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1 **Physical properties of a sandy soil as affected by incubation with a**
2 **synthetic root exudate: strength, thermal and hydraulic conductivity,**
3 **and evaporation**

4 **Running title: Synthetic root exudate on soil physical property**

5 W. ZHANG ^a, W. GAO ^a, W. R. WHALLEY ^b, T. REN ^{a,*}

6 ^a *Department of Soil and Water Sciences, China Agricultural University, Beijing*
7 *100193, China*

8 ^b *Department of Sustainable Agricultural Sciences, Rothamsted Research, Harpenden*
9 *AL5 2JQ, UK*

10

11 Email addresses:

12 Wencan Zhang: wencanzhang@163.com

13 Weida Gao: weida_gao@cau.edu.cn

14 W. Richard Whalley: richard.whalley@rothamsted.ac.uk

15 Tusheng Ren: tsren@cau.edu.cn

16

17 ***Corresponding author:** Tusheng Ren.

18

19 **Summary**

20 Plant roots release various organic materials that may modify soil structure and affect
21 heat and mass transfer processes. The objective of this study was to determine the
22 effects of a synthetic root exudate (SRE) on penetrometer resistance (PR), thermal
23 conductivity (λ), hydraulic conductivity (k) and evaporation of water in a sandy soil.
24 Soil samples, mixed with either distilled water or the SRE, were packed into columns
25 at a designated bulk density and water content, and incubated for 7 days at 18°C. Soil
26 PR, λ , k and evaporation rate were monitored during drying processes. Compared with
27 those incubated with water, samples incubated with SRE had visible hyphae, greater
28 PR (0.7-5.5 MPa higher) and λ (0.2-0.7 W m⁻¹ K⁻¹ higher) in the water content range of
29 0.11-0.22 and 0.05-0.22 m³ m⁻³, increased k in the wet region but decreased k in the dry
30 region. SRE treatment also reduced the overall soil water evaporation rate and
31 cumulative water loss. Analysis of CT scanning showed that the SRE treated samples
32 had a greater proportion of small pores (< 60 μ m). These changes were attributed
33 mainly to SRE-simulated microbial activities.

34

35 **Keywords:** Soil Penetrometer Resistance, Heat Conduction, Water Retention,
36 Microbial Activity

37 **Highlights**

38 □ The effects of incubating a sandy soil with a synthetic root exudate (SRE) on soil
39 physical properties and evaporation are examined.

40 □ SRE incubation increased the fraction of small pores.

41 □ SRE incubation increased soil penetrometer resistance and thermal conductivity.

42 □ Soil hydraulic conductivity was increased in the wet region but was reduced in the
43 dry region.

44 □ SRE incubation reduced the overall evaporation rate and cumulative water loss.

45

46 **Introduction**

47 Secretion or exudation from plant roots and microbes, often termed mucilage or
48 exudates, affects the structure and physical properties of soils (Czarnes *et al.*, 2000;
49 Read *et al.*, 2003; Whalley *et al.*, 2005; Barré & Hallett, 2009; Lambers *et al.*, 2009;
50 Carminati *et al.*, 2010; Choudhury *et al.*, 2018). Mucilage, a matrix of higher-
51 molecular-weight compounds (Sasse *et al.*, 2018), is polysaccharide-rich and can
52 behave as a viscoelastic gel. Exudates are composed of a wide range of components,
53 including mucilage and other low-molecular-weight or soluble high-molecular-weight
54 compounds (Galloway *et al.*, 2020). Holz *et al.* (2015) found that root exudates and
55 mucilage have a high degree of spatial structure with distance from the root tip and with
56 radial distance from the root. The chemical compounds in natural root exudates depend
57 on many factors, such as the plant species and age, the soil microbial communities and
58 activity, as well as the soil chemical and physical status (Naveed *et al.*, 2017).

59 The organic compounds in root exudates play an important role in the plant-
60 microbe interaction by affecting soil microbial communities structure and function (Shi
61 *et al.*, 2011), and the utilization of this type of carbon is an important driver of soil
62 microbial abundance. Soil microbial activity and community development is often
63 stimulated or determined by natural root exudates (Paterson *et al.*, 2007) or synthetic
64 root exudates (Gao *et al.* 2017; Choudhury *et al.*, 2018). The changes in microbial
65 community or microbial activity will in turn alter soil physical properties. Gao *et al.*
66 (2017) found that microbial activity was stimulated by a low molecular synthetic root
67 exudate, and that a poikilothermic temperature response of soil penetrometer resistance

68 seemed to be related to fungi or streptomycetes, a filamentous bacterium. As temporary
69 binding agents, fungal hyphae can bond microaggregates (< 0.25 mm diameter) into
70 stable macroaggregates (> 0.25 mm diameter) (Tisdall 1991). Root exudates also
71 modify soil structure by altering aggregates stability, strength and particle contacts
72 (Czarnes *et al.*, 2000; Read *et al.*, 2003; Whalley *et al.*, 2005; Hinsinger *et al.*, 2009;
73 Carminati *et al.*, 2010). For example, there are reports that root exudates or mucilage
74 increased soil aggregate stability (Morel *et al.*, 1991; Traoré *et al.*, 2000). After adding
75 polygalacturonic acid (PGA) and xanthan, soil strength and stability were increased
76 following wetting and drying cycles, which was attributed to the increased particle
77 bonding strength by PGA (Czarnes *et al.*, 2000). Similarly, some researchers observed
78 that addition of PGA led to the increase of bonding energy among soil particles (Zhang
79 *et al.*, 2008), and enhanced soil hardness and modulus of elasticity, and the degree of
80 these changes varied with the source of the exudates (Naveed *et al.*, 2017). Soil
81 structure can be affected by root exudates *per se*, specifically their high viscosity and
82 low surface tension (Lavelle, 2002; Strayer *et al.*, 2003; Gregory, 2006; Watt *et al.*,
83 2006; Hinsinger *et al.*, 2009).

84 The changes of soil structure in rhizosphere caused by root exudates may alter soil
85 hydraulic properties and processes. Zhang *et al.* (2008) observed a decrease of
86 evaporation rate and an increase of soil water retention, which were attributed to the
87 extracellular polymeric substances (EPS) produced by a plant growth-promoting
88 rhizobacteria. There are reports that root exudates can both increase soil water holding
89 capacity (Young, 1995; Read *et al.*, 1999; Nakanishi *et al.*, 2005) as well as reduce

90 water contents in the rhizosphere (Read *et al.*, 2003; Whalley *et al.*, 2005; Dunbabin *et*
91 *al.*, 2006). An increase in soil water repellence has also been reported (Carminati *et al.*,
92 2010; Moradi *et al.*, 2012). These conflicting findings with respect to water retention
93 are more likely to be explained by the different types of compounds secreted by roots,
94 and some polymeric gels (e.g., glucose) are generally present to increase water holding
95 capacity while some surfactants (e.g., phospholipid) act as decreasing water holding
96 capacity (Read *et al.*, 2003). Drying is an important process and it highlights the
97 structural difference between rhizosphere and bulk soil. For example, Nambiar (1976)
98 and Watt *et al.* (1994) observed the formation of a more stable maize rhizosphere in the
99 drier soils; Albalasmeh & Ghezzehei (2014) reported that drying process enhanced soil
100 aggregation by transporting and depositing binding agents, such as PGA and xanthan;
101 and Benard *et al.* (2017) pointed out that root exudates were deposited preferentially in
102 small pores during drying. A micro-morphological study showed an increase in porosity
103 of mucilage-treated soil following wetting and drying cycles (Czarnes *et al.*, 2000).

104 Heat transfer, available water, nutrient contents, aeration and soil strength are
105 important factors that determine both microbial activity as well as root growth. Soil
106 thermal conductivity (λ) is a parameter that characterizes soil heat conduction. It is
107 determined by the volumetric proportions of solid, liquid and gaseous phases, the
108 arrangement of soil particles, and the interfacial contact between the solid particles as
109 well as between the solid and liquid phases (Carson *et al.*, 2003; Farouki, 1986). It is
110 likely that microbial activity that alters pore size distribution or particle bonding will
111 also affect λ , but to our knowledge this has never been studied. Soil penetrometer

112 resistance (PR), which is commonly used to characterize the soil strength and resistance
113 to root penetration, depends on soil structure and water status (Bengough *et al.*, 2000;
114 Vaz *et al.*, 2011), particle size, shape and distribution, and the chemistry of the soil
115 solution (Horn, 1994). Gao *et al.* (2017) observed an increase of PR in soil samples
116 incubated with a synthetic root exudate. Evaporation of water from a bare soil surface
117 depends on the hydraulic connectivity of soil and it is widely accepted this can be
118 modified by the activity of both roots and microbes.

