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14 Abstract

While soil microbial ecology, soil organic carbon (SOC) and soil physical quality are widely understood to be interrelated — the underlying drivers of emergent properties, from land management to biochemistry, are hotly debated. Biological binding agents, microbial exudates, or 'extracellular polymeric substances' (EPS) in soil are now receiving increased attention due to several of the existing methodological challenges having been overcome. We applied a recently developed approach to quantify soil EPS, as extracellular protein and extracellular polysaccharide, on the well-characterised soils of the Highfield Experiment, Rothamsted Research, UK. Our aim was to investigate the links between agricultural land use, SOC, transient binding agents known as EPS, and their impacts on soil physical quality (given by mean weight diameter of water stable aggregates; MWD). We compared the legacy effects from long-term previous land-uses (unfertilised grassland, fertilised arable, and fallow) which were established >50 years prior to investigation, crossed with the same current land-uses

established for a duration of only 2.5 years prior to sampling. Continuously fallow and grassland soils represented the poorest and greatest states of structural integrity, respectively. Total SOC and %N were found to be affected by both previous and current land-uses, while extractable EPS and MWD were driven primarily by the current land-use. Land-use change between these two extremes (fallow→grass; grass→fallow) resulted in smaller SOC differences (64% increase or 37% loss) compared to MWD (125% increase or 78% loss). SOC concentration correlated well to MWD (adjusted  $R^2 = 0.72$ ) but the high SOC content from previous grassland was not found to contribute directly to the current stability (p < 0.05). Our work thus supports the view that certain distinct components of SOC, rather than the total pool, have disproportionately important effects on a soil's structural stability. EPS-protein was more closely related to aggregate stability than EPS-polysaccharide (p values of 0.002 and 0.027, respectively), and ranking soils with the 5 highest concentrations of EPS-protein to their corresponding orders of stability (MWD) resulted in a perfect match. We confirmed that both EPS-protein and EPS-polysaccharide were transient fractions: supporting the founding models for aggregate formation. We suggest that management of transient binding agents such as EPS —as opposed to simply increasing the total SOC content— may be a more feasible strategy to improve soil structural integrity and help achieve environmental objectives.

44 Keywords

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- Transient binding agents; microbial exudates; protein matrix bonding; exopolysaccharide;
- water stable aggregate (MWD); cation exchange resin (CER)

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#### 1. Introduction

## 1.1. Land use, total SOC and soil structural stability

It is well established that surface vegetation, as controlled by how land is managed, has profound interactions with the underlying soil. In this way, land-use affects many of a soil's fundamental properties including soil organic matter, and its primary constituent: soil organic C (SOC; Jobbágy & Jackson, 2000). Managements that increase SOC in loamy soils enable the development of microporous networks, which are more conducive to the transmission of nutrients, water and air (Bacq-Labreuil et al., 2018; Guo et al., 2020). Without stability in soil structure, these networks and aggregates collapse: restricting porosity, connectivity, nutrient transport, infiltration and root penetration. Soils under arable crops are subjected to annual cultivation and most of the above-ground biomass is harvested, compared to soils under permanent vegetative cover, which have no such disturbance and benefit from increased perennial inputs of C (Chapman et al., 2013; Wiesmeier et al., 2012). It follows that a change in land use alters the balance between inputs and outputs and SOC adjusts to a new equilibrium accordingly (Gregory et al., 2016; Johnston et al., 2009). Soil typically loses SOC faster than it is gained (Johnston et al., 2009; Poeplau et al., 2011; Jones et al., 2016) and so international efforts to increase SOC, such as the '4-per-mille' initiative (Chabbi et al., 2017) are challenged by high frequencies in land-use change.

Alternative policies have been called for which focus more on improving soil physical quality (Poulton et al., 2018) and understanding microbial processes contributing to C storage (Liang, et al., 2017). The water-stability and size distribution of soil aggregates, as described by their mean weight diameter (MWD) reflects the stability against physical stresses typically imposed *in-situ* (Le Bissonais, 1996). Accordingly, MWD is commonly used to build an index of soil health and/or quality. Regression models have gone as far as to explain aggregate stability by simple SOC content (e.g. Kemper and Koch, 1966; Six et al., 2002; Soinne et al., 2016) but the relationships between SOC and aggregate stability are not straightforward (Denef and Six, 2005).

#### 1.2 Organic binding agents

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Both the quality and location of SOC, or more specifically 'binding agents' at the microscale are understood to be important factors determining the overall stability of a soil (Denef et al., 2007; Jensen et al., 2019). The seminal work of Tisdall and Oades (1982) was perhaps the early thesis on this. They proposed an aggregate hierarchy where smaller microaggregates, held together by persistent binding agents (organo-mineral associations), or transient binding agents (microbial exopolysaccharides) combine within macroaggregates and are stabilised by temporary binding agents such as roots and fungal hyphae. Whilst persistent agents were thought to be largely regulated by soil mineralogy and particle size, both the temporary and transient agents were proposed to be controlled by land-use (Tisdall and Oades, 1982). Golchin et al. (1994) further developed the theory to include microbiallydependent cycles of aggregate formation and dispersal, commencing with organic matter inputs from plant cover which stimulate microbial production of transient binding agents. This in turn facilitated the adsorption of aligned clay particles, by electrostatic attraction, forming central units of new stable aggregates. When readily-available C became depleted, their model predicted binding agents would no longer be produced and the aggregates would crumble when subjected to physical stress. Soil biology has since been shown to be a major contributor to soil aggregate stability (Watts et al., 2001; 2005; Jensen et al., 2019), which is highly responsive to management (Kabir, 2005) and especially C inputs (Hirsch et al., 2017).

## 1.3 Extracellular polymeric substances (EPS)

The production of exudates as biological binding agents is a salient feature of individual microorganisms attached to their physical habitat or co-habiting in groups called biofilms (Flemming and Wingender, 2010). This exuded matrix is composed of biopolymers more widely referred to as extracellular polymeric substances or 'EPS', a term we use hereafter. EPS is thought to be composed primarily of polysaccharides but proteins are a crucial functional component (Flemming et al., 2007). Evidence from water sciences shows that protein-rich EPS plays a central role in adhesion and stabilisation of microbial aggregates

(Guo et al., 2016). Indeed, *ex-situ* additions of bacteria selected for their propensity to exude proteinaceous EPS (e.g. *Pseudomonas spp.*) have been shown to aggregate soil particles in solution (Caesar-Tonthat et al., 2014; Krause et al., 2019). While extracellular polysaccharides have long been assumed to be important for soil structure, studies of EPS in soil have been held back by methodological challenges. For example, in the unique study of Tang et al., (2011) the addition of selective fungicides and bactericides were both shown to decrease soil aggregate stability but the methods used to measure extracellular polysaccharides failed to uncover any statistically significant link. Redmile-Gordon et al. (2014) investigated the 'hot sulphuric acid' method used in the study of Tang et al. (2011) and found that the method also co-extracted large amounts of intracellular biomass, and nonspecific extracellular SOC – likely confounding all three classes of biological binding agent. Other investigators have attempted to use hot water as an extractant but conceded that hot water was also not selective for extracellular polysaccharides (Marchus et al., 2018).

Related work on the proteinaceous exudates of arbuscular mychorrhizal fungi, named glomalin, or glomalin-related soil protein (GRSP) are believed to impart aggregate stability and C storage (Wright and Upadhyaya, 1996; Bach et al., 2010). However, the autoclaving step precludes any claim that these extracts represent extracellular material. Furthermore, extracts of GRSP were shown to contain more proteins of bacterial than fungal origin (Gillespie et al., 2011). Redmile-Gordon et al. (2013) proposed that it might be more practical to measure changes in pools of categorically extracellular biopolymers as a biophysically distinct product of the microbial biomass, rather than attempting to ascribe taxonomic origin to materials that could be either intracellular or extracellular. The present lack of studies on EPS, linking soil structural stability, or the effects of land-use change represents an opportunity for researchers to advance our understanding of sustainable soil management in this rapidly expanding area (Costa et al., 2018; Liang, et al., 2017).

