18,19]. Denitrification occurs under anaerobic soil conditions, whereas nitrification occurs under aerobic conditions [6]. Soils with an alkaline pH (7–8) are the most efficient in denitrification, and a pH of 6 to 7.5 favors nitrification [20–22].

Incorporating legumes such as soybeans into crop rotation systems is a production technique offering numerous benefits to soil health, crop productivity, and environmental sustainability [23,24]. It has also been reported to reduce N<sub>2</sub>O emissions in soils by improving nitrogen uptake, thus reducing excess nitrogen [25,26]. Crop rotation, including legume incorporation, can significantly influence N<sub>2</sub>O emissions from agricultural soils through reduced total synthetic N requirements [27,28]. Since legumes fix atmospheric N into the soil, increasing soil N and thus enriching the soil, this can also potentially increase N<sub>2</sub>O emissions if not properly managed [29–31].

Crop rotations that incorporate cover crops and legumes improve soil organic matter, enhancing soil health and structure [32,33]. Improved soil structure helps improve water infiltration and drainage, reducing anaerobic conditions that favor denitrification and N<sub>2</sub>O production and subsequent emissions [34–37]. Rotating crops promote a diverse microbial community in the soil, which can improve nutrient cycling and reduce the likelihood of conditions that favor N<sub>2</sub>O production [37–39]. A healthy and diverse microbial ecosystem can process nitrogen more effectively, reducing the accumulation of nitrates that lead to N<sub>2</sub>O emissions [40–42]. Different crops leave different types, qualities, and amounts of residues. Managing these residues properly (e.g., timely incorporation) can influence microbial activity and nitrogen mineralization, affecting N<sub>2</sub>O emissions [43–46].

Previous studies associated no-till soybean production with reduced N<sub>2</sub>O emissions compared to conventional cropping in multiple-location systems. No-tillage also reduces the cost incurred regarding fuel for cultivation. It improves soil health and structure, as minimal soil disturbance increases soil organic matter and biological activity. This promotes water infiltration. Soil moisture is conserved, since crop residues are left on the soil surface, particularly in rainfed and drought prone areas. In the long-term, yield stability is achieved, since no-till enhances soil resilience, thus improving crop performance under drought stress, which leads to more consistent yield over time. For instance, ref. [47] found that no-till treatments reduced cumulative N<sub>2</sub>O emissions in soybean rotation by 20% to 35.7% compared to conventional tillage, varying with the type of fertilizer type used in South Korea. Similarly, ref. [48] conducted a study in Brazil and found that no-till reduced N<sub>2</sub>O emissions in a soybean–wheat rotation system. In the previous context, the objective of this investigation was to evaluate how conventional tillage and no-tillage can potentially influence N<sub>2</sub>O emissions in soybean rotation in a semi-arid region of the central Free State of South Africa.

## 2. Materials and Methods

### 2.1. Site Description

This research was carried out during the soybean cropping season of October 2022 to December 2023 at the Kenilworth Research Station, located in Bloemfontein, Free State, South Africa, latitude 29.13184 south, longitude 26.24107 east, altitude 1395 m. This is a semi-arid region of South Africa. Bloemfontein receives most of its rainfall in summer (January and February), with an average rainfall of 469 mm per year. Summers are warm; winters are short, cold, and dry. The driest weather is in July and the wettest is in February. The mean maximum temperatures are 19 and 27 °C and the mean minimum temperatures are 1 and 12 °C for winter and summer, respectively. In 2020 to 2023 the Kenilworth Research Station had an average temperature of 17.1 °C, humidity of 47% and precipitation of 33.34 mm. The soil used in this experiment had a pH of 6.57, and an organic matter content of 1.75%, calcium, magnesium, and potassium of 6.91, 1.22 and 1.04 cmol kg<sup>-1</sup>, respectively, in the 0.30 cm deep zone in 2020.

### 2.2. Experimental Design

The experimental design incorporated a randomized complete block design (RCBD). Each block (3 blocks in total) was 192 m (width) × 55 m (length) (10 560 m<sup>2</sup>), and each plot was 8 m (width) × 55 m (length) (440 m<sup>2</sup>). There were three blocks, with two soybean plots in each block, one plot representing each treatment in each block; in total there were six plots. The plant density for soybean was 270,000 plants per ha. Inter-row spacing was 0.75 m and intra-row spacing was 0.04 m.

### 2.3. Treatments

The treatments were planted in soybean rotation systems under no-till (NT) and conventional tillage (CT) [49]. In CT plots, the soil was ploughed with a moldboard prior to planting and, for the NT plot, no ploughing took place, and the crop residues were left on the soil surface. The measurements during the experiment were based on the 3rd year of a 5-year rotation experiment. The crops grown before 2023 were soybean, followed by radish, which was grown as cover crop. The planting of soybean was carried out on 10 December 2022 and the base fertilizer NPK 1:1:1 (30) was applied at a rate of 200 kg ha<sup>-1</sup> at planting. A selective herbicide (Sonalan) was applied to control weeds on 4 February and hand weeding was also carried out. The crop was harvested on 26 June 2023 and yields were calculated as mass of total dry seed weight/unit area, and total biomass/unit area. The total dry seed weight and biomass were then converted to t/ha. The sum of the total dry seed weight and biomass made up the total crop dry weight. Thus, the treatments are as follows:

1. Conventional Tillage: Soybean-Cover crop-Soybean (CT: SB-CC-SB) In these plots, there was crop rotation; soybean was followed by cover crop, then soybean again under conventional tillage; each crop was grown for 12 months, and the soil was cleared before the next crop was planted. The plant density was 270,000 plants per ha.
2. No-tillage: Soybean-Cover crop-Soybean (NT: SB-CC-SB). In these plots, crop rotation took place; soybean was followed by cover crop, then soybean again under no-tillage; each crop was grown for 12 months and the soil was cleared before the next crop was planted. The plant density was 270,000 plants per ha.

### 2.4. Gas Sampling and Flux Measurements

The gas sampling procedure was conducted for a continuous 12-month period between December 2022 and December 2023 [50], making up to 30 measurements. Gas sampling was carried out weekly from December to February, and thereafter twice a month as

the crop matured and once a month in the off-season. Gas sampling was performed between 10:00 a.m. and 12:00 p.m. Four polyvinyl chloride (PVC) static white chambers (40 cm length  $\times$  40 cm width  $\times$  25 cm height) were permanently installed (1 m apart) in each plot to a depth of 5-cm using a steel base before sampling and points were permanently marked using a global positioning system (GPS), so that they could be moved into the same spot after agronomic management practice, i.e., weeding. The static chambers were permanently installed for the duration of the trial; they were only uninstalled at the end of the season. During each gas sampling event, ambient samples ( $T_0$  air from the atmosphere) were taken away from the plots, buildings or cars; five at the beginning and five at the end of each sampling day, using 60 mL of gastight polypropylene syringes. During sampling, four random boxes were chosen for linearity check at 20-min intervals (five different times,  $T_0$ ,  $T_{20}$ ,  $T_{40}$ , and  $T_{60}$ ) to confirm linear gas accumulation with closure time. The remainder of the boxes were terminally sampled at 40 min of closure ( $T_{40}$ ) by drawing air using 60 mL of gastight polypropylene syringes and storing the air in 40 mL vials. Gas samples were then analysed in the laboratory using a gas chromatograph [51] model 107 [52], and the daily fluxes were calculated using the equation below:

$$F = \rho \frac{V}{A} \frac{(C_t - C_0)}{t} \left( \frac{273.15}{T} \right)$$

where  $F$  is the flux of  $N_2O$  ( $g\ N_2O-N\ ha^{-1}d^{-1}$ );  $\rho$  is the gas ( $1.26\ g\ cm^{-3}\ N_2O-N$ ) under STP (273K and 101,325 Pa);  $V$  and  $A$  are the volume ( $0.125\ m^3$ ) and area ( $0.25\ m^2$ ) of  $C_{60}$  and  $C_0$  ( $\mu m^3\ m^{-3}$ ) are the  $N_2O$  concentrations 60 min after chamber closure ( $T_{60}$ ) and the ambient sample ( $T_0$ ), respectively;  $t$  is the time of chamber closure in hours ( $T_{60}$ ); and  $T$  is the air temperature, (K) at the time of sampling. Nitrous oxide fluxes were transformed into daily fluxes ( $kg\ N_2O\ ha^{-1}\ day^{-1}$ ) for the calculation of cumulative  $N_2O$  emissions. The cumulative emissions were determined as the total amount  $N_2O$  released over the growing season. Nitrous oxide fluxes for non-sampling dates were interpolated between two sampling dates, and cumulative emissions calculated using the trapezoidal method [53,54].

## 2.5. Soil Sampling and Analysis

Soil samples were collected (0–10 cm) from each plot in each block at the beginning of the trial in 2020, to determine the soil status before the beginning of the study (baseline values). These soil samples were used to determine pH, texture, bulk density (BD), organic matter (OM), total phosphorus (P), potassium (K), sodium (Na), magnesium (Mg) and calcium (Ca). Soil pH was determined via the water method, using the procedure of the Agri Laboratory Association of Southern Africa Handbook [55]. Soil OM was determined using the loss-on-ignition method, where samples were oven dried in a furnace overnight [55] and the % OM was calculated using the following equation:

$$\% OM = \frac{(W_0 + W_1) - (W_0 + W_2)}{(W_2 + W_1 - W_0)} \times 100$$

where OM is organic matter;  $W_0$  is the weight of the crucible (g);  $W_1$  is the dry soil (g); and  $W_2$  is the dry soil after ignition (g).

Mineral N was determined by the potassium chloride extraction method [56], where 40 g of fresh soil sample were weighed and 80 mL of 2M KCL (1:2 soil/extractant ratio) were added for mineral N extraction and analysed colorimetrically [56]. Total N and total organic carbon (TOC) contents of soil were determined using the LECO method [57] and total P and cation exchange capacity using the Bray 1 method [57]. During each gas sampling event, soil samples were collected (0–10 cm) adjacent to each of the 24 chambers to determine

gravimetric moisture. To measure the gravimetric water content, the soil samples were oven dried at 105 °C for 24 h or until a constant weight was obtained. The %WFPS was calculated using the gravimetric moisture content method, as described by [55]. This was calculated using the following equation:

$$\%WFPS = \frac{SWC}{1 - BD/PD} * 100$$

where WFPS are water filled pores (%); SWC is the volumetric water content (%); BD is the bulk density ( $\text{g cm}^{-3}$ ); and PD is the particle density ( $2.65 \text{ g cm}^{-3}$ ).