119 In brief, root exudates alter the rhizosphere soil structure and water holding
120 capacity, but it is unclear how other soil physical properties and processes respond to
121 these changes. In this study we investigate the changes of soil PR, λ , k , and evaporation
122 following the addition of a synthetic root exudate, which is known to stimulate
123 microbial activity (Gao *et al.*, 2017; Choudhury *et al.*, 2018). The effect of root exudate
124 on soil pore size distribution was also examined with X-ray CT. This the first report of
125 the integrated effects of a root exudate on thermal, hydraulic and mechanical properties
126 of the rhizosphere.

127 **Materials and Methods**

128 **Soil sample and synthetic root exudate**

129 The soil sample was collected in May 2017 from the surface layer (0-20 cm) of a
130 maize field located in Lishu County of Jilin Province, China. The soil has a sand texture
131 with 92% sand, 3% silt, and 5% clay. After removing crop residues and fine roots, the
132 partly air-dried soil sample (with a water content about 0.07 g g⁻¹) was passed through
133 a 2-mm sieve and then stored in the dark at 4°C.

134 A synthetic root exudate (SRE) solution was prepared by mixing 15 compounds,
135 including five large molecular polysaccharides (glucose, fructose, sucrose, arabinose
136 and ribose), five amino acids (glycine, valine, glutamine, serine and alanine) and five
137 organic acids (malic, citric, malonic, oxalic and fumaric), following the procedures of
138 Paterson *et al.* (2007). Each compound contributed 1.39 g C in 500 ml distilled water,
139 gave a solution with 4.167% C and 7.34% N.

140 **Soil column preparation and incubation experiment**

141 To examine the effects of microbial activity on soil PR, λ , k , water evaporation rate
142 and soil pore size distributions with X-ray CT, we prepared soil columns with SRE-
143 treated and distilled water-treated samples (control). The sieved soil sample (with initial
144 water content of about 0.07 g g⁻¹) was mixed with either synthetic root exudate or
145 distilled water at a concentration of 6 ml solution 100 g⁻¹ dry soil, which gave a water
146 content of 0.13 g g⁻¹ (close to field capacity). This concentration was equivalent to 7.2
147 mg SRE g⁻¹ dry soil, which was greater than the nature exudate concentration (4.6 mg
148 exudate g⁻¹ dry soil) reported by Zickenrott *et al.* (2016), but was within the range (4
149 and 8 mg g⁻¹ particle) of Benard *et al.* (2019).

150 For PR, λ , and pore structure measurements, paired soil samples (mixed with SRE
151 and water) were packed into polycarbonate pipes (50-mm high and 50-mm I.D.) with a
152 200 kPa axial pressure (equivalent to the stress of a tractor), which gave an approximate
153 bulk density of 1.61 g cm⁻³. A total of 40 soil columns (20 for each treatment) were
154 prepared. These samples were incubated in the dark for 7 days a temperature of 18°C.
155 Among them, 34 samples (17 for each treatment) were placed in a temperature-

156 regulated room ($25 \pm 1^\circ\text{C}$) and subjected to air-drying, and the PR and λ were
157 monitored continuously: 30 samples (5 times and 3 reps for each treatment) for PR
158 measurement and 4 samples (2 reps of each treatment) were for monitoring λ
159 continuously. The remaining 6 samples (3 reps for each treatment) were air-dried and
160 used to examine the pore structure by using X-ray CT scanning.

161 To examine the effects of SRE addition on soil water evaporation rate and hydraulic
162 conductivity, 6 larger soil columns (about 61-mm high, 72-mm I.D., and a volume of
163 250 cm^3 , triplicate samples for each treatment) were prepared in the same way as the
164 smaller columns. and incubated in the dark for 7 days at 18°C

165 **Measuring soil evaporation and hydraulic conductivity**

166 The six larger samples were saturated with distilled water and placed on a ku-pF
167 apparatus (DT 04-01, Umwelt-Geräte-Technik GmbH, Müncheberg, Germany). The
168 equipment has a star-shaped revolving table that has 10 sample holders, each carries
169 one sample ring and two tensiometers. The distance between the top and bottom
170 tensiometers is 3 cm. Sample weights were recorded with an automatically lowering
171 and lifting balance at a 1-h interval. Soil water evaporation rates were calculated from
172 the dynamics of weight losses. The hydraulic conductivity k , was calculated by using
173 Eq. (1),

$$174 \quad k = \frac{\Delta V}{2A\Delta t(\Psi_t - \Psi_b) - \Delta h} \quad (1)$$

175 where ΔV is the volume of evaporative water loss during time Δt , A is cross-sectional
176 area of sample ring, Ψ_t is tension of top tensiometer (as positive pressure in
177 unsaturated range), Ψ_b is tension of bottom tensiometer, Δz is distance between the

178 tensiometers in the sample ring (= 3 cm), and Δh is the altitude difference between
179 the tensiometer positions (= 3 cm).

180 **Measuring soil penetrometer resistance**

181 The PR of soil samples were measured with a universal testing machine with a
182 measuring range of 0-100 N and an accuracy of 0.00001 N (Model UTM6102,
183 Shenzhen Suns Technology Stock Co., Ltd. Shenzhen, China). The machine was
184 equipped with a 2-mm diameter and 60° cone angle penetrometer. The penetrometer
185 was inserted into soil samples at a constant speed of 20 mm min⁻¹ (Gao *et al.*, 2012;
186 Moraes *et al.*, 2019) from the surface to a depth of 45 mm (to protect the pressure
187 transducer from exceeding maximum load), and the PR was recorded automatically.
188 After PR measurements, the samples were oven-dried at 105°C for more than 24 hours
189 to determine bulk density and water content. The PR measurements were conducted
190 periodically during a 5-day drying period.

191 **Measuring soil thermal conductivity**

192 We used a three-needle heat-pulse probe (40-mm long and 1.3-mm-diameter
193 stainless steel waveguides with 6-mm spacing) to measure λ (Ren *et al.*, 1999). The
194 probe was inserted into the soil column vertically. A constant current was applied to the
195 central needle (which included a heater) for 7-10 s to generate the heat pulse. A data
196 logger (CR 3000, Campbell Scientific, Logan, UT) recorded the temperature change at
197 the sensing needles and the power input at a 1-s interval for 300 s. For each sample, the
198 measurement was repeated for three times at a 60-mins time interval. The temperature-
199 by-time data was analyzed by using the single point method (Bristow *et al.*, 1994,

200 1995). The daily λ measurements were continued for 8 days until the water contents
201 approached a relatively stable state. At the end of drying process, soil bulk density and
202 water content were determined by oven-drying the samples at 105°C for more than 24
203 h.

204 **X-ray CT scanning and image processing**

205 The soil cores (three for each treatment) were scanned with an industrial Phoenix
206 Nanotom X-ray μ -CT (GE, Sensing and Inspection Technologies, GmbH, Wunstorf,
207 Germany) at an energy of 100 kV and a current of 100 μ A. A 0.2-mm Cu filter was
208 used to reduce the beam hardening effect. The filtered back-projection algorithm was
209 used to reconstruct slices from the radiographs. About 2000 slices with a size of 2000
210 \times 2000 pixel for each slice were reconstructed for every sample. The final slices were
211 in 16-bit format, with a resolution of 10 μ m.

212 Images processing, visualization, and quantification were done with an open
213 source software Image J ver. 1.51 (Rasband, 1997-2011). Images were first imported,
214 changed to 8-bit format and adjusted the brightness. We used a global threshold
215 segmentation method to segment the images, and the threshold selection was carefully
216 chosen based on the visual observation method of Zhou *et al.* (2012). A region of
217 interest (ROI) of 1100 \times 1100 \times 1100 voxels was selected from the central part of soil
218 cores, representing a volume of 1.1 \times 1.1 \times 1.1 cm³. The total porosity and pore size
219 distribution were analyzed with the “Friction Volume” and “Thickness” plugins. Pore
220 sizes obtained from “Thickness” were expressed as the equivalent diameter. Pores were
221 classified with a 20- μ m interval diameter from 20 to 500 μ m.