## 1.4 Methodological developments for extracting EPS

The qualitative challenges described above suggest a priority for methods that do not co-extract large amounts of 'non-target' organic matter (intracellular or extracellular). The only method reported to meet these criteria to date is the 'cation exchange resin' (CER) method initially developed by Frolund et al. (1996) and adapted for application in soil by Redmile-Gordon et al. (2014). In the method for soils, a dilute divalent metal salt (CaCl<sub>2</sub>) is used to remove the easily soluble organic fraction. This fraction is highly variable in soil depending on rainfall and moisture content immediately prior to sampling. It is important that the preliminary extractant is not monovalent to maintain EPS stability prior to subsequent removal using CER. The resin extraction step works by reducing binding between negatively charged moieties of the polymeric matrix that are otherwise bridged by divalent cations and releases shorter chains into the extraction buffer (Sheng et al., 2010). This same method was also found to be suitable for EPS in soils dominated by trivalent metal cations (Wang et al., 2019).

The extracellular specificity of the soil EPS extraction method was confirmed using measurements of soil microbial adenosine 5'-triphosphate (ATP; Redmile-Gordon et al., 2014). Their findings concurred with investigators in aquatic sciences (e.g. Takahashi et al., 2009; Ge et al., 2010) that cell-lysis during CER extraction was minimal. To address concerns regarding extracellular contamination, Redmile-Gordon et al. (2015) traced stable isotopes of N through the microbial biomass, showing that CER preferentially extracted synthesised EPS over extracellular SOC of more dubious origin. Colorimetric measurements of EPS protein and polysaccharide in the soil extracts were also corroborated by GC-MS measurements of volatile derivatives, adding support to the more affordable colorimetric determination methods that are normally applied.

## 1.5 Aims and objectives

Our objectives were to investigate the effect of land use or management changes on soil EPS at the field scale and relate these findings to concomitant changes in soil aggregate stability as a fundamental soil physical property. Importantly, the combinations of 'previous'

and 'current' land-uses in this long-term experiment would provide a contrast in the proportion of transient binding agents relative to the total organic matter pool: with 'previous' and 'current' each representing a deficit, and abundance, of land-use-driven binding agents, respectively. Accordingly, we tested three hypotheses: (1) The mixed ley/grass land-use would support the greatest production of microbial EPS, followed by arable, and fallow managements, respectively. (2) EPS concentrations would be more affected by short-term application of the most conducive management than by 'legacy effects' from previous managements – even when that previous management had been applied continuously for a duration of over 50 years. (3) These same EPS concentrations would be significantly correlated to an indicator of soil physical quality *i.e.* aggregate stability.

## 2. Material and methods

## 2.1 Experimental site

This study was conducted on the Highfield experiment at Rothamsted Research, UK (Figure 1; global coordinates 51.804, -0.363) where contrasting land-uses were established previously (duration >50 years) and recently (2.5 years prior to sampling). There is an annual mean temperature (1992–2014) and rainfall (1981–2010) of 10.2 °C and 718 mm, respectively (Scott et al., 2014). The soil is classified as a stagnogleyic paleo-argillic brown earth (Batcombe series), equivalent to a Chromic Luvisol (IUSS Working Group WRB, 2015) with silty clay loam texture (Avery and Catt, 1995). Mineralogy is dominated by expanding 2:1 clays (randomly interstratified montmorillonite and vermiculite). Mineralogy and particle size did not differ significantly between treatments allowing the effect of contrasting land-use to be examined without confounding effects of mineralogy or particle size (Jensen et al., 2019). [Suggested position for Table 1 (soil properties) and Figure 1 (experimental site plan)]

Long-term ley-arable and bare fallow experiments at the site provided 'previous' grassland and arable (both since 1949, in 50 × 7 m plots in a randomized complete block design with four blocks) and 'previous' bare fallow (since 1959, in adjacent land totalling *c*. 1400 m²) land-

uses, as described elsewhere (Gregory et al., 2016). The grassland management supports predominantly rye grass (*Lolium perenne* L.) and clover (*Trifolium* spp.), mown twice in summer with arable represented by winter wheat (*Triticum aestivum* L.) formerly in rotation with winter oats (*Avena sativa* L.). Hay and straw residues are removed after cutting. The fallow treatment is maintained by mouldboard ploughing and cultivating two to four times a year. Arable plots receive 220 kg N ha<sup>-1</sup> annually (primarily as ammonium nitrate, with occasional ammonium sulphate to supply crop S). Selected properties of the long-term treatments are given in Table 1.

In October 2008, three split-plots (10 × 6 m each) were established on each previous grassland and arable treatment in all four blocks, though in the present study we only focussed on the first three. Three sets of three split-plots (of the same size) were established on the adjacent fallow treatment to achieve the same replication. In the split-plots, the same three treatments (grassland, arable and fallow) were established as 'current' management treatments. Thus, the experimental design comprises 9 treatments triplicated in 27 split-plots (3 previous × 3 current management treatments × 3 field replicates). Each of the 9 treatments are therefore characterised by both a *previous* and a *current* management component. This temporal separation implicitly establishes a contrast in the abundance of management-driven transient binding agents (with transient agents being more abundant under the current managements which favour them). Hereafter we use the terms 'grass', 'arable' and 'fallow' to describe the management treatments and the symbol → to indicate where a previous management has been changed to a new current management. For example, grass→arable indicates the treatment previously under grass, but currently under arable.

## 2.2 Sampling and analysis

Soil samples were collected from three random locations in each of the 27 split-plots in February 2011 with a 75-mm diameter auger to a depth of 52 mm. To reduce within-plot variation as much as practicable, sub-samples were bulked to make a single sample per plot. A natural 3-5 mm aggregate fraction was teased from the bulk after partial-drying in a dark,

20 °C ventilated chamber for 18 h to prevent wet-smearing of aggregates upon handling. Roots and macrofauna were removed by hand. The majority of the 3-5 mm fraction was further air-dried in preparation for physical and chemical analyses while portions of the moist 3-5 mm aggregates were crumbled to pass through a 2 mm sieve and chilled to 4 °C overnight. EPS was then extracted as described by Redmile-Gordon et al. (2014). Accordingly, readily soluble organic material was removed from about 3.0 g moist soil (2.5 g dry weight equivalent of soil from each plot) using 0.01 M CaCl<sub>2</sub> at a 1:10 soil:solution ratio, in 50 mL capacity polypropylene centrifuge tubes. These tubes were packed in ice and placed on an end-end reciprocating shaker set to 120 rev. min-1. Following this they were centrifuged at 3200 g for 30 min. Supernatant was discarded. EPS was then extracted from the remaining pellet. This was done by re-suspending the pellet in extraction buffer (2 mM Na<sub>3</sub>PO<sub>4</sub>.12H<sub>2</sub>O (0.760 g L<sup>-1</sup>), 4 mM NaH<sub>2</sub>PO<sub>4</sub>.H<sub>2</sub>O (0.552 g L<sup>-1</sup>), 9 mM NaCl (0.526 g L<sup>-1</sup>), 1 mM KCl (0.0746 g L<sup>-1</sup>), adjusted to pH 7 with 1 M HCl and pre-cooled to 4 °C. CER (Dowex 'Marathon C' sodium form, strongly acidic, 20-50 mesh) was prewashed twice in the above buffer, and 15.98 g CER was added together with the 25 mL buffer. This amount of CER was determined on the basis of 178 mg CER mg<sup>-1</sup> SOC, as required by the soil with highest SOC content. Tubes were shaken for 2 h, and then centrifuged at 4000 g for 30 min. The supernatant containing EPS was transferred into new tubes, frozen in liquid N2, and stored at -80 °C prior to analysis. Total polysaccharide and uronic acids were quantified as described by DuBois et al. (1956) and Mojica et al. (2007), respectively. Extracted protein was measured using a Lowry based technique with reagent concentrations modified for microplate format as described by Redmile-Gordon et al. (2013), except that no additions of model polyphenolics or dilutions of soil extracts were required. As in previous work, EPS-protein was analysed by comparison to standards of bovine serum albumin (BSA; Sigma A7906) prepared in EPS buffer. Redmile-Gordon et al., (2013) tested the approach proposed by Frolund et al. (1995) to correct for interferences in the extracts. The interference of non-proteinaceous chromogens in natural soil extracts was well described. Accordingly, the interference from non-proteinaceous chromogens in the soil extracts was corrected for by i) measuring the absorbance of a second set of samples using Lowry reagents