For soil porosity

$$\text{Soil Porosity} = 1 - \frac{BD}{PD}$$

where PD is the particle density of the soil, assumed to be  $2.65 \text{ g cm}^{-3}$ . Volumetric water content was calculated by multiplying the bulk density (BD) and the gravimetric water content. Soil BD was determined at the beginning of the experiment by collecting an undisturbed soil sample by inserting metal rings into the soil with a hammer and determining the weight of the collected soil samples before and after oven drying at 105 °C for 48 h [58,59]. The bulk density was determined by the following equation:

$$BD = \frac{M}{V}$$

where BD is the bulk density; M is the mass of the soil (g); and V is the volume of the soil ( $\text{cm}^3$ ).

The soil temperature was also measured on each gas sampling event using alcohol thermometers placed vertically in the soil at approximately 10 cm depth. Minimum and maximum temperature and precipitation were measured daily at the weather station located on the farm.

## 2.6. N Loss

The N loss (cumulative  $\text{N}_2\text{O}$ ) for each treatment was calculated using the following equation:

$$\text{N LOSS} = \frac{E_{\text{N}_2\text{O fertilized}}}{R_{\text{N}}} \times 100$$

[58]

Here,  $E_{\text{N}_2\text{O fertilized}}$  represents the measured emissions after application of fertilizer, and  $R_{\text{N}}$  is the N fertilizer rate ( $\text{kg N ha}^{-1}$ ).

## 2.7. Estimated Emissions per Product

The estimated emissions per product for each treatment were calculated using the following equation:

$$\text{Estimated Emissions Product} = \frac{\text{Cumulative Emissions}}{\text{Total Yield}}$$

## 2.8. Data Analysis Trials

Linear models were used to find the differences in  $\text{N}_2\text{O}$  influenced by the no-till and conventional tillage system. If the interactions were significant at  $p < 0.05$ , the Tukey multiple comparison test was used to test differences between treatments. If the interaction was not significant, the analysis was rerun without the interaction term. Response data were transformed before analysis when required to meet the assumptions of the model. Data were sorted in Microsoft Excel; mean separation was performed using the GLM procedure

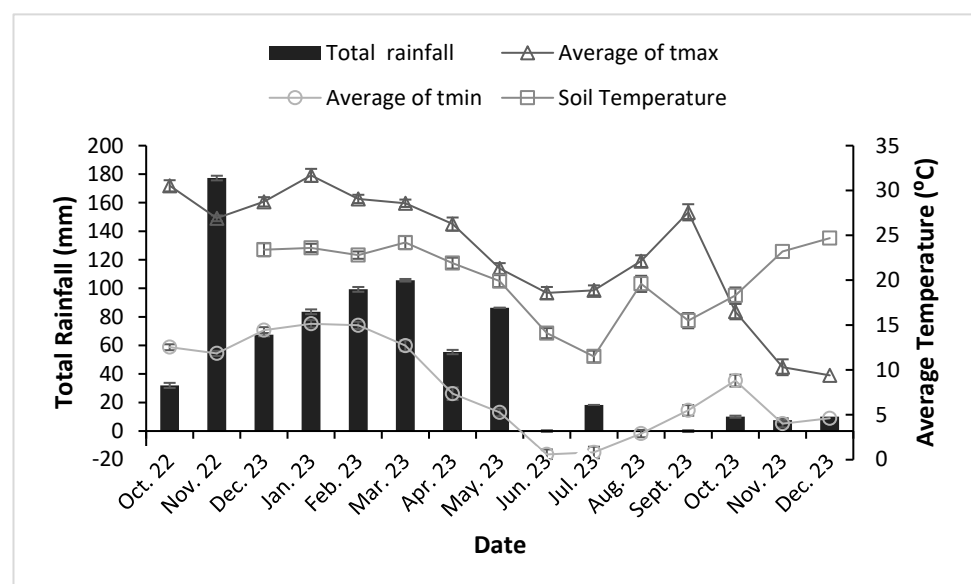
in Statistical Analysis Software (9.0) (SAS). Correlation coefficients were obtained using Microsoft Excel.

### 3. Results

#### 3.1. Meteorological Data

##### 3.1.1. Rainfall Patterns and Atmospheric Temperature During the Experimental Period

The total rainfall for the entire experimental period (October 2022 to December 2023) was 753.22 mm and the highest rainfall event of 177.3 mm was recorded in November 2022; the second highest rainfall events of 105.7 and 99.3 mm was received in March and February 2023, respectively (Figure 1). The highest air temperature was 31.7 °C recorded in January 2023, and the second highest temperature was 30.5 and 29.1 °C recorded in November and March 2023, respectively. The highest soil temperature was 24.7 °C recorded in December 2023, followed by 24.2 °C recorded in March 2023.



**Figure 1.** Monthly rainfall, soil temperature, minimum and maximum temperatures for the experiment.

##### 3.1.2. Soil Variables

##### General Soil Characteristics Before and After the Experiment

Different soil parameters were recorded at the beginning and end of the trial in December 2023 (Table 1). Sand% ranged from  $88 \pm 4.67$  and  $93.67 \pm 4.25\%$ , with the highest of  $93.67 \pm 4.25\%$  in NT at the beginning of the trial, which is significantly different from CT (LSD = 1.95). The highest silt percentage of  $6.92 \pm 2.88$  was recorded in CT after the trial, which was significant (LSD = 1.64) from NT. The highest clay % of  $5.17 \pm 1.59$  was recorded in NT after the test; however, it was not significantly (LSD = 1.35) different from CT. The soil pH ranged from  $5.91 \pm 0.39$  and  $6.3 \pm 0.5$ , with the highest pH of  $6.3 \pm 0.5$  in NT before the trial, which was not significantly (LSD = 0.29) different from CT. Soil bulk density ranged from  $1.4 \pm 0.17$  and  $1.52 \pm 0.12$ , with the highest bulk density ( $1.52 \pm 0.12$ ) recorded in NT before the trial, which was significantly (LSD = 0.09) different from CT. The total C ranged from  $1.44 \pm 0.26$  and  $2.07 \pm 0.77$  g C kg<sup>-1</sup> of dry soil. The highest total C of  $2.07 \pm 0.77$  was in CT after the trial, which was, however, not significant (LSD = 0.4) for NT. The total N ranged from  $27.23 \pm 31.62$  and  $342.59 \pm 84.37$  mg N kg<sup>-1</sup> dry soil, with the highest total N of  $342.59 \pm 84.37$  mg N kg<sup>-1</sup> dry soil in CT before the trial, which was not significantly (LSD = 49.45) different from NT. Organic matter % (OM) ranged from  $3.04 \pm 1.29$  and  $13.88 \pm 4.06\%$ . the highest %OM ( $13.88 \pm 4.06$ ) was recorded CT before



the trial, and there was no significant difference between the treatments. The %WFPS ranged from  $3.05 \pm 1.88$  and  $22.91 \pm 4.74$ , and the highest %WFPS of  $22.91 \pm 4.74\%$  was in NT before the trial, which was, however, not significant ( $LSD = 2.91$ ) in CT. Soil porosity ranged from  $43 \pm 4.4$  and  $46 \pm 6.5\%$ , with the highest 46% in CT, which was not significant ( $LSD = 3.3$ ) from NT. Soil  $NH_4^+$  ranged from  $1.24 \pm 2.19$  and  $5.21 \pm 3.46$  mg N kg<sup>-1</sup> dry soil. The highest  $NH_4$  of  $5.21 \pm 3.46$  was in NT after the trial, which was, however, not significant ( $LSD = 2.32$ ) from CT. Soil  $NO_3$  ranged from  $4.41 \pm 2.65$  and  $47.3 \pm 60.06$  mg N kg<sup>-1</sup> dry soil. The highest  $NO_3$  of  $47.3 \pm 60.06$  mg N kg<sup>-1</sup> dry soil was recorded in CT, which was not significantly ( $LSD = 19.54$ ) different from NT.  $NO_2$  ranged from  $0.015 \pm 0.012$  and  $0.038 \pm 0.057$  mg N kg<sup>-1</sup> dry soil, with the highest of  $0.038 \pm 0.057$  mg N kg<sup>-1</sup> dry soil recorded in NT, which was not significantly ( $LSD = 0.19$ ) different from CT. The soil C/N ratio ranged from  $4.77 \pm 1.17$  and  $104.89 \pm 73.93$ , with the highest C/N ratio of  $104.89 \pm 73.93$  recorded in NT after the trial, which was significantly ( $LSD = 32.66$ ) different from CT. Hydraulic conductivity ranged from  $0.16^{-2} \pm 0.37^{-2}$  and  $0.33^{-2} \pm 0.39^{-3}$  cm/s, with the highest of  $0.33^{-2} \pm 0.39^{-3}$  cm/s recorded in NT, which was, however, not significantly ( $LSD = 0.18^{-2}$ ) different from CT.

**Table 1.** Summary of soil parameters (mean  $\pm$  standard error) in soybean grown under conventional tillage and no-tillage from the commencement of the trial in November 2022 and after the trial in December 2023.