222 **Statistical analysis**

223 For the comparison of pore size distribution obtained by X-ray CT scanning,
224 porosity was modeled using “Gaussian” regression with logarithmically (base 10)
225 transformed soil pore size as the explanatory variate, porosity as the response variate,
226 and treatments of distilled water and root exudates as grouping factors. The curves were
227 fitted to data with grouped regression. We used Genstat (VSN Int. Ltd, Hemel
228 Hempstead, UK, or Payne, 2015) to fit the curves described above to our data and for
229 the statistical analysis.

230 **Results**

231 **Soil penetrometer resistance**

232 Figure 1 shows the measurements of soil PR at different water contents during the
233 drying process. Under both SRE and water treatments, there was an exponential
234 increase in PR with decreasing water content, and addition of root exudate increased
235 PR at a specific water content ($P < 0.05$). Furthermore, the difference between SRE-
236 treated and water-treated samples became larger with decreasing water content. For
237 example, when the evaporation experiment was initiated right after incubation, both
238 root exudate and water treatments had a water content of $0.22 \text{ m}^3 \text{ m}^{-3}$, the PR of SRE-
239 and water-treated samples was 1.29 MPa and 0.58 MPa, respectively; when soil water
240 content was reduced to about $0.12 \text{ m}^3 \text{ m}^{-3}$ (i.e., day 5 for water treatment, and day 6 for
241 SRE treatment), the PR of SRE-treated samples was 5.52 MPa larger than that of water-
242 treated samples. Thus, incubation with the SRE increased PR, and the effect was
243 especially noticeable under dry conditions.

244 **Soil thermal conductivity**

245 Figure 2 presents the $\lambda(\theta_v)$ curves under both SRE and water treatments. The λ
246 values on the first day were obtained right after incubation at 18°C, and the remaining
247 values were daily measurements during the 8-day evaporation process. For both SRE
248 and water treatments, λ decreased nonlinearly with soil drying, and the rate of λ change
249 showed two distinct stages: A slow λ change appeared at soil water contents greater
250 than 0.1 m³ m⁻³, and a relatively rapid reduction at water contents smaller than 0.1 m³
251 m⁻³. At a specific water content, SRE-treated samples had significantly higher λ values
252 than that of water-treated samples. In the 0.05-0.21 m³ m⁻³ water content range, for
253 example, the SRE-treated sample had a λ range of 1.2-2.1 W m⁻¹ K⁻¹, and the
254 corresponding λ range of water-treated samples was 0.8-1.8 W m⁻¹ K⁻¹. The λ difference
255 between the two treatments was reduced slightly in the later stages of drying.

256 **Soil water evaporation**

257 The soil drying process usually involves three stages of water evaporation: a
258 relatively constant rate stage, a falling rate stage, and a residual stage (Amano &
259 Salvucci, 1999; Suleiman & Ritchie, 2003). Addition of the SRE changed soil water
260 evaporation at all three stages (Figure 3a). In stage I, both SRE- and water-treated
261 samples had similar evaporation rates (about 0.10-0.12 mm h⁻¹) but the duration of
262 SRE-treated samples was reduced to 64 h from 94 h of water-treated ones. In stage II,
263 evaporation rates of both SRE- and water-treated samples were reduced significantly,
264 but the samples with SRE had a lower rate of reduction and a relatively longer duration
265 (14 h) than that samples with water. In stage III when both treatments had low and

266 relatively stable evaporation rates, the values (0.034-0.015 mm h⁻¹) of SRE-treated
267 samples were slightly higher than that (0.034-0.007 mm h⁻¹) of water-treated ones.

268 The SRE- and water-treated samples had similar values of cumulative evaporation
269 in the 0-60 h period, but after 60 h when SRE-treated samples had lower cumulative
270 evaporation than that of water-treated samples (Figure 3b). At the end of the
271 evaporation study (393 h), the cumulative evaporation values of the SRE- and water-
272 treated samples were 26.02 mm and 28.31 mm, respectively.

273 **Soil hydraulic conductivity**

274 Figure 4 presents the results of soil hydraulic conductivity as a function of water
275 content (θ_v). For both treatments, the $k(\theta_v)$ curves could be divided into two parts: a
276 relatively flat section with greater k values at higher water contents, and a steep section
277 with k values decreased with water content. The SRE treatment altered the shape of the
278 $k(\theta_v)$ curve significantly: 1) The SRE treatment had a greater θ_v value (0.30 m³ m⁻³)
279 than that of the SRE treatment (0.26 m³ m⁻³) at the inflection point between the two
280 sections; 2) In the flat section, SRE-treated samples had higher hydraulic conductivities
281 than the water-treated samples; 3) In the steep section, a greater falling rate of soil
282 hydraulic conductivity was observed with the SRE-treated samples, and its k values
283 were lower than that of the water-treated samples at $\theta_v < 0.28$ m³ m⁻³.

284 **Soil pore-size distribution**

285 Figure 5 presents the grayscale and binary images of SRE- and water-treated
286 samples from X-ray CT scanning. Visual observation indicated that the images of water
287 -treated sample had more larger pores while the images of SRE-treated sample showed

288 more smaller pores. Pores size distribution data from X-ray CT images are shown in
289 Figure 6. The curves shown were fitted by using grouped regression. We used the
290 Genstat function, PRNORMAL (x, m, v), which is a probability density function for a
291 Normal distribution with mean m and variance v . The fit accounted for 97.4% of the
292 total variance in the data. Due to the limitations of image resolution, sample size and
293 uniformity, we only calculated soil pores with diameters ranging from 20 to 500 μm .
294 While the two treatments had similar total porosity (25.4% vs. 24.8%, obtained from
295 pore size distribution by the “Thickness” plugin), the SRE-treated samples had a greater
296 proportion of smaller pores ($< 60 \mu\text{m}$) and a relatively lower portion of larger pores ($>$
297 $140 \mu\text{m}$). In the pore size range of 20-60 μm , for example, the cumulative porosities
298 were 9.73% and 7.82% for SRE-treated and water-treated samples, respectively. In the
299 pore size range of 140-500 μm , the SRE-treated samples had a cumulative porosity of
300 5.53%, slightly lower than that (6.94%) of the water-treated samples. Thus, SRE
301 incubation significantly altered soil pore-size distribution.

302 **Discussion**

303 Our results support earlier findings that in general, addition of synthetic or natural
304 root exudates increases soil strength (Gao *et al.*, 2017), reduces unsaturated soil
305 hydraulic conductivity (Zheng *et al.*, 2018) and soil water evaporation (Choudhury *et*
306 *al.*, 2018). These changes may relate to the properties of the SRE solution *per se*, and
307 SRE-induced changes in microbial activity that have transformed soil structure. We
308 demonstrated that the changes of soil physical properties and processes induced by the
309 synthetic root exudate depended on soil water status. Comparing with the control

310 samples (augmented with water), the SRE-treated samples have greater hydraulic
311 conductivities at higher water contents (i.e., the flat section of $k(\theta_v)$ curve), and a
312 relatively higher stage III evaporation rate. To the best of our knowledge, this is the
313 first report that SRE treatment causes an increase of soil thermal conductivity.

314 **Root exudates as drivers of microbial activities that promote soil structure** 315 **formation**

316 It was possible that enhanced microbial activity due to SRE incubation was
317 responsible for the modification of soil structure and pore-size distribution (i.e., a
318 greater fraction of smaller pores and a relatively lower fraction of larger pores, Figure
319 5) and the subsequent changes of soil physical properties. Firstly, root exudates are
320 sources of carbon and energy for the heterotrophic soil microflora (Morel *et al.*, 1991).
321 Soil microbial activity is usually stimulated by root-released carbon, either in the form
322 of natural root exudate (Paterson *et al.*, 2007) or artificially prepared materials
323 (Choudhury *et al.*, 2018). Several studies have shown that in sandy soils, the hyphae
324 network is mainly responsible for stabilizing soil particles into aggregates (Degens &
325 Sparling, 1995; Degens *et al.*, 1996) by cross-linkage and entanglement of particles
326 (Oades, 1993; Degens *et al.*, 1996; Moreno-Espíndola *et al.*, 2007; Tisdall *et al.*, 2012).
327 In this study, the addition of SRE almost certainly promoted microbial activity (see Gao
328 *et al.*, 2017; Choudhury *et al.*, 2018), which enhanced the soil particle-to-particle
329 contacts and led to greater soil PR and λ values (Figures 1 and 2). For SRE-treated
330 samples, we observed that hyphae appeared on the 2nd day of incubation (at 18°C), grew
331 prolifically on the 4th - 5th days, and some dead or dry hyphae at the end of 7-days

332 incubation, while no hyphae could be seen on the water-treated samples. By using the
333 same SRE and similar incubation experiments, Gao *et al.* (2017) found increased
334 numbers of several kinds of bacteria. They further showed that suppression of bacteria
335 was more effective in increasing soil strength than suppression of fungi, suggesting the
336 important role of fungi in shaping soil structure.