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without CuSO<sub>4</sub>, and ii) applying the mathematical correction as presented by Frolund et al.,

236 (1995) i.e.

$$237 \quad Abs_{protein} = 1.25(Abs_A - Abs_B) \tag{1}$$

Where  $Abs_A$  is the absorbance given for soil extracts using Lowry reagents,  $Abs_B$  is the absorbance given by the set analysed using Lowry reagents without added CuSO<sub>4</sub>, and  $Abs_{protein}$  is the theoretical absorbance due to protein, which is compared against a standard curve of BSA standards made in EPS extraction buffer.

SOC and total N concentrations were determined on the 3-5 mm fraction which was firstly air-dried, and fine-milled (< 355 µm) before dry combustion in a Leco TruMac Combustion Analyser (LECO Corp., St Joseph, MI, USA) after carbonate removal with HCI. The MWD (µm) of stable 3-5 mm aggregates was obtained following the procedure for slaking by 'fast wetting' described by Le Bissonnais (1996; ISO, 2012). We selected the 'fast wetting' component of the test as this assesses the response of soil to a highly-disaggregating force, as controlled by the strength of aggregates and their wetting rate.

#### 2.3 Statistical analyses

All statistical analyses were completed using the GenStat (18<sup>th</sup> Edition) programme (VSN International Ltd., Hemel Hempstead, UK). We first assessed whether the residuals were normally distributed, and transformed (log<sub>10</sub>) the SOC, N and MWD variates accordingly. The complexity of the previous treatments in the Highfield experiments prior to the imposition of the current treatments was recently described by Gregory et al. (2016). Following their approach, we tested for significant effects of previous and current management treatments on the variates measured using analysis of variance (ANOVA) with the following model structures:

The 'previous' treatment factor was first nested (/) within an 'experiment' factor which allocated data according to whether it derived from either the original ley-arable or the bare fallow experiment, prior to forming the cross-products (\*) with the 'current' treatment factor. Likewise, the block model confirmed that there was no single block factor, but rather there were separate blocks for the different 'experiments' interacting (·). By convention in a factorial design, we compared the effects of the 'previous' and 'current' land use main factors (treatments averaged over their current and previous land use, respectively; n = 9), and their interaction (n = 3). We deemed treatment differences to be 'significant' if the probability (p) of the ANOVA variance ratio statistic (F) was < 0.05. We also conducted regression analysis to test for significant relationships between SOC, EPS and MWD variates.

272 3. Results

As an individual factor, previous land-use had no significant legacy effects on soil aggregate stability (MWD) or EPS (Table 2). However, there was strong legacy effect on total SOC and N concentrations which decreased significantly in the order grass > arable > fallow (p < 0.05; Table 2). A significant effect was also observed for current management with SOC and N (p < 0.05) all increasing under grass with the exception that there was no significant difference ascribed to current management between the SOC concentrations currently under arable and fallow (Table 2). Moreover, the extractable EPS-protein, EPS-polysaccharide and EPS-uronic acid pools were all strongly affected by current management, with the variates being significantly greater under grass compared to either arable or fallow (p < 0.05), which did not differ significantly. Stable aggregate MWD was also only influenced by current management (Table 2), being significantly greater for soil under grass compared to arable, and significantly greater under arable compared to fallow (p < 0.05; Table 3). Under the scheme of Le Bissonnais (1996), soils currently under grass, arable and fallow are classed as 'stable', 'unstable' and 'very unstable', respectively.

[Suggested position for Table 2]

The only variate that revealed a significant interaction between previous and current land-use was MWD (p = 0.035; Table 2). Within each previous management treatment, MWD showed statistically significant differences (p < 0.05) as a result of current land-use: MWD being highest for grass, intermediate for arable, and lowest for fallow within just 2.5 years since the current land-uses were established. Plots previously under grass showed enhanced structural stability when the current land-use was also grass (grass→grass).

## [Suggested position for Table 3]

Significant regression models (p < 0.05) were found for MWD ( $log_{10}$ ) as a function of SOC, EPS-protein and EPS-polysaccharide (adjusted  $R^2$  = 0.72, 0.30, and 0.15, respectively; Table 3). Accordingly, these are presented as Figure 2.

## [Suggested position for Figure 2]

Notably, all soils under grass appeared to exhibit MWDs in excess of those predicted by the regression between MWD and SOC (Figure 2). If it is assumed that the measured organic substances contribute to an increase in MWD (as is supposed; Tisdall and Oades, 1982) then the upper limit of the magnitude of the 'binding effect' per unit binder on a weight-for-weight basis (w/w) can be compared by standardising units SOC to the same scale as EPS parameters (Table 3). Standardising all to mg binder g-1 soil gives mass-adjusted coefficients for SOC, EPS-polysaccharide, and EPS-protein of 0.027, 1.6, and 3.6, respectively.

## [Suggested position for Figure 3]

Since there was a significant previous—current land use interaction on MWD (Table 2) we explored the links to SOC and EPS-protein by plotting the two most significant predictors of MWD on the natural scale (Figure 3). This provides visual clarity that something subtler than total SOC is affecting the MWD, and that appreciable changes in aggregate stability have been achieved within 2.5 year's application of the new managements. For example, soils previously under grass retained high SOC contents, but the MWDs of these treatments (grass—arable and grass—fallow) were only about half those of the arable—grass soils. Furthermore, arable—grass soils contained less SOC than grass—arable (Figure 3a). Accordingly, prediction of the expected stability order, based on ranking of SOC content was

successful in only 3 out of 9 cases with a mismatch occurring between SOC rank and MWD order for most land-use combinations (Figure 3a). In contrast, EPS-protein (Figure 3b) enabled better prediction of rankings for management-driven stabilities across the range of stability classes from 'very stable' to 'unstable' (Figure 3b): being successful in 6 out of 9 cases (Figure 3b). This relationship was more apparent with the more stable soils, where rank of EPS-protein matched the order of the top five structural stabilities: greatest EPS-protein for the greatest MWD, second greatest EPS-protein for the second greatest MWD, and so on.

All treatments previously under grass showed the greatest changes in SOC, with losses outweighing the loss or gain of any other permutation. This was not the case with EPS-protein (Figure 3b) where the magnitude of increase from fallow (fallow→grass) was similar to the loss of EPS-protein when transitioning from the reverse permutation (grass→fallow). There were greater losses of MWD, SOC and EPS-protein associated with transition to fallow (grass→fallow) over arable (grass→arable). Both transitions resulted in aggregates being classed as 'unstable' compared to those remaining under grass which were 'very stable' (Le Bissonnais, 1996). In contrast, soils previously under arable showed very little loss in SOC or EPS-protein content (1.64 to 1.56%; and 185 to 176 μg g<sup>-1</sup>) when converted to fallow, but the decrease in MWD was much greater: from 584 to 395 μm with a change of class from 'unstable' to 'very unstable' (Le Bissonnais, 1996).