Parameter	Soybea-CT	Soybean-NT	LSD
<b>Before 06/12/2022</b>			
Soil Texture			
Sand (%)	$91.00 \pm 4.67^b$	$93.67 \pm 4.25^a$	1.95
Silt (%)	$6.00 \pm 4.82^a$	$3.00 \pm 3.36^b$	1.91
Clay (%)	$2.92 \pm 2.07^c$	$3.33 \pm 1.97^b$	0.8
Soil pH	$6.05 \pm 0.57^a$	$6.30 \pm 0.50^a$	0.29
Bulk density (g/cm <sup>3</sup> )	$1.44 \pm 0.17^b$	$1.52 \pm 0.12^a$	0.09
Total C (g C/kg dry soil)	$1.65 \pm 0.41^a$	$1.44 \pm 0.26^a$	0.22
Total N (mg N/kg dry soil)	$342.59 \pm 84.37^a$	$333.08 \pm 69.78^a$	49.45
Organic matter (% w/w)	$13.88 \pm 4.06^a$	$11.96 \pm 1.99^a$	1.78
WFPS (%)	$21.63 \pm 4.48^a$	$22.91 \pm 4.74^a$	2.91
$NH_4^+$ (mg N/kg dry soil)	$1.24 \pm 2.19^b$	$3.05 \pm 3.74^a$	1.58
$NO_3$ (mg N/kg dry soil)	$47.3 \pm 60.06^a$	$33.87 \pm 21.24^a$	19.54
$NO_2$ (mg N/kg dry soil)	$0.018 \pm 0.061^a$	$0.038 \pm 0.086^a$	0.052
C/N Ratio	$4.77 \pm 1.17^a$	$4.86 \pm 1.64^c$	1.38
<b>After 06/12/2023</b>			
Soil Texture			
Sand (%)	$88.00 \pm 4.67^a$	$89.00 \pm 2.89^a$	0.24
Silt (%)	$6.92 \pm 2.88^a$	$5.83 \pm 2.41^a$	1.64
Clay (%)	$5.08 \pm 3.00^a$	$5.17 \pm 1.59^a$	1.35
Soil pH	$6.06 \pm 0.25^a$	$5.91 \pm 0.39^b$	0.16
Bulk density (g/cm <sup>3</sup> )	$1.40 \pm 0.17^c$	$1.50 \pm 0.13^a$	0.087
Total C (g C/kg dry soil)	$2.07 \pm 0.77^a$	$1.97 \pm 0.81^a$	0.4
Total N (mg N/kg dry soil)	$50.03 \pm 49.61^a$	$27.23 \pm 31.62^a$	33.59
Organic matter (% w/w)	$3.12 \pm 1.19^a$	$3.04 \pm 1.29^a$	0.69

Table 1. Cont.

Parameter	Soybean-CT	Soybean-NT	LSD
WFPS (%)	$3.05 \pm 1.88^a$	$4.98 \pm 4.21^a$	1.86
Soil porosity (%)	$0.46 \pm 0.065^a$	$0.43 \pm 0.044^{ab}$	0.033
NH <sub>4</sub> <sup>+</sup> (mg N/kg dry soil)	$5.05 \pm 4.65^a$	$5.21 \pm 3.46^a$	2.32
NO <sub>3</sub> (mg N/kg dry soil)	$4.41 \pm 2.65^b$	$6.01 \pm 3.97^a$	2.39
NO <sub>2</sub> (mg N/kg dry soil)	$0.015 \pm 0.012^a$	$0.038 \pm 0.057^a$	0.19
Hydraulic conductivity (cm/s)	$0.16^{-2} \pm 0.37^{-3}a$	$0.33^{-2} \pm 0.39^{-3}a$	$0.18^{-2}$
C/N Ratio	$49.96 \pm 31.3^a$	$104.87 \pm 73.93^b$	32.66

WFPS (Water Filled Pore Spaces). Means separated by different lower-case letters (a, b, c) in each column are significantly different at  $p \leq 0.05$ .

### Soil Measurements

The average WFPS for the trial was 22.2%. The %WFPS ranged from 3 to 43%; the highest of  $43 \pm 4.73$  was recorded in December 2022 in NT and the lowest of  $3.05 \pm 1.88$  was recorded in CT in December 2023 (Figure 2), which was not significantly (LSD = 2.91) different from CT. Soil NH<sub>4</sub> ranged from  $1.24 \pm 2.19$  to  $5.21 \pm 3.46$  mg N kg<sup>-1</sup> dry soil in (Figure 3). The highest NH<sub>4</sub><sup>+</sup> was  $5.21 \pm 3.46$  in NT, which was however, not significant (LSD = 2.32) in CT. The NO<sub>3</sub><sup>-</sup>-N of the soil ranged from 1.59 to 104.56 in 2023 (Figure 3). The highest NO<sub>3</sub>-N was 104.56 recorded in CT, which was significantly (LSD = 2.39) different from NT. NO<sub>2</sub>-N ranged from  $0.015 \pm 0.012$  to  $0.038 \pm 0.057$  mg N kg<sup>-1</sup> dry soil; the highest NO<sub>2</sub>-N was  $0.038 \pm 0.057$  mg N kg<sup>-1</sup> dry soil recorded in NT, which was not significantly different (LSD = 0.19) from CT (Table 1).

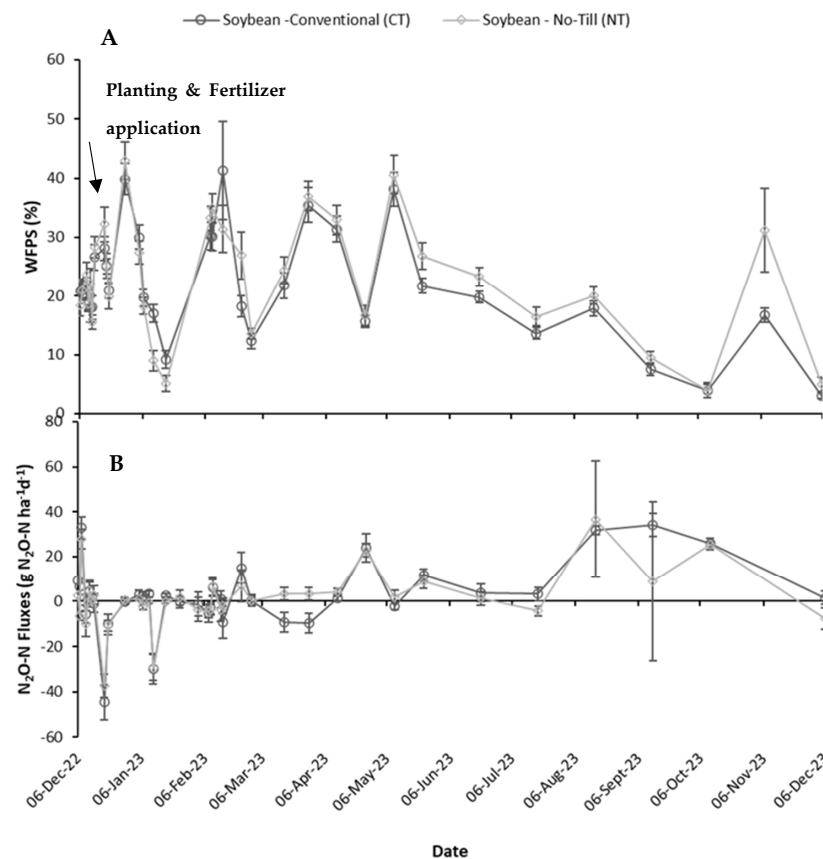
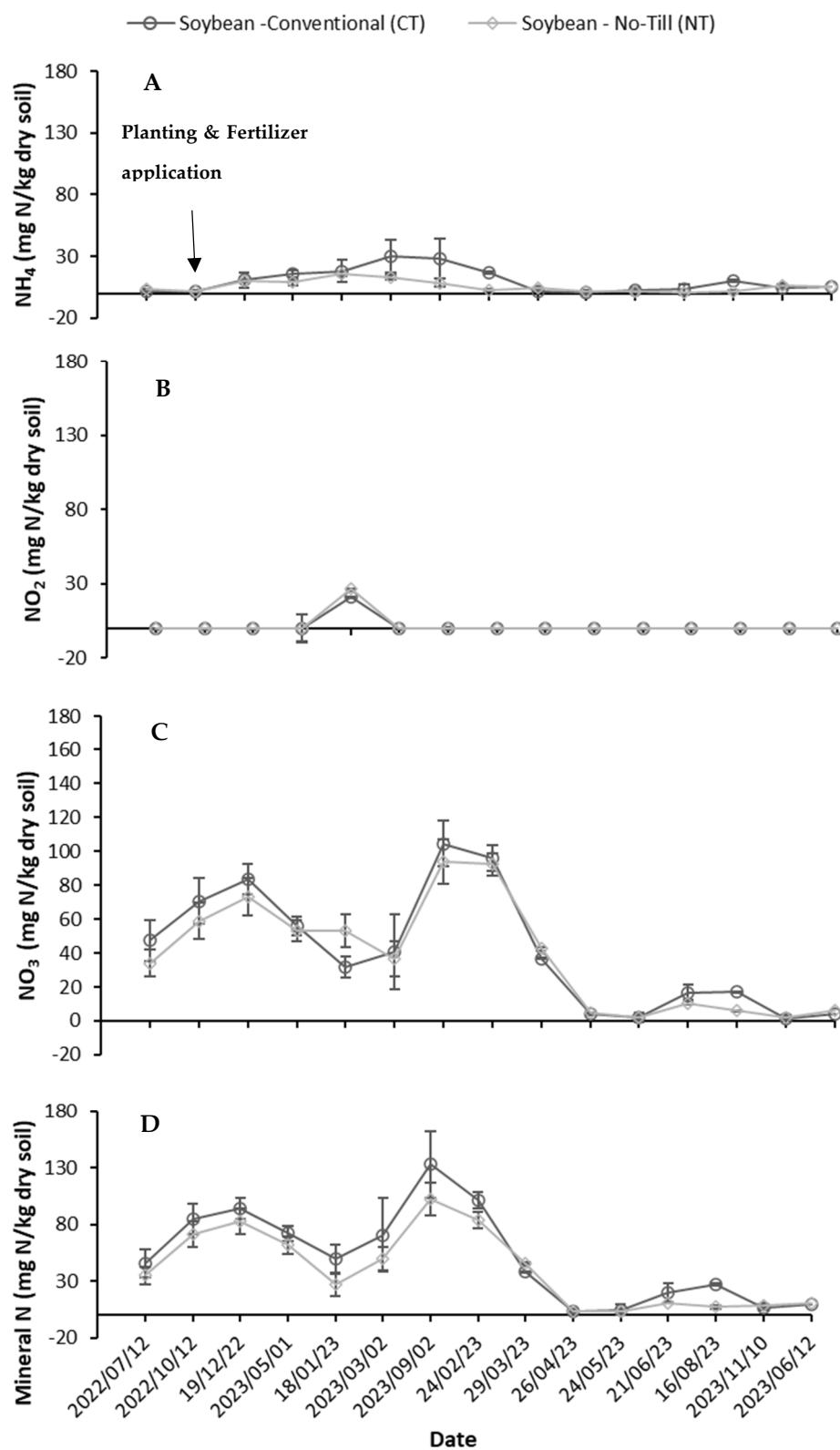


Figure 2. Dynamics of N<sub>2</sub>O and % WFPS in the different treatments during the experimental period.