337 Other microorganisms might also have contributed to the structural changes of
338 SRE-treated samples. There are reports that bacteria and their metabolites can improve
339 soil stability by enhancing particle-to-particle adhesion, and affect water retention and
340 reduce soil water diffusivity by blocking smaller pores and altering surface tension of
341 water (Or *et al.*, 2007; Benard *et al.*, 2017; Papadakos *et al.*, 2017; Choudhury *et al.*
342 2018).

343 **Synthetic root exudates as a bonding agent**

344 Root exudates usually consist of a group of organic compounds (e.g.,
345 polysaccharides, organic acids, glucose, and sugars) that are generally viscoelastic, thus
346 cause changes in soil interparticle contacts, mechanical stability (Naveed *et al.*, 2017)
347 and hydrological processes (Carminati *et al.*, 2016). The anionic forms of organic acids
348 in root exudate may disperse soil particles (Shanmuganathan & Oades, 1983), whereas
349 the sugars offset this effect by gelling soil particles together (Oades, 1984).
350 Polygalacturonic acid, a compound which can be used to simulate root exudates,
351 improved the interparticle bond energy, and as a result, the fracture toughness of clay
352 was increased exponentially with added PGA (Zhang *et al.*, 2008). In addition, some
353 compounds in root exudates are powerful surfactants, which decrease the surface

354 tension at the gas-liquid surface and change soil water retention properties immediately
355 (Read *et al.*, 2003; Raaijmakers *et al.*, 2010).

356 It is hardly possible to differentiate the bonding effects of the SRE from microbial
357 induced soil structural changes. For this purpose, we compared the PR and λ values in
358 this study against measurements from another experiment conducted at 4°C, in which
359 microbial activity was inhibited, and the changes of soil physical properties were
360 supposed to be caused merely by the SRE as a bonding agent. Paired (SRE-treated and
361 water-treated) samples were prepared in the same procedure except that the samples
362 were incubated at 4°C for 7 days. At the end of incubation, the SRE treatment had a
363 higher PR (974 kPa) than that of water-treated treatment (634 kPa, Figure S1). The
364 difference, however, was much smaller compared with the results from 18°C incubation
365 (1331 kPa vs. 597 kPa). Additionally, the λ values of SRE-treated samples (2.06 W m⁻¹
366 K⁻¹) showed no significant difference from that of water-treated samples incubated
367 either at 4°C (1.94 W m⁻¹ K⁻¹) or at 18°C (2.08 W m⁻¹ K⁻¹, Figure S1). Thus, the SRE
368 used in this study could have promoted bonding at low temperatures when the role of
369 microorganisms was minimized. At normal temperatures (e.g., 18°C), the direct
370 “sticking effect” of the SRE is minor comparing to its indirect function of simulating
371 microbial activity.

372 **Effects of synthetic root exudates on soil evaporation and hydraulic conductivity**

373 The influence of root exudates and similar materials (e.g., EPS produced by
374 microbes) on soil hydraulic conductivity, water retention, and evaporation have been
375 studies extensively. In general, the addition of root exudates and EPS leads to a

376 reduction of hydraulic conductivity (Vandevivere & Baveye, 1992; Or *et al.*, 2007),
377 increase of water retention, and a decline of evaporative water loss (Chenu &
378 Roberson, 1996; Choudhury *et al.*, 2018). These phenomena are explained by the
379 experimental evidence that these materials have a larger water holding capacity, alter
380 soil matrix structure and connectivity of pore space, and modify the surface tension and
381 viscosity of soil water (Naveed *et al.*, 2018; Zheng *et al.*, 2018). Our results support the
382 previous findings in that the SRE-treated samples had lower average evaporation rates
383 and reduced cumulative evaporative water losses than the water-treated samples (Figure
384 3). However, the two treatments differed considerably in terms of duration and rate of
385 evaporation at the three stages. During stage I evaporation, both treatments had a
386 relatively high and constant evaporation rate controlled by atmospheric conditions. Yet
387 the SRE-treated samples had a shorter stage I period, since the soils were not able to
388 sustain liquid water flow to meet the evaporative demand due to the sudden reduction
389 of hydraulic conductivity at the inflection point (Figure 4). Additionally, due to its
390 greater water holding capacity and lower hydraulic conductivity than that of the water-
391 treated samples, the SRE-treated samples maintained a longer but slower water loss rate
392 in stage II evaporation. It was surprising that the SRE-treated samples had a relatively
393 higher rate of stage III evaporation than the water treated ones, which has not been
394 reported previously. This is likely to be because the SRE-treated samples maintained
395 higher water contents (mostly absorbed water) at the end of stage II evaporation.

396 Interestingly, our data showed that addition of the SRE did not always reduce soil
397 hydraulic conductivity. Comparing to the water-treated samples, elevated hydraulic

398 conductivities were obtained in the SRE-treated ones in the flat section of the $k(\theta)$ curve
399 (Figure 4). We are not clear about the explanation of this phenomena because both
400 treatments had similar total porosities, and the SRE treatment possessed a greater
401 proportion of smaller pores (Figures 5 and 6). A potential explanation is that some
402 preferential flow paths had been created due to SRE-induced microbial activities.

403 **Conclusions**

404 In this study, we investigated the mechanical strength (penetrometer resistance),
405 thermal conductivity, hydraulic conductivity and evaporation process of a sandy soil
406 treated with a synthetic root exudate. After incubation at 18°C, both penetrometer
407 resistance and thermal conductivity were increased, soil hydraulic conductivity was
408 increased in the wet region but decreased in the dry region, and overall evaporation rate
409 and cumulative water loss were reduced significantly, which were attributed to
410 reinforced particle-to-particle contacts and changes of soil pore-size distribution due
411 mainly to exudate-stimulated microbial activity. X-ray CT scanning images provided
412 direct evidence that root exudates treatment had little effect on total porosity but
413 increased the number of smaller pores. Further studies are required to examine if these
414 conclusions also apply to field soils with natural root exudates.

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423 Centre for Monitoring the Rhizosphere’.

424 **Authors’ Contributions**

425 WZ conducted the experiment, collected the data, and drafted the article. WG,
426 WRW and TR participated in design of the work, helped in data analysis and
427 interpretation, and revised the manuscript. All authors approved the final version to be
428 published.

429 **Conflicts of Interest**

430 The authors declare that there is no conflict of interest regarding the publication of
431 this article.

432 **Data Availability**

433 The data that support the findings of this study are available from the corresponding
434 author upon reasonable request.

435 **References**

- 436 Albalasmeh, A.A. & Ghezzehei, T.A. 2014. Interplay between soil drying and root
437 exudation in rhizosphere development. *Plant and Soil*, **374**, 739–751.
- 438 Amano, E. & Salvucci, G.D. 1999. Detection and use of three signatures of soil-
439 limited evaporation. *Remote Sensing of Environment*, **67**, 108–122.
- 440 Barré, P. & Hallett, P.D. 2009. Rheological stabilization of wet soils by model root
441 and fungal exudates depends on clay mineralogy. *European Journal of Soil
442 Science*, **60**, 525–538.

443 Benard, P., Zarebanadkouki, M., Hedwig, C., Holz, M., Ahmed, M.A. & Carminati,
444 A. 2017. Pore-scale distribution of mucilage affecting water repellency in the
445 rhizosphere. *Vadose Zone Journal*, **17**, 1–9.

446 Bengough, A.G., Campbell, D. & O’Sullivan, M. 2000. Penetrometer Techniques
447 in Relation to Soil Compaction and Root Growth. In: *In: Smith, K.A., Mullins,*
448 *C.E. (Eds.), Soil and Environmental Analysis: Physical Methods, Revised, and*
449 *Expanded, 2nd ed. Marcel Dekker, New York, pp. 377–403.*

450 Bristow, K.L., Bilskie, J.R., Kluitenberg, G.J. & Horton, R. 1995. Comparison of
451 techniques for extracting soil thermal properties from dual-probe heat-pulse data.
452 *Soil Science*, **160**, 1–7.

453 Bristow, K.L., Kluitenberg, G.J. & Horton, R. 1994. Measurement of soil thermal
454 properties with a dual-probe heat-pulse technique. *Soil Science Society of*
455 *America Journal*, **58**, 1288–1294.