The greatest total improvement in MWD was seen with transitions from arable to grass (increase of 590 μm), and while the SOC content of this treatment was less than the SOC content of the reverse grass—arable treatment, the EPS-protein was indeed greater, and the MWD was much greater too (at 1174 'medium' vs. 684 μm 'unstable'; Le Bissonnais, 1996). Likewise comparing the grass—fallow with fallow—grass treatment changes we see that the former retained a considerably greater SOC concentration (2.33 vs. 1.43%) but a considerably lesser EPS-protein concentration (157 vs. 191 μg g<sup>-1</sup>), while the latter had the greater EPS-protein concentration and structural stability as indicated by MWD (494 vs. 595 μm; Table 2).

We established that there was no significant interaction effect between previous × current land use on EPS fractions, however we did note some unexpected observations. EPS-

polysaccharide decreased with a transition from fallow to arable, as did EPS-protein (Table 2), but apart from these soils previously managed as fallow, there were no other similarities in land-use responses between these EPS pools. EPS-polysaccharide and EPS-uronic acid both unexpectedly increased when converted to the reduced C-input fallow (Table 2; arable→fallow; 318 to 375 μg EPS-polysaccharide; 167 to 194 μg EPS-uronic acid g-1 soil). Accordingly, the reverse fallow→arable conversion decreased all EPS fractions, but increased the MWD (Table 2). Additionally, the greatest MWD of all land-uses (grass→grass) was associated with merely average EPS-polysaccharide concentrations.

352 4. Discussion

4.1 Effects of land-use and land-use change on SOC, EPS and MWD

Continuous grassland clearly resulted in the greatest SOC, EPS and structural stability (MWD). We found total SOC concentration to be the best *overall* predictor of MWD according to the adjusted R<sup>2</sup> (Table 3). However, the maximum possible contribution of SOC to MWD was relatively low when considered on an equivalent mass basis: about 60 or 140 x smaller than that of EPS-polysaccharide or EPS-protein, respectively (Table 3). This stands to reason as not all SOC will have a binding effect. This also suggests that the relationship may not always be sensitive enough to describe how small changes in SOC quantity or quality affect a soil's stability. The lack of a decisive relationship between SOC and aggregate stability has been identified before (Tisdall and Oades, 1982; Liu et al., 2005; Denef et al., 2007; Sarker et al., 2018) and we explore this further in our data.

We found greater scattering in EPS-protein's relationship to MWD from the data shown in Figure 2b compared to Figure 2a, which could be due to variability introduced during the extraction of EPS prior to measurement, incomplete extraction, and/or reflect some natural underlying variability in the relationship. Indeed, one of the strengths of the total SOC measurement is the complete combustion of C in the analysis. Nonetheless, comparisons of Figures 3a and 3b show that EPS-protein may be more mechanistic in the formation —or maintenance— of the more stable aggregates in response to land-use and land-use change.

For example, total SOC (Figure 3a) did not explain the mean stability order of land-use combinations, with a mismatch occurring between SOC rank and MWD rank in most cases. In contrast, EPS-protein, which was controlled by current land-use (Table 1), described the stability order more accurately (Figure 3b). From a biofilm perspective this might be expected, because proteinaceous moieties in EPS are widely known to mediate biological attachment to abiotic surfaces (e.g. Taglialegna et al., 2016). A notable example of these impacts on stability at odds with total SOC is found in the extreme land-use changes between grass and fallow. While grass→fallow retained considerably greater SOC than fallow→grass (2.33 vs. 1.43%), the latter yielded more EPS-protein (191 vs. 157 μg g<sup>-1</sup>) and with this, significantly greater structural stability (MWD; 595 vs. 494 μm; Table 2).

There were examples where considerable decreases in MWD were not fully explained by either SOC or EPS. Treatments previously under arable showed very little loss in SOC when converted to fallow (1.64 to 1.56%, a relative decrease of 5%), but the decline in soil physical stability (MWD) was much greater and significant ( p <0.05) from 584 μm to 395 μm: a relative decrease of 32%, or transition from qualitative classes 'unstable' to 'very unstable' (Le Bissonnais, 1996). In this instance, the loss of EPS-protein was similar to SOC (185 to  $176~\mu g~EPS~g^{-1}$ ; relative decrease of 5%). This decline is likely to be more related to differences in temporary binding agents (plant roots and hyphae) because increased tillage is well-known known for its disruptive influence on fungal hyphae in fallow soils (Kabir, 2005). This magnitude of decline in aggregate stability is also similar to that found by Blankinship et al. (2016) where plants were manually removed from a seasonally dry grassland. We also found examples where a considerable increase in MWD was not fully explained by either SOC or EPS. The transition from arable to grass corresponded to increases of 7-48% in the EPS fractions and a 46% increase in SOC but a much greater (101%) increase in MWD. As above this suggests temporary binding agents were also implicated. Grass leys in arable rotation are widely recognised for their capacity to improve soil physical quality, which is understood to be largely owed to improvements in both temporary and transient binding agents (Tisdall and Oades, 1982). Accordingly, the size of the microbial biomass and abundance of fungi at our

experimental site were reported by other authors to increase in the order fallow>arable>grass (Gregory et al., 2009; Hirsch et al., 2009; Wu et al., 2012). In the present study, the only binding agents measured were *transient*. However, most roots were removed in the preparation (Section 2.2) and so we assume that fungal hyphae likely accounted for the majority of *temporary* binding effects.

Considering the positive interaction between previous and current land-uses observed for MWD (p < 0.05; Table 2; Figure 3a) it is apparent that the beneficial effects of current grass and arable land-uses on MWD must have been enhanced by specific conditions at the time of land-use change (owed to the preceding 50 year's land-use). Root establishment of the grass ley and winter wheat (2.5 years prior to sampling) would have clearly benefitted from the soil physical and chemical properties originating from the previous grass land-use (Table 1). Increased plant cover and/or reduced tillage therefore can be seen to have provided positive feedback for the subsequently beneficial land-uses. Nevertheless, no measurable legacy effects or interactions were detected for any of the extracted EPS (Table 2) confirming that the EPS themselves were transient. Furthermore, it is not likely that persistent binding agents declined substantially in the 2.5 years following land-use change, because organo-mineral associations tend to be resistant to oxidation and microbial metabolism (Kögel-Knabner et al., 2008). Therefore, we expect that this interaction effect is due to temporary binding agents coexisting at the time of sampling. Both temporary and transient binding agents are predominantly composed of relatively 'labile' carbon compounds, which are a temporally unstable pool known to decline relative to total SOC in response to reduced plant cover and increased tillage (Haynes, 2000).

These combined findings above support the claim of Jensen et al. (2019) that distinct components of SOC, rather than size of the total pool, have a disproportionate effect on aggregate stability. We also observed a larger range, and greater maximum total SOC between previous land-use classes than between current ones (Table 2). This suggests that total SOC content was primarily owed to previous land-use, and moreover that the bulk of SOC in soil previously under grass persisted > 2.5 years. This is supported by historically

fallow plots containing the least SOC whereas historically grass-covered plots contain the most (Table 2). While the concentration of 'persistent' or 'older' SOC cannot be used as a surrogate for *persistent binding agents*, it is important that this large quantity of organic material associated with previous land-uses does not appear to contribute significantly to the stability of soils in our study (see MWD; Table 2; p = 0.137). Furthermore, this form —or location— of SOC was not readily available for production of microbial EPS.