**Figure 3.** Mineral N dynamics in no-till and conventional tillage in soybean during the experimental period.

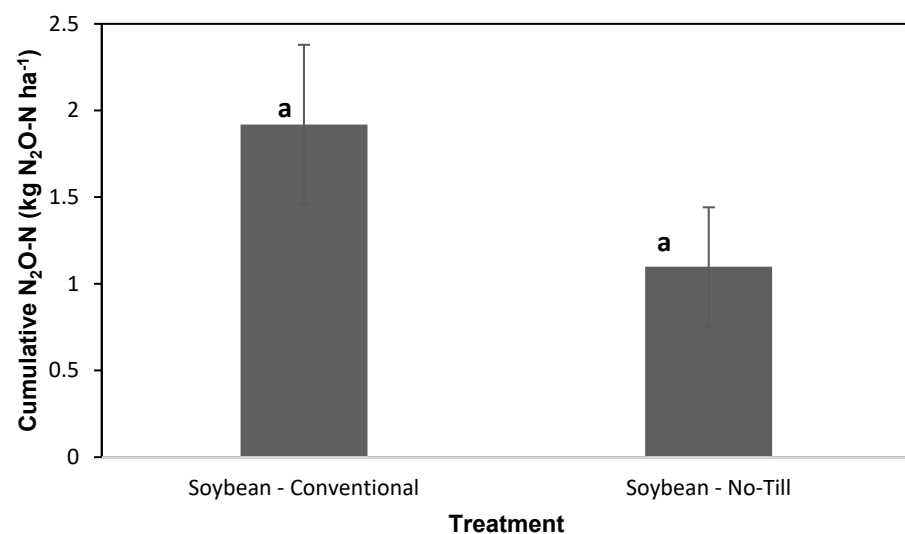
### 3.1.3. Nitrous Oxide Emissions

The N<sub>2</sub>O fluxes ranged from  $-44.6$  to  $36.6$  g of N<sub>2</sub>O-N ha<sup>-1</sup> d<sup>-1</sup>. The lowest flux was recorded on 19 December 2022 (Figure 2). There was a peak in February after the application of top-dressing fertilizers on 3 February 2023, and after an increase in %WFPS.

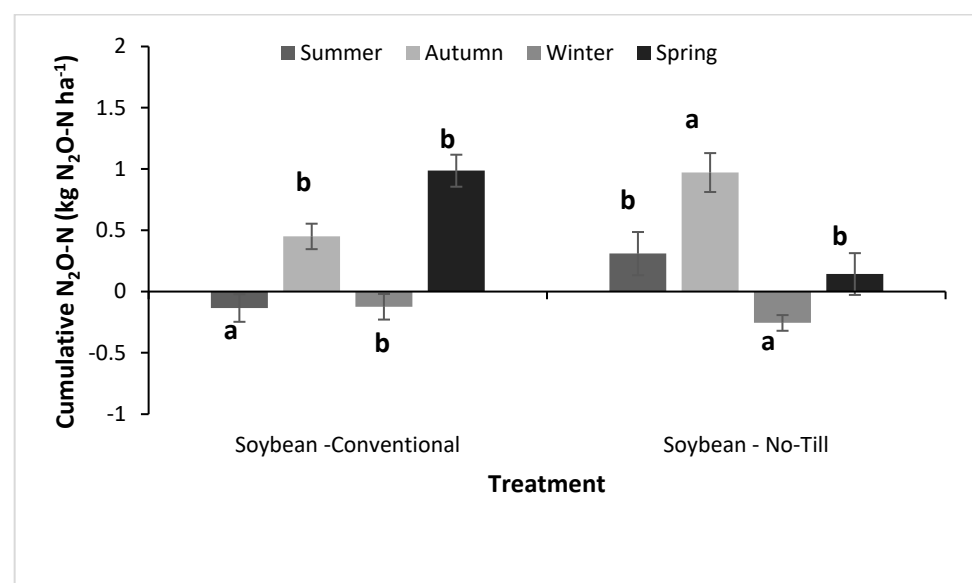
The highest peak was observed in August–September from both treatments after harvest, the fluxes were significantly ( $LSD = 0.3$ ) higher from CT to NT in the plots sampled. Fluxes decreased thereafter; fluxes decreased in NT compared to CT. Low  $N_2O$  emissions were at the same time with low %WFPS in December 2023 (Figure 2).

### 3.1.4. Cumulative $N_2O$ Emissions and N Lost as $N_2O$

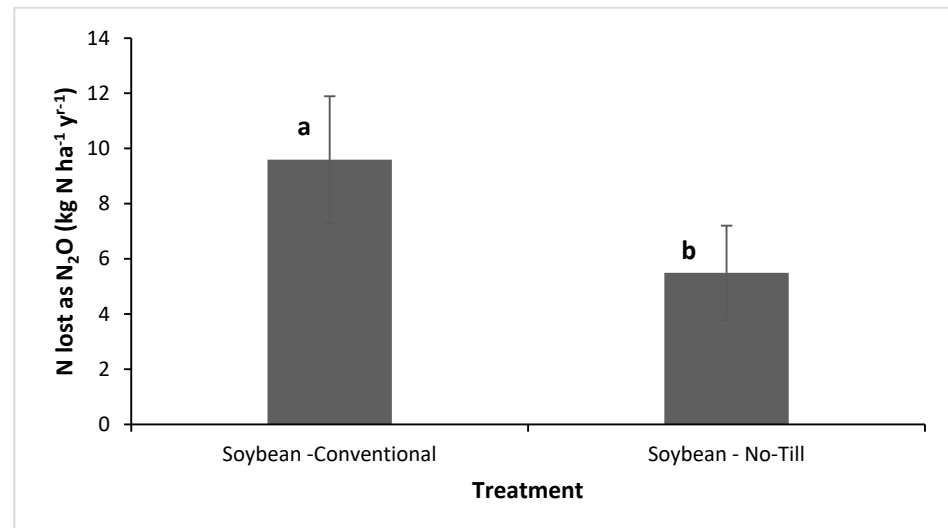
The highest cumulative emissions ( $1.9 \text{ kg of } N_2O\text{-N ha}^{-1}$ ) were recorded in CT plots (Figure 4). The highest cumulative  $N_2O$  emissions for the different seasons show that CT has the highest  $N_2O$  emissions in spring  $0.99 \pm 4.45 \text{ kg } N_2O\text{-N ha}^{-1}$ , which was, however, not significantly ( $LSD = 0.29$ ) different from NT (Figure 5). The lowest  $N_2O\text{-N}$  emissions were recorded during the winter season ( $-0.26 \pm 0.22 \text{ kg of } N_2O\text{-N ha}^{-1}$ ) in NT, which was significantly different ( $LSD = 0.19$ ) from CT (Figure 5). The N lost as  $N_2O$  ranged from 0 to 9.6% (Figure 6). The highest N lost as  $N_2O$  of 9.68% was recorded in CT; there was a significant difference between CT and NT ( $LSD = 1.6$ ).



**Figure 4.** Cumulative  $N_2O$  in the different treatments during the experimental period. Means separated by different lower-case letters (a, b, c) in each column are significantly different at  $p \leq 0.05$ .



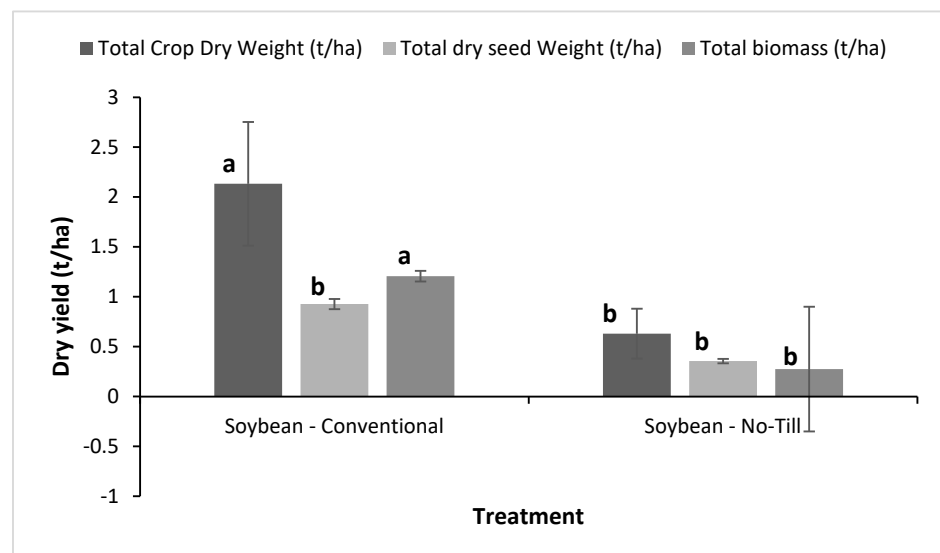
**Figure 5.** Cumulative  $N_2O$  in the different treatments during the different seasons of the experimental period. Means separated by different lower-case letters (a, b, c) are significantly different at  $p \leq 0.05$ .



**Figure 6.** N lost as N<sub>2</sub>O in the different treatments during the experimental period. Means separated by different lower-case letters (a, b, c) are significantly different at  $p \leq 0.05$ .

### 3.1.5. NT and CT Practices on Yields

The total yield was higher in CT compared to NT in the rotation of the soybeans. The highest total dry seed yield was 0.93 t/ha recorded in CT (Figure 7). The highest total crop dry weight and total biomass were recorded in CT. These were 2.13 and 1.21 t/ha, respectively. There was a significant difference between NT and CT (LSD = 0.23). The estimated emissions per product were higher in NT (1.67 kgNha<sup>-1</sup> dry matter) and lower in CT (0.95 kgNha<sup>-1</sup> dry matter) (Table 2) for the duration of the trial.



**Figure 7.** Dry grain, biomass and total crop yield in the different treatments during the experimental period. Means separated by different lower-case letters (a, b, c) are significantly different at  $p \leq 0.05$ .