456 Carminati, A., Moradi, A.B., Vetterlein, D., Vontobel, P., Lehmann, E., Weller, U.,
457 Vogel, H.J. & Oswald, S.E. 2010. Dynamics of soil water content in the
458 rhizosphere. *Plant and Soil*, **332**, 163–176.

459 Carminati, A., Zarebanadkouki, M., Kroener, E., Ahmed, M.A. & Holz, M. 2016.
460 Biophysical rhizosphere processes affecting root water uptake. *Annals of Botany*,
461 **118**, 561–571.

462 Chenu, C. & Roberson, E.B. 1996. Diffusion of glucose in microbial extracellular
463 polysaccharide as affected by water potential. *Soil Biology and Biochemistry*, **28**,
464 877–884.

465 Choudhury, B.U., Ferraris, S., Ashton, R.W., Powlson, D.S. & Whalley, W.R. 2018.
466 The effect of microbial activity on soil water diffusivity. *European Journal of*
467 *Soil Science*, **69**, 407–413.

468 Czarnes, S., Hallett, P.D., Bengough, A.G. & Young, I.M. 2000. Root- and microbial-
469 derived mucilages affect soil structure and water transport. *European Journal of*
470 *Soil Science*, **51**, 435–443.

471 Degens, B.P. & Sparling, G.P. 1995. Repeated wet-dry cycles do not accelerate the
472 mineralization of organic C involved in the macro-aggregation of a sandy loam
473 soil. *Plant and Soil*, **175**, 197–203.

474 Degens, B.P., Sparling, G.P. & Abbott, L.K. 1996. Increasing the length of hyphae in
475 a sandy soil increases the amount of water-stable aggregates. *Applied Soil*
476 *Ecology*, **3**, 149–159.

477 Dunbabin, V.M., McDermott, S. & Bengough, A.G. 2006. Upscaling from
478 rhizosphere to whole root system: Modelling the effects of phospholipid
479 surfactants on water and nutrient uptake. *Plant and Soil*, **283**, 57–72.

480 Farouki, O.T. 1986. *Thermal properties of soils. Series on rock and soil mechanics*
481 *series*.

482 Galloway, A.F., Knox, P. & Krause, K. 2020. Sticky mucilages and exudates of
483 plants: putative microenvironmental design elements with biotechnological
484 value. *New Phytologist*, **225**, 1461–1469.

485 Gao, W., Muñoz-Romero, V., Ren, T., Ashton, R.W., Morin, M., Clark, I.M.,
486 Powlson, D.S. & Whalley, W.R. 2017. Effect of microbial activity on

487 penetrometer resistance and elastic modulus of soil at different temperatures.
488 *European Journal of Soil Science*, **68**, 412–419.

489 Gao, W., Ren, T., Bengough, A.G., Auneau, L., Watts, C.W. & Whalley, W.R. 2012.
490 Predicting Penetrometer Resistance from the Compression Characteristic of Soil.
491 *Soil Science Society of America Journal*, **76**, 361–369.

492 Gregory, P.J. 2006. Roots, rhizosphere and soil: The route to a better understanding of
493 soil science? *European Journal of Soil Science*, **57**, 2–12.

494 Hinsinger, P., Bengough, A.G., Vetterlein, D. & Young, I.M. 2009. Rhizosphere:
495 Biophysics, biogeochemistry and ecological relevance. *Plant and Soil*, **321**, 117–
496 152.

497 Holz, M., Carminati, A. & Kuzyakov, Y. 2015. Distribution of root exudates and
498 mucilage in the rhizosphere: combining ¹⁴C imaging with neutron
499 radiography. **17**, 2015.

500 Horn, R. 1994. The Effect of Aggregation of Soils on Water, Gas, and Heat Transport.
501 In: *Flux Control in Biological Systems*, pp. 335–361. Elsevier.

502 Lambers, H., Mougél, C., Jaillard, B. & Hinsinger, P. 2009. Plant-microbe-soil
503 interactions in the rhizosphere: An evolutionary perspective. *Plant and Soil*, **321**,
504 83–115.

505 Lavelle, P. 2002. Functional domains in soils. *Ecological Research*, **17**, 441–450.

506 Moradi, A.B., Carmina, A., Lamparter, A., Woche, S.K., Bachmann, J., Vetterlein, D.,
507 Vogel, H.-J. & E., S.O. 2012. Is the Rhizosphere Temporarily Water Repellent?
508 *Vadose Zone Journal*, **11**.

509 Moraes, M., Debiiasi, H., Franchini, J., Bonetti, J., Levien, R., Schnepf, A. & Leitner.
510 2019. Mechanical and Hydric Stress Effects on Maize Root System Development
511 at Different Soil Compaction Levels. *Frontiers in Plant Science*, **10**, 1–18.

512 Morel, J.L., Habib, L., Plantureux, S. & Guckert, A. 1991. Influence of maize root
513 mucilage on soil aggregate stability. *Plant and Soil*, **136**, 111–119.

514 Moreno-Espíndola, I.P., Rivera-Becerril, F., de Jesús Ferrara-Guerrero, M. & De
515 León-González, F. 2007. Role of root-hairs and hyphae in adhesion of sand
516 particles. *Soil Biology and Biochemistry*, **39**, 2520–2526.

517 Nakanishi, T.M., Okuni, Y., Hayashi, Y. & Nishiyama, H. 2005. Water gradient
518 profiles at bean plant roots determined by neutron beam analysis. *Journal of*
519 *Radioanalytical and Nuclear Chemistry*, **264**, 313–317.

520 Nambiar, E.K.S. 1976. Uptake of Zn⁶⁵ from dry soil by plants. *Plant and Soil*, **44**,
521 267–271.

522 Naveed, M., Brown, L.K., Raffan, A.C., George, T.S., Bengough, A.G., Roose, T.,
523 Sinclair, I., Koebernick, N., Cooper, L., Hackett, C.A. & Hallett, P.D. 2017.
524 Plant exudates may stabilize or weaken soil depending on species, origin and
525 time. *European Journal of Soil Science*, **68**, 806–816.

526 Naveed, M., Brown, L.K., Raffan, A.C., George, T.S., Bengough, A.G., Roose, T.,
527 Sinclair, I., Koebernick, N., Cooper, L. & Hallett, P.D. 2018. Rhizosphere-scale
528 quantification of hydraulic and mechanical properties of soil impacted by root
529 and seed exudates. *Vadose Zone Journal*, **17**.

530 Oades, J.M. 1984. Soil organic matter and structural stability: mechanisms and

531 implications for management. *Plant and Soil*, **76**, 319–337.

532 Oades, J.M. 1993. The role of biology in the formation, stabilization and degradation
533 of soil structure. *Geoderma*, **56**, 377–400.

534 Or, D., Phutane, S. & Dechesne, A. 2007. Extracellular polymeric substances
535 affecting pore-scale hydrologic conditions for bacterial activity in unsaturated
536 soils. *Vadose Zone Journal*, **6**, 298–305.

537 Papadakos, G.N., Karangelos, D.J., Petropoulos, N.P., Anagnostakis, M.J., Hinis, E.P.
538 & Simopoulos, S.E. 2017. Uncertainty assessment method for the Cs-137 fallout
539 inventory and penetration depth. *Journal of Environmental Radioactivity*, **171**,
540 234–245.

541 Paterson, E., Gebbing, T., Abel, C., Sim, A. & Telfer, G. 2007. Rhizodeposition
542 shapes rhizosphere microbial community structure in organic soil. *New*
543 *Phytologist*, **173**, 600–610.

544 Raaijmakers, J.M., de Bruijn, I., Nybroe, O. & Ongena, M. 2010. Natural functions of
545 lipopeptides from *Bacillus* and *Pseudomonas*: More than surfactants and
546 antibiotics. *FEMS Microbiology Reviews*, **34**, 1037–1062.

547 Read, D.B., Bengough, A.G., Gregory, P.J., Crawford, J.W., Robinson, D.,
548 Scrimgeour, C.M., Young, I.M., Zhang, K. & Zhang, X. 2003. Plant roots release
549 phospholipid surfactants that modify the physical and chemical properties of soil.
550 *New Phytologist*, **157**, 315–326.

551 Read, D.B., Gregory, P.J. & Bell, A.E. 1999. Physical properties of axenic maize root
552 mucilage. *Plant and Soil*, **211**, 87–91.