In accordance with the present findings, Redmile-Gordon et al. (2015) took soils from the same grass—arable plots in February 2012 and found that the relatively high SOC concentration did not support microbial assimilation of inorganic N (15NH<sub>4</sub>15NO<sub>3</sub>) provided in the laboratory. Neither was there any exudation of 15N labelled EPS into the extracellular space *unless* 15N was accompanied with simultaneous inputs of labile C. The above study highlighted the importance of fresh C inputs for EPS production, which is reflected in the present findings of elevated quantities of EPS-protein, EPS-polysaccharide and EPS-uronic acids in soils under dense vegetation cover at the time of sampling (i.e. grass; Table 2). Moreover, these findings reinforce the notion of a 'turning point' where older SOC, which is presumably more decomposed or physically occluded, ceases to be a viable source of C to measurably support exudation of EPS. This is further illustrated by the considerably greater EPS concentrations (protein, polysaccharide and uronic acid) under fallow—grass compared to grass—fallow (Table 2) and thus supports the theory presented by Golchin et al. (1994). Our findings show that measurement of 'transient' binding agents using the method of Redmile-Gordon et al., (2014) can add useful depth and contrast to measures of total SOC.

## 4.2 SOC vs. biological binding agents as drivers of soil stability

There remains a small paradox to be settled in that SOC shows a good correlation to aggregate stability, while the previous land-managements that contribute the majority of this SOC were not directly related to the MWD observed (Table 2; p = 0.137). However, as discussed in section 4.1, a positive interaction arising from the previous grass land-use was driving improved soil quality generally, for example with high SOC contributing a much

reduced bulk density (Table 1). More successful plant establishment in 2008 would therefore be expected, and is confirmed by yield measurements taken by other investigators at the same site (e.g. Hirsch et al., 2017). This increased plant biomass would in turn have increased the abundance of associated binding agents such as roots and hyphae - contributing to the MWD measured at the site. Importantly, this clarifies an important point that residual SOC needs no direct contribution to soil stability to indirectly contribute to a good R² for MWD via positive feedback effects. This example illustrates the case that large quantities of 'inactive' SOC can contribute relatively small concentrations of temporary and transient binding agents, which are overwhelmed by the contributions from current land-use (Table 2; Figure 3b).

While it could be argued that EPS production may also have been boosted by the same factors presented in Table 1, this fraction is sufficiently 'transient' that any increases in EPS that occurred previous to the moment of land-use change were no longer detectable at the time of sampling (Table 2). However, there would have been an abundance of EPS in plots formerly under grass, which interestingly corresponds to the increase in exchangeable multivalent cations (Ca²+ and Mg²+) under this land-use (Table 1). As discussed in section 4.1, fresh C was previously seen to be vital for production of EPS in these soils. Multivalent cations not only bridge carboxylic acid moieties within the EPS, but when sufficiently concentrated, contribute electrostatic interactions between the EPS and negatively charged mineral surfaces to form considerably stable bonds (Berne et al., 2015). We propose that electrostatic bonding between clays and biota could therefore depend on soil biota being able to maintain production of EPS to retain anchorage of these cations. Our data suggest that the majority of all stabilising effects subsided within 2.5 years of transition away from the high-C grass land-use, with MWD of grass—arable becoming similar to that of continuous arable by the time of measurement (Table 2; LSD 0.16 log<sub>10</sub> μm).

## 4.3 EPS-protein vs. EPS-polysaccharide

While Tisdall and Oades (1982) claimed that extracellular polysaccharides would be the most important transient binding agents in soil, investigations of biofilms in other environments show that protein and polysaccharide pools are not mutually exclusive, with glycoproteins typically being abundant, and essential, in bonding (Flemming et al., 2007). Among the transient EPS parameters measured here, EPS-protein showed the greatest coefficient of regression and R<sup>2</sup> against MWD (Table 3). Proteinaceous moieties in the EPS matrix are especially recognised for their contribution to stable biofilm structure (Vlamakis et al., 2013). The relatively low R<sup>2</sup> for polysaccharide and protein (0.15, and 0.30, respectively) are expected because the contributions of *temporary* (and *persistent*) binding agents are, of course, excluded.

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With regard to the mechanisms underlying the apparently EPS-protein driven stabilisation in the more stable soils (Fig. 3b), BsIA proteins exuded from the model soil organism (Bacillus subtilis) were found to self-assemble extracellularly; providing architectural stability and protecting the otherwise fragile polysaccharides against disruption from fast wetting (Arnaouteli et al., 2016). Earlier work found that hydrophobic amino acids —most notably phenylalanine and tyrosine— were characteristic of community EPS produced in response to a hydrophilic source of C (Redmile-Gordon et al., 2015). Tyrosine has since been shown to be of mechanistic importance in the formation of adhesive peptides, which impart strength and elasticity to 'living glues' produced by soil organisms (Zhang et al., 2019). The elevated concentrations of proteinaceous EPS in soils currently under grass (Table 2) is perhaps not surprising given that increased availability of plant polysaccharides is known to trigger production of extracellular proteins in the laboratory (Beauregard et al., 2013). However, given that in the present study a) these pools of EPS are transient, and b) solar radiation was at a seasonal low, it is somewhat surprising that these increases in community EPS were still measurable in the field by extraction in the temperate midwinter. Whether this is indicative of basal winter photosynthesis supporting sufficient rhizodeposit-C or a minimum turnover rate equal to several months we cannot say. Nonetheless, the majority do not remain extractable for more than 2.5 years, with previous land-use having no direct effect on the size of this pool (p = 0.823; Table 2).

Exceptions to the straightforward relationship of [increased plant-C] + [decreased tillage] = [greater EPS] are seen with land-use changes between fallow and arable. Fallow—arable conversion, with its implicit increases in C-inputs and decreased tillage, showed marked declines in EPS-polysaccharide and uronic acid fractions amounting to -51 and -41 µg EPS g-1 soil, respectively (Table 2). Likewise, arable→fallow conversion had notably increased EPS-polysaccharide and EPS-uronic acid fractions compared to its arable →arable counterpart, despite clearly greater C inputs from winter wheat and comparatively reduced tillage. Precedent exists for two feasible explanations. Firstly, the increased solar radiation for cyanobacteria and other photoautotrophs on the fallow surface may enable sufficient production of extracellular polysaccharides to be detected in the bulk soil. Cyanobacterial polysaccharides can stabilise soil surface crusts (Cania et al., 2019) but are apparently not highly influential for the stability of bulk soil (Table 2). Secondly, and it seems more likely, the inorganic N fertiliser applied annually to these arable plots somehow reduced the efficiency of microbial EPS production in the bulk soil (mass of EPS per unit microbial ATP), as shown previously by Redmile-Gordon et al. (2015). In every other sense, the arable land-use represents the 'intermediate' of individual factors contributing to the contrasting land-use pressures, i.e. tillage frequency, C inputs, duration of vegetative cover, and pH (Table 1). Wu et al. (2012) also reported intermediate soil microbial biomass concentrations, as expected between grass and fallow. Nonetheless, the arable land-use is exceptional in that inorganic N fertiliser is applied annually. Our data therefore support the conceptual model proposed by Redmile-Gordon et al. (2015) that while C is a fundamental requirement for significant EPS production, surplus inorganic N acts as a 'switch': shifting soil microbial investment of C from EPS to intracellular investment for growth of cells.