**Table 2.** Estimated emissions per product in soybean grown under conventional tillage and no-tillage for the duration of the trial.

Treatment	N <sub>2</sub> O kg N ha <sup>-1</sup>	Yield t ha <sup>-1</sup>	Kg N ha <sup>-1</sup> Dry Matter
Soybean-CT	2	2.1	0.95
Soybean-NT	1	0.6	1.67

## 4. Discussion

### 4.1. The Role of Soil Variables in Daily $N_2O$ Fluxes

The highest  $N_2O$  fluxes in this trial were recorded in December 2022–March 2023 (Figure 2). These high fluxes were recorded at the same time with planting and fertilizer application and episodes of precipitation on December 2022 and February 2023. These peaks followed rainfall events occurring in February–March. During this period of the year the soil temperature was high (24.2 °C) (Figure 1). The cause of this peak may have been the high soil temperature and moisture content significantly influencing  $N_2O$  production by increasing microbial activity, enhancing enzyme activity and accelerating organic matter decomposition [12,44]. This is in line with other studies such as [59] who found that %WFPS and temperature are the main drivers of  $N_2O$  fluxes; also [60–62] who reported that  $N_2O$  fluxes increased exponentially with soil temperature and, who reported that  $N_2O$  fluxes increase by five to nine times with each 10 °C temperature increase and microbial nitrification and denitrification increase three-fold with a 10 °C increase. During denitrification,  $NO_3^-$  is reduced to  $NO$  to  $N_2O$ . The high temperature (25–35 °C) encourages microbial enzyme activity. This accelerates denitrification (anaerobic) and nitrification (aerobic).

The high temperature and %WFPS may have stimulated high microbial activity, which in turn increased denitrification when the soil became saturated and oxygen levels decreased [63,64]. High  $N_2O$  fluxes also responded to the application of inorganic fertilizers in the soil and the amount of crop residues from crop rotation present in the soil, as this may have contributed soluble C to denitrifiers when mineralized [25,65–67].

Ref. [68] found that  $N_2O$  emissions were high at 65% WFPS and in water-saturated soils, where  $N_2O$  is mainly formed by denitrification of nitrate. The highest %WFPS in our study was approximately 44%, indicating that  $N_2O$  emissions in our study may have mainly been the result of nitrification, but we did not quantify this in the current study. This is in agreement with [69,70], who reported that most of the  $N_2O$  emissions are from nitrification when the WFPS is below 60%, while an increased conversion from  $N_2O$  to  $N_2$  occurs at increased soil water content.

The large peak in  $N_2O$  fluxes immediately after N fertilizer application in December 2022 was the result of that N application (Figure 2), similar to findings by [71], who reported a 75% increase in mean  $N_2O$  in soybean after N fertilizer application. There was a positive correlation ( $p = 0.05$ ) between  $N_2O$  emissions and N in the soil (Figure 8). These results are in line with previous authors [72–74], who found that N application increases N availability in the soil and promotes  $N_2O$  emissions compared to unfertilized soil. Refs. [75–77] found that high  $N-NH_4^+$  in the soil leads to greater nitrification. Refs. [78,79] highlighted that a high number of  $N_2O$  emissions may have been because the  $NO_2^-$  formed during the nitrification process can be used as electron receptor if  $O_2$  is limited, and denitrification can occur after nitrification, when soil conditions are conducive.

### 4.2. NT and CT Practices on Cumulative $N_2O$ Emissions and N Lost as $N_2O$

Cumulative  $N_2O$  emissions were higher in CT compared to NT, and there was a peak during planting (Figure 4). NT reduced emissions, in agreement with the results of [64], who found that soybean rotation had lower  $N_2O$  emissions compared to plots with monoculture soybean under NT and CT. The lower rate of  $N_2O$  emissions from soybean crops supported the conclusion of [80], who found that N is released from root exudates during the growing season, and decomposition of previous crop residues influence  $N_2O$  emissions much more than the biological N fixation process. In August immediately after harvesting, the emissions were higher in NT compared to CT, in line with an increase in % WFPS, bulk density and high C in the soil from crop residues. High WFPS in NT is due to

residues covering the soil surface, thus reducing water evaporation from the soil. Since the NT soils hold more water, this led to heavier and more compacted soils, contributing to higher bulk density in NT. There was a positive correlation ( $p = 0.21$ ) between  $N_2O$  emissions and soil C (Figure 8). These results agree with [81–83], who found that increased soil moisture (WFPS) in no-till treatments increased the amount of denitrifying activity and, subsequently,  $N_2O$  production. In September,  $N_2O$  emissions were higher in CT compared to NT; this was after harvest, when the soil had been disturbed as the plants were pulled from the soil and some residues were incorporated into the soil. This increased the organic matter in the soil, leading to a higher emission of  $N_2O$ . There was a strong correlation ( $p = 0.43$ ) between  $N_2O$  emission and soil organic matter (Figure 8). These results are in line with [6,84,85], who found that incorporating crop residues into the soil increased  $N_2O$  emissions, mainly in CT. The decrease in  $N_2O$  emissions with NT treatment could also be due to the fact that soybean roots have the potential to denitrify the last step of the denitrification process ( $N_2O$  to  $N_2$ ) [86], and this also has the potential to reduce  $N_2O$  derived from nodules, fertilizer N, and soil organic matter [87].

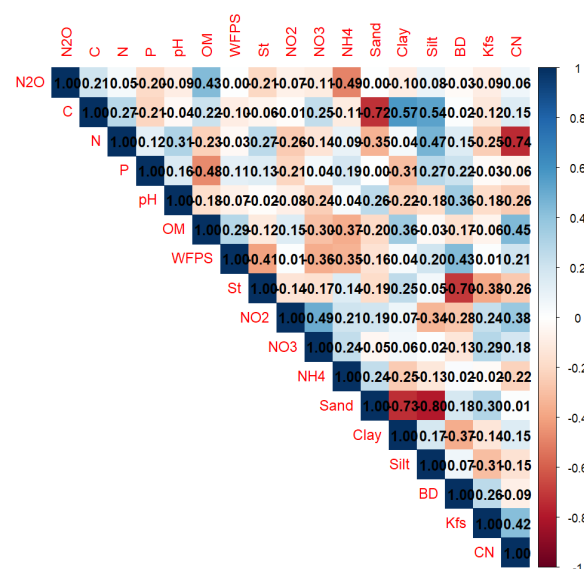


Figure 8. Correlations Matrix between cumulative  $N_2O$  and soil variables.

Refs. [88,89] reported reduced  $N_2O$  emissions in NT compared to CT. This is possibly due to sequential nitrification and denitrification by nitrifiers, such as *Nitrosomonas* sp. and *Nitrobacter* sp., and denitrifiers in optimal soil temperature and moisture conditions. In our trial, in CT most of the emissions occurred in spring, followed by autumn; this is when the temperature gradually increased (Figure 5). In NT, the highest emissions occurred in autumn, when the average soil temperature was 22 °C (Figure 1). This is in line with Signor [11], who found that  $N_2O$  production during nitrification increases when the soil temperature is between 5 and 40 °C. Peigne et al., 2007 [90] reported that N availability was lower in NT than in CT, which is consistent with our study (Table 1). Refs. [91,92] found that the lower availability of N in NT was due to a low N mineralisation rate in NT and immobilisation accompanied by slow soil organic matter decomposition [93].

The loss of N as  $N_2O$  was the highest in CT, rather than in NT (Figure 6). These results are in line with [94] who found that most of the simulated loss of nitrogen are from drains flow and net mineralisation, leaching. Ref. [95] found that N loss to drain flow was highest mainly due to high precipitation events during or after fertilizer application.

#### 4.3. NT and CT Practices on Yields

The total dry crop yields under CT were found to be higher in CT than in NT. Total dry crop yield in CT was 61% higher than in NT (Figure 6). The higher yields in CT may be related to the high N, C, porosity and organic matter recorded in CT, as these soil properties improve nutrients available for plant uptake, thus increasing yields (Table 1). Crop residues are mixed with soil under CT, thus reducing soil porosity and bulk density. CT mix crop residues into the soil this leads to high organic matter, C and N temporarily, especially in the topsoil. It also creates a uniform mixture of organic and mineral soil layers. This blending effect can lead to high average organic matter content. Aeration from the tillage accelerates microbial decomposition. Initially, this can release nutrients (including nitrate and mineral N) and increase microbial biomass, temporarily raising measurable N and increase yields. The NT plots had a higher % of sand, high bulk density, and lower total C and N. These results are in line with [96], who found that soybean yields were lower on sandy soils compared to loamy soils. Ref. [97] also found low yields in soybeans under NT rather than in CT, due to the soil having lower water holding capacity, N insufficiency due to soil leaching, and the reduced root length under NT. Ref. [51] found significantly low yields in the dry season. The possibility of low yield in NT is possibly due to the fact that soybean NT did not respond to the extra soil moisture available at the lower depth because plant root depth may be delayed under NT [98]. The emissions per product was higher in NT (1.67 kg N ha<sup>-1</sup> dry matter) compared to CT (0.95 kg N ha<sup>-1</sup> dry matter), as presented in Table 2. These results were almost similar to those of [99], who found emissions per product of 1 kg N ha<sup>-1</sup> dry matter, both under CT and NT.

## 5. Conclusions

This study provided comprehensive and quantitative results of NT and CT effects on N<sub>2</sub>O emissions in a semi-arid region. In general, NT reduced N<sub>2</sub>O emissions in soybean rotation. The reduction in NT treatment was more prominent during the dry season. No-tillage is worth practicing. These findings agree with those in the published literature. The total dry matter yield was lower under NT; however, the dry matter yield may be improved with long term duration of NT.

**Author Contributions:** N.O.D. was involved in conceptualizing, methodology, formal analysis, data curation, writing—original draft, writing—review and editing. L.B. was involved in planning and supervising the work. L.M.C. was involved in planning and supervising the work. She commented on the manuscript. A.L.-L. was involved in processing the gas data, she also commented on the manuscript. J.C.D. was involved in planning and supervising the work. He contributed to the design and implementation of the research, to the analysis of the results and to the writing of the manuscript. All authors have read and agreed to the published version of the manuscript.