553 Sasse, J., Martinoia, E. & Northen, T. 2018. Feed Your Friends: Do Plant Exudates
554 Shape the Root Microbiome? *Trends in Plant Science*, **23**, 25–41.

555 Shanmuganathan, R.T. & Oades, J.M. 1983. Modification of soil physical properties
556 by addition of calcium compounds. *Australian Journal of Soil Research*, **21**,
557 285–300.

558 Shi, S., Richardson, A.E., O’Callaghan, M., Deangelis, K.M., Jones, E.E., Stewart, A.,
559 Firestone, M.K. & Condon, L.M. 2011. Effects of selected root exudate
560 components on soil bacterial communities. *FEMS Microbiology Ecology*, **77**,
561 600–610.

562 Strayer, D.L., Power, M.E., Fagan, W.F., Pickett, S.T.A. & Belnap, J. 2003. A
563 Classification of Ecological Boundaries. *BioScience*, **53**, 723–729.

564 Suleiman, A.A. & Ritchie, J.T. 2003. Modeling soil water redistribution during
565 second-stage evaporation. *Soil Science Society of America Journal*, **67**, 377–386.

566 Tisdall, J.M. 1991. Fungal hyphae and structural stability of soil. *Australian Journal*
567 *of Soil Research*, **29**, 729–743.

568 Tisdall, J.M., Nelson, S.E., Wilkinson, K.G., Smith, S.E. & McKenzie, B.M. 2012.
569 Stabilisation of soil against wind erosion by six saprotrophic fungi. *Soil Biology*
570 *and Biochemistry*, **50**, 134–141.

571 Traoré, O., Groleau-Renaud, V., Plantureux, S., Tubeileh, A. & Boeuf-Tremblay, V.
572 2000. Effect of root mucilage and modelled root exudates on soil structure.
573 *European Journal of Soil Science*, **51**, 575–581.

574 Vandevivere, P. 1995. Bacterial clogging of porous media: A new modelling

575 approach. *Biofouling*, **8**, 281–291.

576 Vandevivere, P. & Baveye, P. 1992. Relationship between transport of bacteria and
577 their clogging efficiency in sand columns. *Applied and Environmental*
578 *Microbiology*, **58**, 2523–2530.

579 Vaz, C.M.P., Manieri, J.M., de Maria, I.C. & Tuller, M. 2011. Modeling and
580 correction of soil penetration resistance for varying soil water content.
581 *Geoderma*, **166**, 92–101, (At:
582 <http://dx.doi.org/10.1016/j.geoderma.2011.07.016>).

583 Watt, M., McCully, M.E. & Canny, M.J. 1994. Formation and stabilization of
584 rhizosheaths of *Zea mays* L.: Effect of soil water content. *Plant Physiology*, **106**,
585 179–186.

586 Watt, M., Silk, W.K. & Passioura, J.B. 2006. Rates of root and organism growth, soil
587 conditions, and temporal and spatial development of the rhizosphere. *Annals of*
588 *Botany*, **97**, 839–855.

589 Whalley, W.R., Riseley, B., Leeds-Harrison, P.B., Bird, N.R.A., Leech, P.K. &
590 Adderley, W.P. 2005. Structural differences between bulk and rhizosphere soil.
591 *European Journal of Soil Science*, **56**, 353–360.

592 Young, I.M. 1995. Variation in moisture contents between bulk soil and the
593 rhizosheath of wheat (*Triticum aestivum* L. cv. Wembley). *New Phytologist*, **130**,
594 135–139.

595 Zhang, B., Hallett, P.D. & Zhang, G. 2008. Increase in the fracture toughness and
596 bond energy of clay by a root exudate. *European Journal of Soil Science*, **59**,

597 855–862.

598 Zheng, W., Zeng, S., Bais, H., LaManna, J.M., Hussey, D.S., Jacobson, D.L. & Jin, Y.

599 2018. Plant Growth-Promoting Rhizobacteria (PGPR) Reduce Evaporation and

600 Increase Soil Water Retention. *Water Resources Research*, **54**, 3673–3687.

601 Zhou, H., Peng, X., Peth, S. & Xiao, T.Q. 2012. Effects of vegetation restoration on

602 soil aggregate microstructure quantified with synchrotron-based micro-computed

603 tomography. *Soil and Tillage Research*, **124**, 17–23.

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605

606 **FIGURE CAPTIONS**

607 **Figure 1.** Dynamics of penetrometer resistance of synthetic root exudate (SRE)- and
608 water-treated soil samples during a drying process following incubation at 18°C.

609 **Figure 2.** Dynamics of thermal conductivity of synthetic root exudate (SRE)- and
610 water- treated soil samples during a drying period following incubation at 18°C.

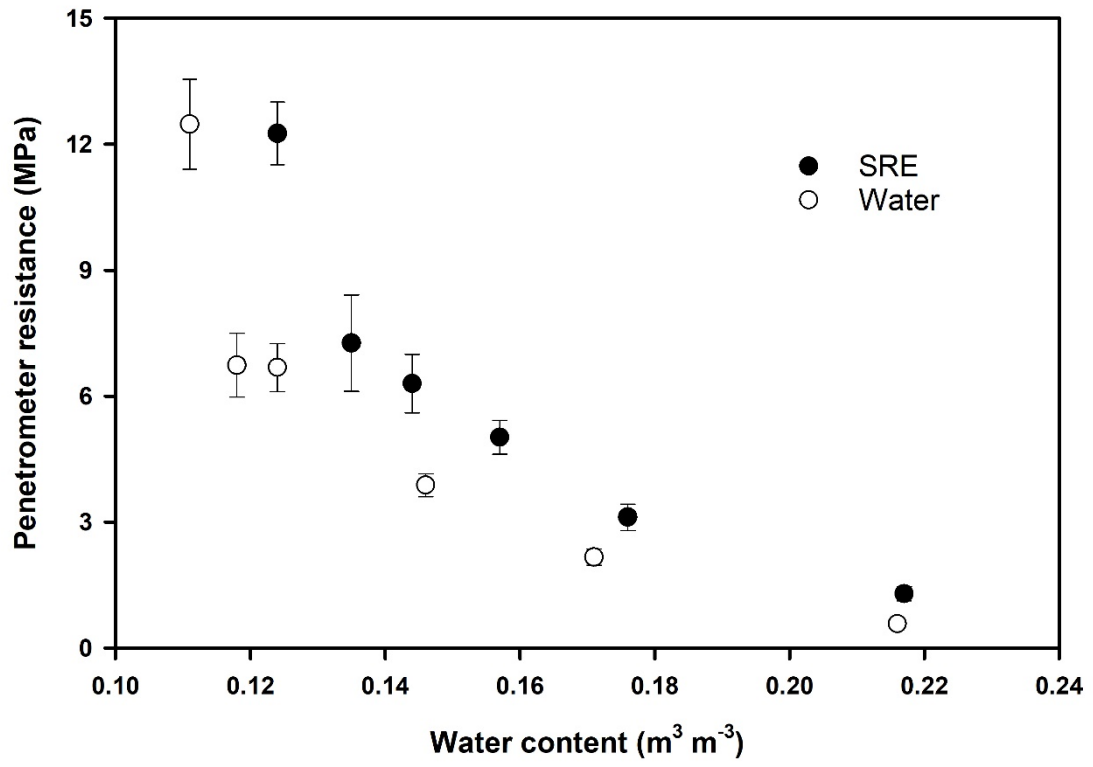
611 **Figure 3.** Changes of soil water evaporation rates (a) and cumulative evaporation (b)
612 of synthetic root exudate (SRE)- and water-treated samples during a drying process.

613 The samples were incubation at 18°C for 7 days, saturated with water, and then dried
614 at room temperature.

615 **Figure 4.** Soil hydraulic conductivity as a function of water content for synthetic root
616 exudate (SRE)- and water-treated samples obtained with the ku-pF apparatus.

617 **Figure 5.** Grayscale and binary images of synthetic root exudate (SRE)- and water-
618 treated soil samples following incubation at 18°C for 7 days. The images were
619 obtained using an X-ray μ -CT scanner with a 10 μ m-resolution. Images are displayed
620 as 1100×1100 pixels.