#### 4.4 Implications for land-use and soil management strategies

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Most SOC (> 98%) in sieved soil taken from a grass—arable plot of the same longterm experiment was shown to be non-living and inaccessible to the soil microbial biomass (Redmile-Gordon et al., 2015). In the present study, the residual fraction of SOC from the same previous land-use was also found to not be directly influential for soil structural stability (Table 2) with the primary binding agents operating at our study site being attributed to the influence of current land-use over the long-term previous land-use (Table 2). Taken together, these results suggest that soil biology, or biological binding agents are vital for favourable soil quality (MWD). The significant shifts in this reasonably 'holistic' measure of soil quality (Six et al., 2000) were achieved within a relatively short period (< 2.5 years). Management of biological binding agents, including EPS as a highly responsive *transient binding agent* may therefore be useful to rapidly improve soil structural stability to meet various environmental objectives. The approach of building a smaller more 'active' subset of total SOC may also help to avoid some of the seasonal risks associated with large pools of total SOC, including emissions of carbon dioxide, leaching of N overwinter, contamination of rivers, aquifers, and the subsequent indirect emissions of nitrous oxide arising from leached N (Reay et al., 2012).

5. ConclusionWe confirmed that the primary binding agents affected by current land-use were transient and/or temporary. Direct effects of current mixed grass and clover, without fertiliser inputs, were more important for both soil physical stability, and EPS, than any legacy from a previous land-use (including greater SOC). Among the extracellular fractions studied, the stability of aggregates was most strongly related to increases in EPS-protein, less related to EPS-polysaccharide, and not related to uronic acids. We also demonstrated that measurement of transient binding agents, using a recently developed method, adds useful depth and contrast to measures of total SOC. We conclude that there may be significant environmental benefits to managing smaller specific 'functional' pools of C in agriculture and horticulture, rather than increasing total SOC *per se*.

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## 564 References

- Arnaouteli, S., MacPhee, C.E., Stanley-Wall, N.R., 2016. Just in case it rains: building a
- 566 hydrophobic biofilm the *Bacillus subtilis* way. Curr. Opin. Microbiol. 34, 7-12.
- Avery, B.W., Catt, J.A., 1995. The Soil at Rothamsted. Lawes Agricultural Trust, Harpenden,
- 568 UK, pp. 44.
- Bach, E.M., Baer, S.G., Meyer, C.K., Six, J., 2010. Soil texture affects soil microbial and
- 570 structural recovery during grassland restoration. Soil Biol. Biochem. 42, 2182-2191.
- 571 Bacq-Labreuil, A., Crawford, J., Mooney, S.J., Neal, A.L., Akkari, E., McAuliffe, C., Zhang, X.,
- Redmile-Gordon, M., Ritz, K., 2018. Effects of cropping systems upon the three-dimensional
- architecture of soil systems are modulated by texture. Geoderma, 332, pp. 73-83.
- Beauregard, P.B., Chai, Y., Vlamakis, H., Losick, R., Kolter, R., 2013. *Bacillus subtilis* biofilm
- 575 induction by plant polysaccharides. Proc. Nat. Acad. Sci. USA 110, 1621-1630.
- 576 Berne, C., Ducret, A., Hardy, G.G., & Brun, Y.V., 2015. Adhesins involved in attachment to
- abiotic surfaces by Gram-negative bacteria. Microbiol. Spectr. 3, MB-0018-2015.
- 578 Blankinship, J.C., Fonte, S.J., Six, J., Schimel, J.P., 2016. Plant versus microbial controls on
- 579 soil aggregate stability in a seasonally dry ecosystem. Geoderma 272, 39-50.
- 580 Caesar-Tonthat, T.C., Stevens, W.B., Sainju, U.M., Caesar, A.J., West, M., Gaskin, J. F.,
- 581 2014. Soil-aggregating bacterial community as affected by irrigation, tillage, and cropping
- system in the northern great plains. Soil Sci. 179, 11–20.
- 583 Cania, B., Vestergaard, G., Kublik, S., Köhne, J.M., Fischer, T., Albert, A., Winkler, B.,
- 584 Schloter, M., Schulz, S., 2019. Biological soil crusts from different soil substrates harbor
- 585 distinct bacterial groups with the potential to produce exopolysaccharides and
- 586 lipopolysaccharides. Microb. Ecol.

- Chabbi, A., Lehmann, J., Ciais, P., Loescher, H.W., Cotrufo, M.F., Don, A., SanClements, M.,
- Schipper, L., Six, J., Smith, P., Rumpel, C., 2017. Aligning agriculture and climate policy. Nat.
- 589 Climate Change 7, 307-309.
- 590 Chapman, S.J., Bell, J.S., Campbell, C.D., Hudson, G., Lilly, A., Nolan, A.J., Robertson,
- 591 A.H.J., Potts, J.M. & Towers, W., 2013. Comparison of soil carbon stocks in Scottish soils
- 592 between 1978 and 2009. Eur. J. Soil Sci. 64, 455-465.
- 593 Costa, O., Raaijmakers, J.M., Kuramae, E.E, 2018. Microbial extracellular polymeric
- 594 substances: ecological function and impact on soil aggregation. Front. Microbiol. 9, 1636.
- Denef, K., Zotarelli, L., Boddey, R.M., Six, J., 2007. Microaggregate-associated carbon as a
- 596 diagnostic fraction for management-induced changes in soil organic carbon in two Oxisols.
- 597 Soil Biol. Biochem. 39, 1165-1172.
- 598 Denef, K., Six, J., 2005. Clay mineralogy determines the importance of biological versus
- abiotic processes for macroaggregate formation and stabilization. Eur. J. Soil Sci. 56, 469-
- 600 479.
- Flemming, H.C., Neu, T.R., Wozniak, D.J., 2007. The EPS matrix: the "house of biofilm cells".
- 602 J. Bacteriol. 189, 7945-7947.
- Flemming, H.C., Wingender, J., 2010. The biofilm matrix. Nature Rev. Microbiol. 8, 623-633.
- Frolund, B., Griebe, T., Nielsen, P.H., 1995. Enzymatic-activity in the activated-sludge
- floc matrix. Appl. Microbiol. Biotechnol. 43, 755-761.
- 606 Frolund, B., Palmgren, R., Keiding, K., Nielsen, P.H., 1996. Extraction of extracellular
- polymers from activated sludge using a cation exchange resin. Water Res. 30, 1749-1758.
- 608 Ge, L.Y., Deng, H.H., Gao, H.W., Wu, F., 2010. Optimized extraction protocol for extracellular
- polymeric substances (EPS) from two activated sludges. Res. J. Chem. Environ. 14, 78-82.
- 610 Gillespie, A.W., Farrell, R.E., Walley, F.L., Ross, A.R.S., Leinweber, P., Eckhardt, K.-U.,
- Regier, T.Z., Blyth, R.I.R., 2011. Glomalin-related soil protein contains nonmycorrhizal-
- related heat-stable proteins, lipids and humic materials. Soil Biol. Biochem. 43, 766-777.
- 613 Golchin, A., Oades, J.M., Skjemstad, J.O. Clarke, P., 1994. Soil structure and carbon cycling.
- 614 Aus. J. Soil Res. 32, 1043-1068.