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**Data Availability Statement:** Data are contained within the article.



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## References

1. Ramzan, S.; Rasool, T.; Bhat, R.A.; Ahmad, P.; Ashraf, I.; Rashid, N.; ul Shafiq, M.; Mir, I.A. Agricultural soils a trigger to nitrous oxide: A persuasive greenhouse gas and its management. *Environ. Monit. Assess.* **2020**, *192*, 436. [\[CrossRef\]](#)
2. Montzka, S.A.; Dlugokencky, E.J.; Butler, J.H. Non-CO<sub>2</sub> greenhouse gases and climate change. *Nature* **2011**, *476*, 43–50. [\[CrossRef\]](#)
3. Butterbach-Bahl, K.; Dannenmann, M. Denitrification and associated soil N<sub>2</sub>O emissions due to agricultural activities in a changing climate. *Curr. Opin. Environ. Sustain.* **2011**, *3*, 389–395. [\[CrossRef\]](#)
4. Sajjad, M.; Hussain, K.; Wajid, S.A.; Saqib, Z.A. The Impact of Split Nitrogen Fertilizer Applications on the Productivity and Nitrogen Use Efficiency of Rice. *Nitrogen* **2024**, *6*, 1. [\[CrossRef\]](#)
5. Ciarlo, E.A.; Llorente, M.M.; Conti, M.E.; Giardina, E.B. Soybean residue effect on carbon fractions and gaseous nitrogen losses at contrasting moisture contents. *Commun. Soil Sci. Plant Anal.* **2009**, *40*, 2756–2772. [\[CrossRef\]](#)
6. Aulakh, M.; Walters, D.; Doran, J.; Francis, D.; Mosier, A. Crop residue type and placement effects on denitrification and mineralization. *Soil Sci. Soc. Am. J.* **1991**, *55*, 1020–1025. [\[CrossRef\]](#)
7. Lai, T.V.; Farquharson, R.; Denton, M.D. High soil temperatures alter the rates of nitrification, denitrification and associated N<sub>2</sub>O emissions. *J. Soils Sediments* **2019**, *19*, 2176–2189. [\[CrossRef\]](#)
8. Russell, C.; Fillery, I.; Bootsma, N.; McInnes, K. Effect of temperature and nitrogen source on nitrification in a sandy soil. *Commun. Soil Sci. Plant Anal.* **2002**, *33*, 1975–1989. [\[CrossRef\]](#)
9. Hayatsu, M.; Tago, K.; Saito, M. Various players in the nitrogen cycle: Diversity and functions of the microorganisms involved in nitrification and denitrification. *Soil Sci. Plant Nutr.* **2008**, *54*, 33–45. [\[CrossRef\]](#)
10. Van Groenigen, J.; Huygens, D.; Boeckx, P.; Kuypers, T.W.; Lubbers, I.; Rütting, T.; Groffman, P. The soil N cycle: New insights and key challenges. *Soil* **2015**, *1*, 235–256. [\[CrossRef\]](#)
11. Signor, D.; Cerri, C.E.P. Nitrous oxide emissions in agricultural soils: A review. *Pesqui. Agropecuária Trop.* **2013**, *43*, 322–338. [\[CrossRef\]](#)
12. Bizimana, F.; Dong, W.; Timilsina, A.; Raseduzzaman, M.; Li, X.; Zhang, Y.; Hu, C. Effects of Straw Amendment in Combination with Synthetic N Fertilizer Addition on N<sub>2</sub>O, N<sub>2</sub>, and Their Stoichiometric Ratios in Three Different Agro-Ecosystems. *Agronomy* **2024**, *14*, 887. [\[CrossRef\]](#)
13. Powlson, D.; Saffigna, P.; Kragt-Cottaar, M. Denitrification at sub-optimal temperatures in soils from different climatic zones. *Soil Biol. Biochem.* **1988**, *20*, 719–723. [\[CrossRef\]](#)
14. Canion, A.; Kostka, J.E.; Gihring, T.; Huettel, M.; Van Beusekom, J.; Gao, H.; Lavik, G.; Kuypers, M.M. Temperature response of denitrification and anammox reveals the adaptation of microbial communities to in situ temperatures in permeable marine sediments that span 50 in latitude. *Biogeosciences* **2014**, *11*, 309–320. [\[CrossRef\]](#)
15. Adouani, N.; Limousy, L.; Lendormi, T.; Sire, O. N<sub>2</sub>O and NO emissions during wastewater denitrification step: Influence of temperature on the biological process. *Comptes Rendus. Chim.* **2015**, *18*, 15–22. [\[CrossRef\]](#)
16. Lazcano, C.; Zhu-Barker, X.; Decock, C. Effects of organic fertilizers on the soil microorganisms responsible for N<sub>2</sub>O emissions: A review. *Microorganisms* **2021**, *9*, 983. [\[CrossRef\]](#)
17. Wrage-Mönnig, N.; Horn, M.A.; Well, R.; Müller, C.; Velthof, G.; Oenema, O. The role of nitrifier denitrification in the production of nitrous oxide revisited. *Soil Biol. Biochem.* **2018**, *123*, A3–A16. [\[CrossRef\]](#)
18. Senbayram, M.; Chen, R.; Budai, A.; Bakken, L.; Dittert, K. N<sub>2</sub>O emission and the N<sub>2</sub>O/(N<sub>2</sub>O+N<sub>2</sub>) product ratio of denitrification as controlled by available carbon substrates and nitrate concentrations. *Agric. Ecosyst. Environ.* **2012**, *147*, 4–12. [\[CrossRef\]](#)
19. Thangarajan, R.; Bolan, N.S.; Tian, G.; Naidu, R.; Kunhikrishnan, A. Role of organic amendment application on greenhouse gas emission from soil. *Sci. Total Environ.* **2013**, *465*, 72–96. [\[CrossRef\]](#)
20. Broadbent, F.; Clark, F. Denitrification. *Soil Nitrogen* **1965**, *10*, 344–359.
21. Šimek, M.; Jiřová, L.; Hopkins, D.W. What is the so-called optimum pH for denitrification in soil? *Soil Biol. Biochem.* **2002**, *34*, 1227–1234. [\[CrossRef\]](#)
22. Coyne, M.S. Denitrification in soil. *Soil Nitrogen Uses Environ. Impacts* **2018**, *1*, 95–139.
23. Sumner, D.R. Crop rotation and plant productivity. In *Handbook of Agricultural Productivity*; CRC Press: Boca Raton, FL, USA, 2018; pp. 273–314.
24. Ghosh, P.K.; Hazra, K.K.; Venkatesh, M.S.; Praharaj, C.S.; Kumar, N.; Nath, C.P.; Singh, U.; Singh, S.S. Grain legume inclusion in cereal–cereal rotation increased base crop productivity in the long run. *Exp. Agric.* **2020**, *56*, 142–158. [\[CrossRef\]](#)