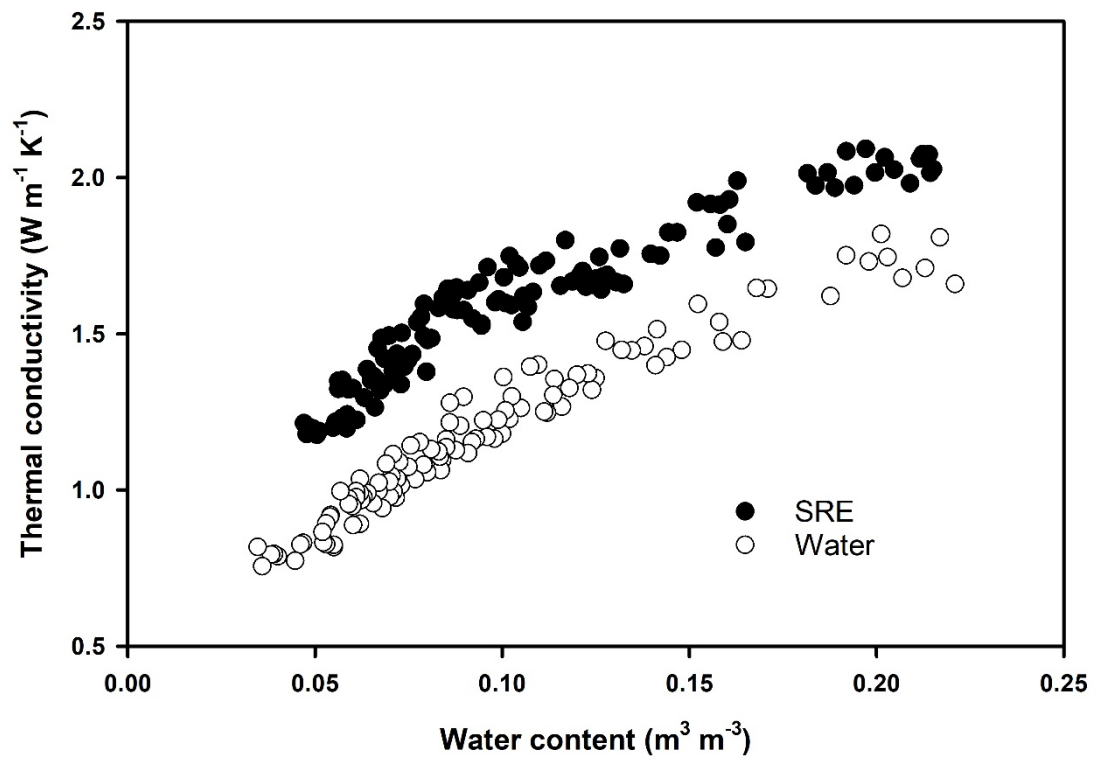
621 **Figure 6.** Pore-size distribution of synthetic root exudate (SRE)- and water-treated
622 soil samples following incubation at 18°C for 7 days. The porosity data were
623 estimated indirectly from X-ray CT scanned images. The values of the X-axis
624 represent logarithmic results of pore diameters.



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Figure 1



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Figure 2

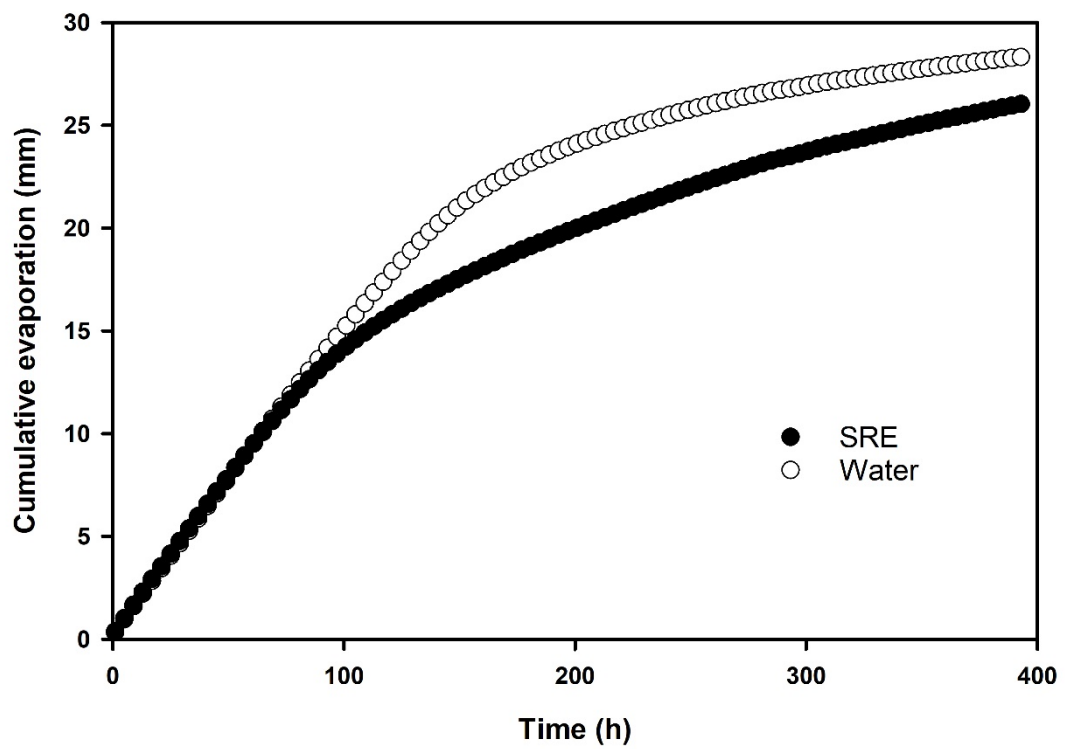
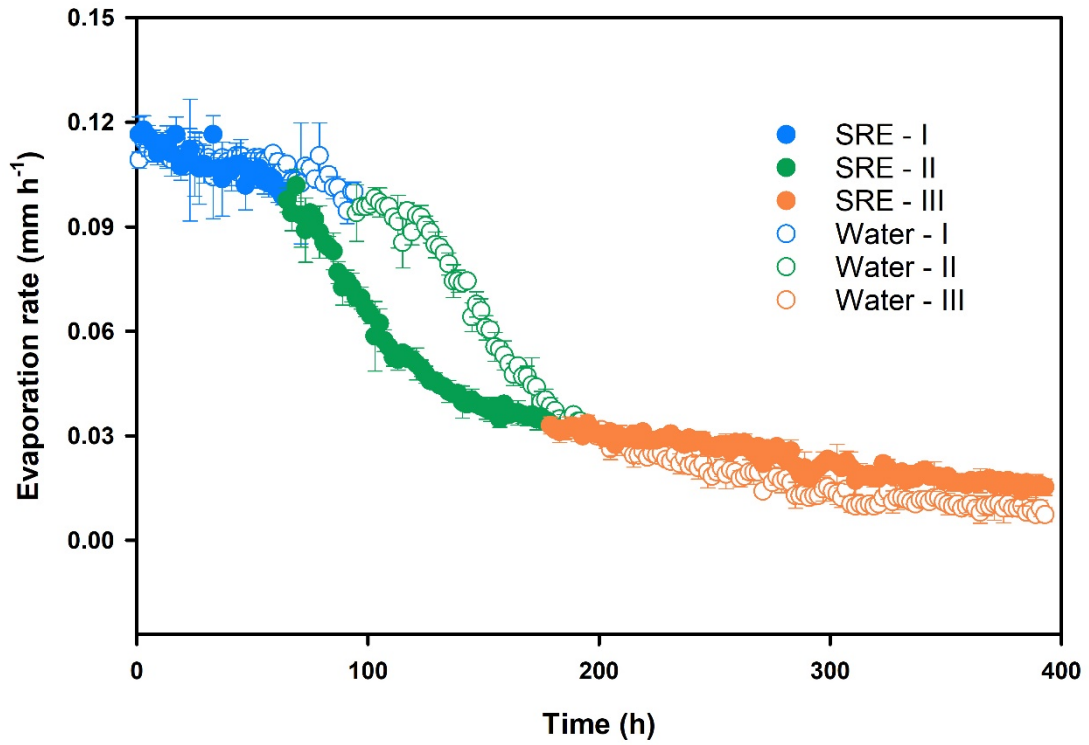
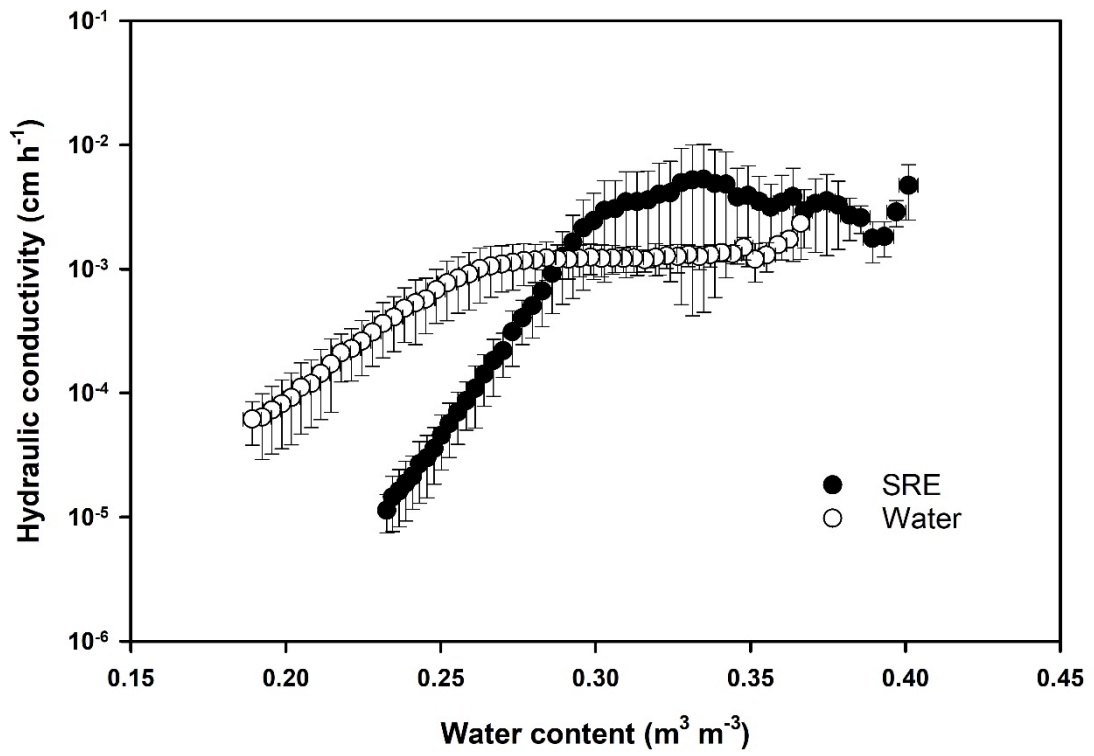