- 615 Gregory, A.S., Watts, C.W., Griffiths, B.S., Hallett, P.D., Kuan, H.L., Whitmore, A.P., 2009.
- The effect of long-term soil management on the physical and biological resilience of a range
- of arable and grassland soils in England. Geoderma 153, 172-185.
- Gregory, A.S., Dungait, J.A.J., Watts, C.W., Bol, R., Dixon, E.R., White, R.P., Whitmore, A.P.,
- 619 2016. Long-term management changes topsoil and subsoil organic carbon and nitrogen
- dynamics in a temperate agricultural system. Eur. J. Soil Sci. 67, 421-430.
- 621 Guo, X., Wang, X., & Liu, J., 2016. Composition analysis of fractions of extracellular polymeric
- 622 substances from an activated sludge culture and identification of dominant forces affecting
- 623 microbial aggregation. Sci. Rep. 6, 28391.
- 624 Guo, Y., Fan, R., Zhang, X., Zhang, Y., Wu, D., McLaughlin, N., Zhang, S., Chen, X., Jia, S.,
- 625 Liang, A., 2020. Tillage-induced effects on SOC through changes in aggregate stability and
- 626 soil pore structure. Sci. Tot. Environ. 703. https://doi.org/10.1016/j.scitotenv.2019.134617
- 627 Haynes, R.J., 2000. Labile organic matter as an indicator of organic matter quality in arable
- and pastoral soils in New Zealand. Soil Biol. Biochem. 32, 211-219.
- Hirsch, P.R., Jhurreea, D., Williams, J.K., Murray, P.J., Scott, T., Misselbrook, T.H., Goulding,
- 630 K.W.T., Clark, I.M., 2017. Soil resilience and recovery: rapid community responses to
- management changes. Plant Soil 412, 283-297.
- 632 ISO, 2012. ISO10930:2012. Soil Quality Measurement of the Stability of Soil Aggregates
- 633 Subjected to the Action of Water. International Organization for Standardization, Geneva,
- 634 Switzerland, pp. 20.
- 635 IUSS Working Group WRB. 2015. World Reference Base for Soil Resources 2014, Update
- 636 2015. International soil classification system for naming soils and creating legends for soil
- 637 maps. World Soil Resources Report 106. Food and Agriculture Organization of the United
- Nations International Union of Soil Sciences. FAO, Rome, pp. 192.
- Jensen, J.L., Schjönning, P., Watts, C.W., Christensen, B.T., Peltre, C., Munkholm, L.J., 2019.
- Relating soil C and organic matter fractions to soil structural stability. Geoderma 337, 834-
- 641 843.

- Jobbágy, E.G. & Jackson, R.B. 2000. The vertical distribution of soil organic carbon and its
- relation to climate and vegetation. Ecological Applications, 10, 423-436.
- 644 Johnston, A.E., Poulton, P.R., Coleman, K., 2009. Soil organic matter: its importance in
- sustainable agriculture and carbon dioxide fluxes. Adv. Agron. 101, 1-57.
- Jones, A.R., Orton, T.G., Dalal, R.C., 2016. The legacy of cropping history reduces the
- recovery of soil carbon and nitrogen after conversion from continuous cropping to permanent
- 648 pasture. Agric. Ecosyst. Environ. 216, 166-176.
- 649 Kabir, Z., 2005. Tillage or no-tillage: impact on mycorrhizae. Can. J. Plant Sci. 85, 23-29.
- 650 Kemper, W.D., Koch, E.J., 1966. Aggregate stability of soils from western United States and
- 651 Canada. United States Department of Agriculture Technical Bulletin 1355. US Government
- 652 Printing Office, Washington DC, USA, pp. 52.
- 653 Kögel-Knabner, I., Guggenberger, G., Kleber, M., Kandeler, E., Kalbitz, K., Scheu, S.,
- 654 Eusterhues, K., Leinweber, P., 2008. Organo-mineral associations in temperate soils:
- Integrating biology, mineralogy, and organic matter chemistry. J. Plant Nutr. Soil Sci. 171, 61-
- 656 82.
- 657 Krause, L., Biesgen, D., Treder, A., Schweizer, S.A., Klumpp, E., Knief, C., Siebers, N., 2019.
- 658 Initial microaggregate formation: Association of microorganisms to montmorillonite-goethite
- aggregates under wetting and drying cycles. Geoderma, 351, 250-260.
- Le Bissonnais, Y., 1996. Aggregate stability and assessment of soil crustability and erodibility.
- 1. Theory and methodology. Eur. J. Soil Sci. 47, 425-437.
- Liang, C., Schimel, J.P., Jastrow, J.D., 2017. The importance of anabolism in microbial control
- over soil carbon storage. Nat. Microbiol. 2, 1–6.
- Liu, A.G., Ma, B.L., Bomke, A.A., 2005. Effects of cover crops on soil aggregate stability, total
- organic carbon, and polysaccharides. Soil Sci. Soc. Am. J. 69, 2041-2048.
- 666 Marchus, K.A., Blankinship, J.C., Schimel, J.P., 2018. Environmental controls on extracellular
- 667 polysaccharide accumulation in a California grassland soil. Soil Biol. Biochem. 125, 86-92.

- Poeplau, C., Don, A., Vesterdal, L., Leifeld, J., van Wesemael, B., Schumacher, J., Gensior,
- 669 A., 2011. Temporal dynamics of soil organic carbon after land-use change in the temperate
- zone carbon response functions as a model approach. Glob. Change Biol. 17, 2415-2427.
- 671 Poulton, P., Johnston, J., Macdonald, A., White, R., Powlson, D., 2018. Major limitations to
- achieving "4 per 1000" increases in soil organic carbon stock in temperate regions: evidence
- 673 from long-term experiments at Rothamsted Research, United Kingdom, Glob, Change Biol.
- 674 24, 2563-2584.
- 675 Reay, D.S., Davidson, E.A., Smith, K.A., Smith, P., Melillo, J.M., Dentener, F., Crutzen, P.J.,
- 676 2012. Global agriculture and nitrous oxide emissions. Nat. Clim. Change 2, 410-416.
- Redmile-Gordon, M., White, R.P., Brookes, P.C., 2011. Evaluation of substitutes for paraguat
- 678 in soil microbial ATP determinations using the trichloroacetic acid-based reagent of Jenkinson
- and Oades (1979). Soil Biol. Biochem. 43, 1098-1100.
- Redmile-Gordon, M.A., Armenise, E., White, R.P., Hirsch, P.R., Goulding, K.W.T., 2013. A
- 681 comparison of two colorimetric assays, based upon Lowry and Bradford techniques, to
- estimate total protein in soil extracts. Soil Biol. Biochem. 67, 166-173.
- Redmile-Gordon, M.A., Brookes, P.C., Evershed, R.P., Goulding, K.W.T., Hirsch, P.R., 2014.
- 684 Measuring the soil-microbial interface: extraction of extracellular polymeric substances (EPS)
- from soil biofilms. Soil Biol. Biochem 72, 163-171.
- Redmile-Gordon, M.A., Evershed, R.P., Hirsch, P.R., White, R.P., Goulding, K.W.T., 2015.
- 687 Soil organic matter and the extracellular microbial matrix show contrasting responses to C and
- 688 N availability. Soil Biol. Biochem. 88, 257-267.
- Sarker, J.R., Singh, B.P., Cowie, A.L., Fang, Y.Y., Collins, D., Badgery, W., Dalal, R.C., 2018.
- 690 Agricultural management practices impacted carbon and nutrient concentrations in soil
- aggregates, with minimal influence on aggregate stability and total carbon and nutrient stocks
- in contrasting soils. Soil Tillage Res. 178, 209-223.
- 693 Scott, T., Macdonald, A.J., Goulding, K.W.T., 2014. The UK Environmental Change Network,
- 694 Rothamsted. Physical and Atmospheric Measurements: The First 20 Years. Lawes
- 695 Agricultural Trust Co. Ltd., Harpenden, UK, pp. 32.