25. Drury, C.; Yang, X.; Reynolds, W.; McLaughlin, N. Nitrous oxide and carbon dioxide emissions from monoculture and rotational cropping of corn, soybean and winter wheat. *Can. J. Soil Sci.* **2008**, *88*, 163–174. [\[CrossRef\]](#)
26. Jeuffroy, M.; Baranger, E.; Carrouée, B.; De Chezelles, E.; Gosme, M.; Hénault, C.; Schneider, A.; Cellier, P. Nitrous oxide emissions from crop rotations including wheat, oilseed rape and dry peas. *Biogeosciences* **2013**, *10*, 1787–1797. [\[CrossRef\]](#)
27. Gomes, J.; Bayer, C.; de Souza Costa, F.; de Cássia Piccolo, M.; Zanatta, J.A.; Vieira, F.C.B.; Six, J. Soil nitrous oxide emissions in long-term cover crops-based rotations under subtropical climate. *Soil Tillage Res.* **2009**, *106*, 36–44. [\[CrossRef\]](#)
28. Li, T.; Li, Y.; Shi, Z.; Wang, S.; Wang, Z.; Liu, Y.; Wen, X.; Mo, F.; Han, J.; Liao, Y. Crop development has more influence on shaping rhizobacteria of wheat than tillage practice and crop rotation pattern in an arid agroecosystem. *Appl. Soil Ecol.* **2021**, *165*, 104016. [\[CrossRef\]](#)
29. Kakraliya, S.; Singh, U.; Bohra, A.; Choudhary, K.; Kumar, S.; Meena, R.S.; Jat, M. Nitrogen and legumes: A meta-analysis. In *Legumes for Soil Health and Sustainable Management*; Springer: Delhi, India, 2018; pp. 277–314.
30. Fagodiya, R.K.; Sharma, G.; Verma, K.; Rai, A.K.; Prajapat, K.; Singh, R.; Chandra, P.; Sheoran, P.; Yadav, R.K.; Biswas, A. Computation of soil quality index after fifteen years of long-term tillage and residue management experiment (LT&RE) under rice wheat system. *Agric. Syst.* **2024**, *219*, 104039.
31. Zhang, B.; Wang, G.; Liu, X. Research on corn and soybean rotation: A review of ecological benefits and economic potential. *Resour. Data J.* **2024**, *3*, 208–222.
32. Fageria, N.K.; Baligar, V.C.; Bailey, B.A. Role of cover crops in improving soil and row crop productivity. *Commun. Soil Sci. Plant Anal.* **2005**, *36*, 2733–2757. [\[CrossRef\]](#)
33. Singh, G.; Bhattacharyya, R.; Das, T.; Sharma, A.; Ghosh, A.; Das, S.; Jha, P. Crop rotation and residue management effects on soil enzyme activities, glomalin and aggregate stability under zero tillage in the Indo-Gangetic Plains. *Soil Tillage Res.* **2018**, *184*, 291–300. [\[CrossRef\]](#)
34. Drury, C.; Yang, X.; Reynolds, W.; Tan, C. Influence of crop rotation and aggregate size on carbon dioxide production and denitrification. *Soil Tillage Res.* **2004**, *79*, 87–100. [\[CrossRef\]](#)
35. Halvorson, A.D.; Del Grosso, S.J.; Reule, C.A. Nitrogen, tillage, and crop rotation effects on nitrous oxide emissions from irrigated cropping systems. *J. Environ. Qual.* **2008**, *37*, 1337–1344. [\[CrossRef\]](#) [\[PubMed\]](#)
36. Zhu, Y.; Li, Z.; Zhao, D.; Zhang, B.; Zhu, B.; Yao, Z.; Kiese, R.; Butterbach-Bahl, K.; Zhou, M. Effects of Conservation Agriculture on Soil N<sub>2</sub>O Emissions and Crop Yield in Global Cereal Cropping Systems. *Glob. Change Biol.* **2025**, *31*, e70048. [\[CrossRef\]](#) [\[PubMed\]](#)
37. Tariq, A.; Menheere, N.; Gao, Y.; Brown, S.; Van Eerd, L.L.; Lauzon, J.D.; Bruun, S.; Wagner-Riddle, C. Increased N<sub>2</sub>O emissions by cover crops in a diverse crop rotation can be mediated with dual nitrification and urease inhibitors. *Agric. Ecosyst. Environ.* **2024**, *374*, 109178. [\[CrossRef\]](#)
38. Kumar, T.; Kundu, M.S.; Jha, R.K. Impact of crop rotation and tillage operations on mitigating greenhouse gas emissions and evaluation of sustainability index in rice-wheat-green gram cropping system of north Bihar. *J. Environ. Manag.* **2024**, *366*, 121689. [\[CrossRef\]](#) [\[PubMed\]](#)
39. Wu, G.; Yang, S.; Luan, C.-s.; Wu, Q.; Lin, L.-l.; Li, X.-x.; Che, Z.; Zhou, D.-b.; Dong, Z.-r.; Song, H. Partial organic substitution for synthetic fertilizer improves soil fertility and crop yields while mitigating N<sub>2</sub>O emissions in wheat-maize rotation system. *Eur. J. Agron.* **2024**, *154*, 127077. [\[CrossRef\]](#)
40. Braker, G.; Matthies, D.; Hannig, M.; Brandt, F.B.; Brenzinger, K.; Gröngroft, A. Impact of land use management and soil properties on denitrifier communities of namibian savannas. *Microb. Ecol.* **2015**, *70*, 981–992. [\[CrossRef\]](#)
41. Desloover, J.; Vlaeminck, S.E.; Clauwaert, P.; Verstraete, W.; Boon, N. Strategies to mitigate N<sub>2</sub>O emissions from biological nitrogen removal systems. *Curr. Opin. Biotechnol.* **2012**, *23*, 474–482. [\[CrossRef\]](#)
42. Hu, H.-W.; Chen, D.; He, J.-Z. Microbial regulation of terrestrial nitrous oxide formation: Understanding the biological pathways for prediction of emission rates. *FEMS Microbiol. Rev.* **2015**, *39*, 729–749. [\[CrossRef\]](#)
43. Snyder, C.S.; Bruulsema, T.W.; Jensen, T.L.; Fixen, P.E. Review of greenhouse gas emissions from crop production systems and fertilizer management effects. *Agric. Ecosyst. Environ.* **2009**, *133*, 247–266. [\[CrossRef\]](#)
44. Hassan, M.U.; Aamer, M.; Mahmood, A.; Awan, M.I.; Barbanti, L.; Seleiman, M.F.; Bakhsh, G.; Alkharabsheh, H.M.; Babur, E.; Shao, J. Management strategies to mitigate N<sub>2</sub>O emissions in agriculture. *Life* **2022**, *12*, 439. [\[CrossRef\]](#) [\[PubMed\]](#)
45. Haas, E.; Carozzi, M.; Massad, R.S.; Butterbach-Bahl, K.; Scheer, C. Long term impact of residue management on soil organic carbon stocks and nitrous oxide emissions from European croplands. *Sci. Total Environ.* **2022**, *836*, 154932. [\[CrossRef\]](#)
46. Otchere, O. The Impact of Cover Cropping on Soil Nitrogen Availability, Crop Nitrogen Use, and Nitrous Oxide Emissions in a Prairie Potato—Grain Crop Rotation. Doctoral Dissertation, University of Saskatchewan, Saskatoon, SK, Canada, 2022.
47. Yoo, J.; Woo, S.-H.; Park, K.-D.; Chung, K.-Y. Effect of no-tillage and conventional tillage practices on the nitrous oxide (N<sub>2</sub>O) emissions in an upland soil: Soil N<sub>2</sub>O emission as affected by the fertilizer applications. *Appl. Biol. Chem.* **2016**, *59*, 787–797. [\[CrossRef\]](#)

48. Monteiro, R.C.; Franchini, J.C.; Jantalia, C.P.; Urquiaga, S.; Alves, B.J.R.; Boddey, R.M. Soil nitrous oxide emissions from a soybean-wheat succession under different tillage systems in Southern Brazil. *Rev. Bras. De Ciência Do Solo* **2023**, *47*, e0220135. [\[CrossRef\]](#)
49. Smith, K.; Thomson, P.; Clayton, H.; McTaggart, I.; Conen, F. Effects of temperature, water content and nitrogen fertilisation on emissions of nitrous oxide by soils. *Atmos. Environ.* **1998**, *32*, 3301–3309. [\[CrossRef\]](#)
50. Chadwick, D.; Cardenas, L.; Misselbrook, T.; Smith, K.; Rees, R.; Watson, C.; McGeough, K.; Williams, J.; Cloy, J.; Thorman, R. Optimizing chamber methods for measuring nitrous oxide emissions from plot-based agricultural experiments. *Eur. J. Soil Sci.* **2014**, *65*, 295–307. [\[CrossRef\]](#)
51. West, T.; Griffith, D.; Steinhardt, G.; Klodivko, E.; Parsons, S. Effect of tillage and rotation on agronomic performance of corn and soybean: Twenty-year study on dark silty clay loam soil. *J. Prod. Agric.* **1996**, *9*, 241–248. [\[CrossRef\]](#)
52. Connor, B.F.; Rose, D.L.; Noriega, M.C.; Murtaugh, L.K.; Abney, S.R. *Methods of Analysis by the US Geological Survey National Water Quality Laboratory—Determination of 86 Volatile Organic Compounds in Water by Gas Chromatography/Mass Spectrometry, Including Detections Less Than Reporting Limits*; US Geological Survey: Denver, CO, USA, 1998.
53. Cardenas, L.; Thorman, R.; Ashlee, N.; Butler, M.; Chadwick, D.; Chambers, B.; Cuttle, S.; Donovan, N.; Kingston, H.; Lane, S. Quantifying annual N<sub>2</sub>O emission fluxes from grazed grassland under a range of inorganic fertiliser nitrogen inputs. *Agric. Ecosyst. Environ.* **2010**, *136*, 218–226. [\[CrossRef\]](#)
54. Louro, A.; Sawamoto, T.; Chadwick, D.; Pezzolla, D.; Bol, R.; Báez, D.; Cardenas, L. Effect of slurry and ammonium nitrate application on greenhouse gas fluxes of a grassland soil under atypical South West England weather conditions. *Agric. Ecosyst. Environ.* **2013**, *181*, 1–11. [\[CrossRef\]](#)
55. McLean, E. Soil pH and lime requirement. *Methods Soil Anal. Part 2 Chem. Microbiol. Prop.* **1983**, *9*, 199–224.
56. Wheatley, R.; MacDonald, R.; Smith, A.M. Extraction of nitrogen from soils. *Biol. Fertil. Soils* **1989**, *8*, 189–190. [\[CrossRef\]](#)
57. Sims, J.T. Soil test phosphorus: Bray and Kurtz P-1. In *Methods of Phosphorus Analysis for Soils, Sediments, Residuals, and Waters*; Researchgate.net: Newark, NJ, USA, 2000; Volume 13.
58. Dickey, G.; Allen, R.; Wright, J.; Murray, N.; Stone, J.; Hunsaker, D. Soil bulk density sampling for neutron gauge calibration. *Irrig. Drain. Syst.* **1993**, 1103–1111.
59. Amirinejad, A.A.; Kamble, K.; Aggarwal, P.; Chakraborty, D.; Pradhan, S.; Mittal, R.B. Assessment and mapping of spatial variation of soil physical health in a farm. *Geoderma* **2011**, *160*, 292–303. [\[CrossRef\]](#)
60. Gasche, R.; Papen, H. A 3-year continuous record of nitrogen trace gas fluxes from untreated and limed soil of a N-saturated spruce and beech forest ecosystem in Germany: 2. NO and NO<sub>2</sub> fluxes. *J. Geophys. Res. Atmos.* **1999**, *104*, 18505–18520. [\[CrossRef\]](#)
61. Roelle, P.A.; Aneja, V.P.; Gay, B.; Geron, C.; Pierce, T. Biogenic nitric oxide emissions from cropland soils. *Atmos. Environ.* **2001**, *35*, 115–124. [\[CrossRef\]](#)
62. Schindlbacher, A.; Zechmeister-Boltenstern, S.; Butterbach-Bahl, K. Effects of soil moisture and temperature on NO, NO<sub>2</sub>, and N<sub>2</sub>O emissions from European forest soils. *J. Geophys. Res. Atmos.* **2004**, *109*. [\[CrossRef\]](#)
63. Luo, J.; Tillman, R.; Ball, P. Grazing effects on denitrification in a soil under pasture during two contrasting seasons. *Soil Biol. Biochem.* **1999**, *31*, 903–912. [\[CrossRef\]](#)
64. Drury, C.; Reynolds, W.; Tan, C.; Welacky, T.; Calder, W.; McLaughlin, N. Emissions of nitrous oxide and carbon dioxide: Influence of tillage type and nitrogen placement depth. *Soil Sci. Soc. Am. J.* **2006**, *70*, 570–581. [\[CrossRef\]](#)
65. Johansen, A.; Carter, M.S.; Jensen, E.S.; Hauggard-Nielsen, H.; Ambus, P. Effects of digestate from anaerobically digested cattle slurry and plant materials on soil microbial community and emission of CO<sub>2</sub> and N<sub>2</sub>O. *Appl. Soil Ecol.* **2013**, *63*, 36–44. [\[CrossRef\]](#)
66. Coppens, F.; Garnier, P.; De Gryze, S.; Merckx, R.; Recous, S. Soil moisture, carbon and nitrogen dynamics following incorporation and surface application of labelled crop residues in soil columns. *Eur. J. Soil Sci.* **2006**, *57*, 894–905. [\[CrossRef\]](#)
67. Chen, Z.; Ge, S.; Zhang, Z.; Du, Y.; Yao, B.; Xie, H.; Liu, P.; Zhang, Y.; Wang, W.; Zhou, H. Soil moisture but not warming dominates nitrous oxide emissions during freeze–thaw cycles in a Qinghai–Tibetan plateau alpine meadow with discontinuous permafrost. *Front. Ecol. Evol.* **2021**, *9*, 676027. [\[CrossRef\]](#)
68. Russow, R.; Sich, I.; Neue, H.-U. The formation of the trace gases NO and N<sub>2</sub>O in soils by the coupled processes of nitrification and denitrification: Results of kinetic <sup>15</sup>N tracer investigations. *Chemosphere-Glob. Change Sci.* **2000**, *2*, 359–366. [\[CrossRef\]](#)
69. Wu, D.; Dong, W.; Oenema, O.; Wang, Y.; Trebs, I.; Hu, C. N<sub>2</sub>O consumption by low-nitrogen soil and its regulation by water and oxygen. *Soil Biol. Biochem.* **2013**, *60*, 165–172. [\[CrossRef\]](#)
70. Bateman, E.; Baggs, E. Contributions of nitrification and denitrification to N<sub>2</sub>O emissions from soils at different water-filled pore space. *Biol. Fertil. Soils* **2005**, *41*, 379–388. [\[CrossRef\]](#)
71. Iqbal, J.; Mitchell, D.C.; Barker, D.W.; Miguez, F.; Sawyer, J.E.; Pantoja, J.; Castellano, M.J. Does nitrogen fertilizer application rate to corn affect nitrous oxide emissions from the rotated soybean crop? *J. Environ. Qual.* **2015**, *44*, 711–719. [\[CrossRef\]](#) [\[PubMed\]](#)