Figure 3

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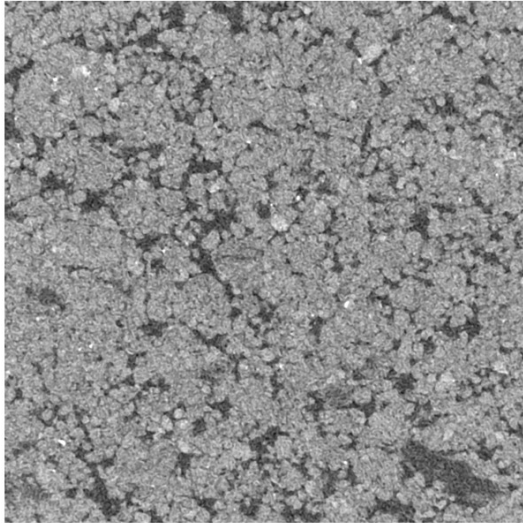


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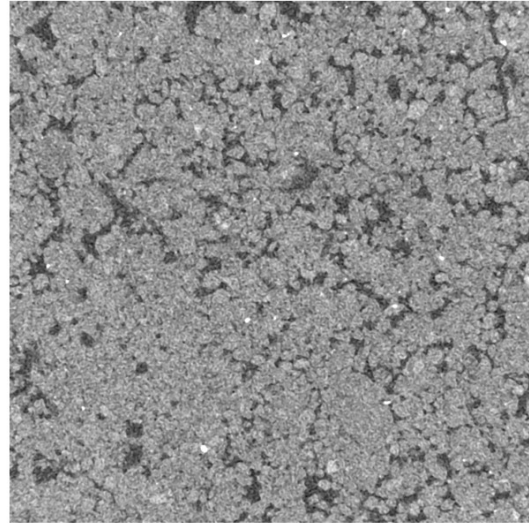
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Figure 4

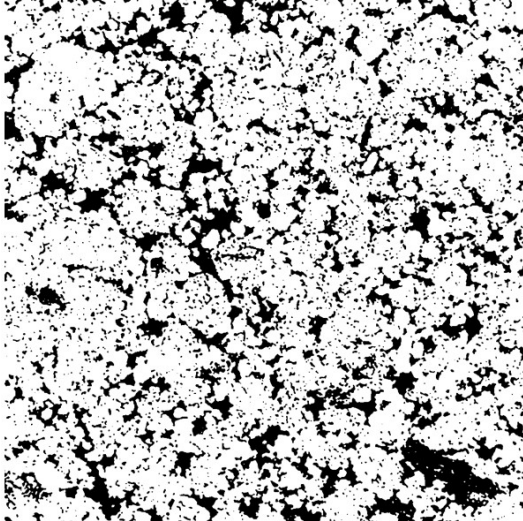
Water sample – Grayscale image



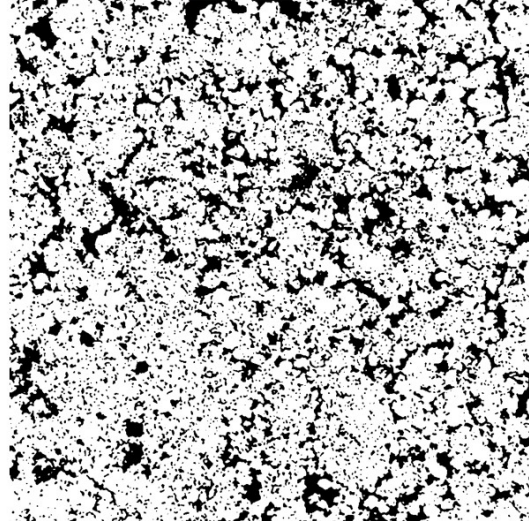
SRE sample – Grayscale image



Water sample – Binary image



SRE sample – Binary image

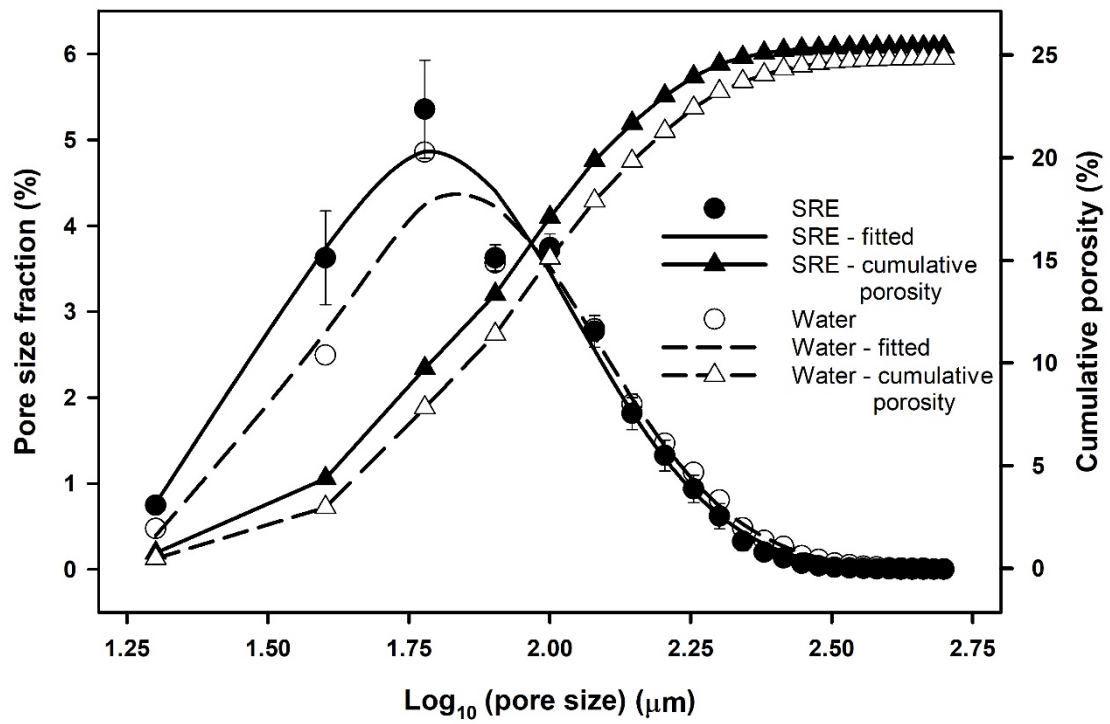


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Figure 5



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Figure 6