72. Wang, X.; Lu, P.; Yang, P.; Ren, S. Effects of fertilizer and biochar applications on the relationship among soil moisture, temperature, and N<sub>2</sub>O emissions in farmland. *PeerJ* **2021**, *9*, e11674. [\[CrossRef\]](#)
73. Del Prado, A.; Merino, P.; Estavillo, J.; Pinto, M.; González-Murua, C. N<sub>2</sub>O and NO emissions from different N sources and under a range of soil water contents. *Nutr. Cycl. Agroecosyst.* **2006**, *74*, 229–243. [\[CrossRef\]](#)
74. Li, Y.; Barton, L.; Chen, D. Simulating response of N<sub>2</sub>O emissions to fertiliser N application and climatic variability from a rain-fed and wheat-cropped soil in Western Australia. *J. Sci. Food Agric.* **2012**, *92*, 1130–1143. [\[CrossRef\]](#)
75. Carmo, J.B.d.; Neill, C.; Garcia-Montiel, D.C.; de Cássia Piccolo, M.; Cerri, C.C.; Steudler, P.A.; de Andrade, C.A.; Passianoto, C.C.; Feigl, B.J.; Melillo, J.M. Nitrogen dynamics during till and no-till pasture restoration sequences in Rondônia, Brazil. *Nutr. Cycl. Agroecosyst.* **2005**, *71*, 213–225. [\[CrossRef\]](#)
76. Ruser, R.; Flessa, H.; Russow, R.; Schmidt, G.; Buegger, F.; Munch, J. Emission of N<sub>2</sub>O, N<sub>2</sub> and CO<sub>2</sub> from soil fertilized with nitrate: Effect of compaction, soil moisture and rewetting. *Soil Biol. Biochem.* **2006**, *38*, 263–274. [\[CrossRef\]](#)
77. Zanatta, J.A.; Bayer, C.; Vieira, F.C.; Gomes, J.; Tomazi, M. Nitrous oxide and methane fluxes in South Brazilian Gleysol as affected by nitrogen fertilizers. *Rev. Bras. De Ciência Do Solo* **2010**, *34*, 1653–1665. [\[CrossRef\]](#)
78. Mosier, A.R. Exchange of gaseous nitrogen compounds between agricultural systems and the atmosphere. *Plant Soil* **2001**, *228*, 17–27. [\[CrossRef\]](#)
79. Khalil, K.; Mary, B.; Renault, P. Nitrous oxide production by nitrification and denitrification in soil aggregates as affected by O<sub>2</sub> concentration. *Soil Biol. Biochem.* **2004**, *36*, 687–699. [\[CrossRef\]](#)
80. Rochette, P.; Janzen, H.H. Towards a revised coefficient for estimating N<sub>2</sub>O emissions from legumes. *Nutr. Cycl. Agroecosyst.* **2005**, *73*, 171–179. [\[CrossRef\]](#)
81. Escobar, L.F.; Amado, T.J.C.; Bayer, C.; Chavez, L.F.; Zanatta, J.A.; Fiorin, J.E. Postharvest nitrous oxide emissions from a subtropical oxisol as influenced by summer crop residues and their management. *Rev. Bras. De Ciência Do Solo* **2010**, *34*, 507–516. [\[CrossRef\]](#)
82. Rochette, P. No-till only increases N<sub>2</sub>O emissions in poorly-aerated soils. *Soil Tillage Res.* **2008**, *101*, 97–100. [\[CrossRef\]](#)
83. Zschornack, T.; Bayer, C.; Zanatta, J.A.; Vieira, F.C.B.; Anghinoni, I. Mitigation of methane and nitrous oxide emissions from flood-irrigated rice by no incorporation of winter crop residues into the soil. *Rev. Bras. De Ciência Do Solo* **2011**, *35*, 623–634. [\[CrossRef\]](#)
84. Almaraz, J.J.; Zhou, X.; Mabood, F.; Madramootoo, C.; Rochette, P.; Ma, B.-L.; Smith, D.L. Greenhouse gas fluxes associated with soybean production under two tillage systems in southwestern Quebec. *Soil Tillage Res.* **2009**, *104*, 134–139. [\[CrossRef\]](#)
85. Nouchi, I.; Yonemura, S. CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O fluxes from soybean and barley double-cropping in relation to tillage in Japan. *Phyton* **2005**, *45*, 327–338.
86. Asakawa, S. Denitrifying ability of indigenous strains of Bradyrhizobium japonicum isolated from fields under paddy-upland rotation. *Biol. Fertil. Soils* **1993**, *15*, 196–200. [\[CrossRef\]](#)
87. Inaba, S.; Ikenishi, F.; Itakura, M.; Kikuchi, M.; Eda, S.; Chiba, N.; Katsuyama, C.; Suwa, Y.; Mitsui, H.; Minamisawa, K. N<sub>2</sub>O emission from degraded soybean nodules depends on denitrification by Bradyrhizobium japonicum and other microbes in the rhizosphere. *Microbes Environ.* **2012**, *27*, 470–476. [\[CrossRef\]](#) [\[PubMed\]](#)
88. Kim, S.U.; Lee, H.H.; Moon, S.M.; Han, H.R.; Hong, C.O. Nitrous oxide emissions and maize yield as influenced by nitrogen fertilization and tillage operations in upland soil. *Appl. Biol. Chem.* **2021**, *64*, 1–12. [\[CrossRef\]](#)
89. Omonode, R.A.; Halvorson, A.D.; Gagnon, B.; Vyn, T.J. Achieving lower nitrogen balance and higher nitrogen recovery efficiency reduces nitrous oxide emissions in North America's maize cropping systems. *Front. Plant Sci.* **2017**, *8*, 1080. [\[CrossRef\]](#) [\[PubMed\]](#)
90. Peigné, J.; Ball, B.; Roger-Estrade, J.; David, C. Is conservation tillage suitable for organic farming? A review. *Soil Use Manag.* **2007**, *23*, 129–144. [\[CrossRef\]](#)
91. Franzluebbers, A.; Hons, F.; Zuberer, D. Tillage and crop effects on seasonal soil carbon and nitrogen dynamics. *Soil Sci. Soc. Am. J.* **1995**, *59*, 1618–1624. [\[CrossRef\]](#)
92. Malhi, S.S.; Lemke, R.; Wang, Z.; Chhabra, B.S. Tillage, nitrogen and crop residue effects on crop yield, nutrient uptake, soil quality, and greenhouse gas emissions. *Soil Tillage Res.* **2006**, *90*, 171–183. [\[CrossRef\]](#)
93. Pekrun, C.; Messelhäuser, M.H.; Finck, M.; Hartung, K.; Möller, K.; Gerhards, R. Yield, soil Nitrogen content and weed control in six years of conservation agriculture on-farm field trials in Southwest Germany. *Soil Tillage Res.* **2023**, *227*, 105615. [\[CrossRef\]](#)
94. Kaspar, T.; Jaynes, D.; Parkin, T.; Moorman, T.; Singer, J. Effectiveness of oat and rye cover crops in reducing nitrate losses in drainage water. *Agric. Water Manag.* **2012**, *110*, 25–33. [\[CrossRef\]](#)
95. Gillette, K.; Malone, R.; Kaspar, T.; Ma, L.; Parkin, T.; Jaynes, D.; Fang, Q.; Hatfield, J.; Feyereisen, G.; Kersebaum, K. N loss to drain flow and N<sub>2</sub>O emissions from a corn-soybean rotation with winter rye. *Sci. Total Environ.* **2018**, *618*, 982–997. [\[CrossRef\]](#)
96. Toliver, D.K.; Larson, J.A.; Roberts, R.K.; English, B.C.; De La Torre Ugarte, D.G.; West, T.O. Effects of no-till on yields as influenced by crop and environmental factors. *Agron. J.* **2012**, *104*, 530–541. [\[CrossRef\]](#)

97. Evanylo, G. No-Till Corn Response to Nitrogen Rate and Timing in the Middle Atlantic Coastal Plain. *J. Prod. Agric.* **1991**, *4*, 180–185. [[CrossRef](#)]
98. Yin, X.; Al-Kaisi, M.M. Periodic response of soybean yields and economic returns to long-term no-tillage. *Agron. J.* **2004**, *96*, 723–733. [[CrossRef](#)]
99. Parkin, T.B.; Kaspar, T.C. Nitrous oxide emissions from corn–soybean systems in the Midwest. *J. Environ. Qual.* **2006**, *35*, 1496–1506. [[CrossRef](#)]

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