- 696 Sheng, G.-P., Yu, H.-Q., Li, X.-Y., 2010. Extracellular polymeric substances (EPS) of microbial
- aggregates in biological wastewater treatment systems: a review. Biotech. Adv. 28, 882–894.
- 698 Six, J., Elliott, E.T., & Paustian, K., 2000. Soil structure and soil organic matter: II. A normalized
- stability index and the effect of mineralogy. Soil Sci. Soc. Am. J. 64, 1042-1049.
- 700 Six, J., Conant, R.T., Paul, E.A., Paustian, K., 2002. Stabilization mechanisms of soil organic
- 701 matter: implications for C-saturation of soils. Plant Soil 241, 155-176.
- 702 Soinne, H., Hyvaluoma, J., Ketoja, E., Turtola, E., 2016. Relative importance of organic
- carbon, land use and moisture conditions for the aggregate stability of post-glacial clay soils.
- 704 Soil Tillage Res. 158, 1-9.
- 705 Takahashi, E., Ledauphin, J., Goux, D., Orvain, F., 2009. Optimising extraction of extracellular
- 706 polymeric substances (EPS) from benthic diatoms: comparison of the efficiency of six EPS
- 707 extraction methods. Mar. Freshwater Res. 60, 1201-1210.
- Taglialegna, A., Lasa, I., Valle, J., 2016. Amyloid structures as biofilm matrix scaffolds.
- 709 J Bacteriol. 198, pp. 2579-2588.
- 710 Tang, J., Mo, Y.H., Zhang, J.Y., Zhang, R.D., 2011. Influence of biological aggregating agents
- associated with microbial population on soil aggregate stability. Appl. Soil Ecol. 47, 153-159.
- 712 Tisdall, J.M., Oades, J.M., 1982. Organic-matter and water-stable aggregates in soils. J. Soil
- 713 Sci. 33, 141-163.
- Vlamakis, H., Chai, Y., Beauregard, P., Losick, R., Kolter, R., 2013. Sticking together: building
- a biofilm the *Bacillus subtilis* way. Nat Rev. Microbiol. 11, 157–168.
- Wang, S., Redmile-Gordon, M., Mortimer, M., Cai, P., Wu, Y., Peacock, C.L., Gao, C., Huang,
- 717 Q. Extraction of extracellular polymeric substances (EPS) from red soils (Ultisols), 2019. Soil
- 718 Biol. and Biochem. 135, 283-285.
- Watts, C.W., Whalley, W.R., Longstaff, D., White, R.P., Brookes, P.C., Whitmore, A.P., 2001.
- 720 Aggregation of a soil with different cropping histories following the addition of organic
- 721 materials. Soil Use Manage. 17, 263-268.
- Watts, C.W., Whalley, W.R., Brookes, P.C., Devonshire, B.J., Whitmore, A.P., 2005. Biological
- and physical processes that mediate micro-aggregation of clays. Soil Sci. 170, 573-583.

- Wiesmeier, M., Spörlein, P., Geuß, U., Hangen, E., Haug, S., Reischl, A., Schilling, B., von 724
- Lützow, M., Kögel-Knabner, I., 2012. Soil organic carbon stocks in southeast Germany 725
- (Bavaria) as affected by land use, soil type and sampling depth. Glob. Change Biol. 18, 2233-726
- 727 2245.
- Wright, S.F., Upadhyaya, A., 1996. Extraction of an abundant and unusual protein from soil 728
- and comparison with hyphal protein of arbuscular mycorrhizal fungi. Soil Sci., 161, 575-586. 729
- Wu, Y., Kemmitt, S., White, R.P., Xu, J., Brookes, P.C., 2012. Carbon dynamics in a 60 year 730
- fallowed loamy-sand soil compared to that in a 60 year permanent arable or permanent 731
- grassland UK soil. Plant Soil, 352, 51-63. 732
- Zhang, C., Huang, J., Zhang, J., Liu, S., Cui, M., An, B., Wang, X., Pu, J., Zhao, T., Fan, C., 733
- Lu, T.K., Zhong, C., 2019. Engineered Bacillus subtilis biofilms as living glues. Mat. Today 28, 734
- 735 40-48.

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Fig. 1. The Highfield experiment with previous grass (G), arable (A) (both since 1949, in 50 738  $\times$  7 m plots) and bare fallow (F) (since 1959, in adjacent land totalling c. 1400 m<sup>2</sup>) land uses, 739 with superimposed current grass (black), arable (grey) and fallow (white) land uses. The 740 experimental design comprises 9 treatments triplicated in 27 split-plots (3 previous × 3 741 current land uses × 3 field replicates). Plot numbers are given. 742

743

Fig. 2. Stable aggregate mean weight diameter (MWD) as a linear function of (a) soil organic 744 C concentration (y-axis), and (b) EPS-protein concentration. Current grass, arable and fallow 745 land uses are shown (black, grey and white symbols, respectively). See Table 3 for further 746 details of the linear regression analyses. 747

748

- Fig. 3. Mean (n=3) (a) soil organic C concentration (y-axis), (b) EPS-protein concentration as 749 related to stable aggregate mean weight diameter (MWD; circles) for the nine previous × 750 current land use combinations. Previous land use (x-axis) is shown in factorial combination 751
- with current grass, arable and fallow land uses (black, grey and white symbols, respectively). 752
- The diameter of each circle represents the stability (scale bar at 1000 µm MWD). 753

**Table 1**Selected physical and chemical properties of the soil (upper 10 cm) at Highfield under long-term grass and arable (since 1949) and fallow (since 1959) land uses as measured in 2005 by Gregory et al. (2009) and Watts (personal communication) unless stated.

| Property                  | Unit  | Grass | Arable | Fallow |
|---------------------------|---|-------|--------|--------|
| Sand                      | g kg <sup>-1</sup>                          | 179   | 189    | 178    |
| Silt                      | g kg <sup>-1</sup>                          | 487   | 504    | 525    |
| Clay                      | g kg <sup>-1</sup>                          | 333   | 306    | 297    |
| Particle density          | g cm <sup>-3</sup>                          | 2.46  | 2.57   | 2.61   |
| Bulk density <sup>a</sup> | g cm <sup>-3</sup>                          | 0.99  | 1.44   | 1.32   |
| Plastic limit             | g kg <sup>-1</sup>                          | 484   | 225    | 208    |
| Liquid limit              | g kg <sup>-1</sup>                          | 663   | 364    | 319    |
| Plasticity index          | %   | 17.9  | 13.9   | 11.1   |
| Linear shrinkage          | %   | 11.0  | 10.0   | 8.2    |
| Water content –1 kPa      | g kg <sup>-1</sup>                          | 449   | 254    | 231    |
| Water content –5 kPa      | g kg <sup>-1</sup>                          | 407   | 244    | 219    |
| Water content –30 kPa     | g kg <sup>−1</sup>                          | 355   | 234    | 221    |
| Water content –300 kPa    | g kg <sup>-1</sup>                          | 178   | 126    | 115    |
| Water content –1500 kPa   | g kg <sup>-1</sup>                          | 176   | 115    | 99     |
| Exchangeable Ca           | mmol <sub>c</sub> kg <sup>−1</sup>          | 61.8  | 23.8   | 10.3   |
| Exchangeable Mg           | mmol <sub>c</sub> kg <sup>−1</sup>          | 6.0   | 3.5    | 4.2    |
| Exchangeable Mn           | mmol <sub>c</sub> kg <sup>−1</sup>          | 11    | 9.1    | 4.2    |
| Exchangeable K            | mmol <sub>c</sub> kg <sup>−1</sup>          | 4.2   | 1.4    | 8.0    |
| Exchangeable Na           | mmol <sub>c</sub> kg <sup>-1</sup>          | 1.0   | 1.9    | 8.0    |
| Cation exchange capacity  | mmol₀ kg <sup>-1</sup>                      | 78.9  | 36.4   | 18.7   |
| рН                        | -log (g [H <sup>+</sup> ] L <sup>-1</sup> ) | 6.30  | 5.76   | 4.43   |
| Electrical conductivity   | mS cm <sup>-1</sup>                         | 0.21  | 0.08   | 0.05   |

<sup>&</sup>lt;sup>a</sup> Gregory et al. (2016)

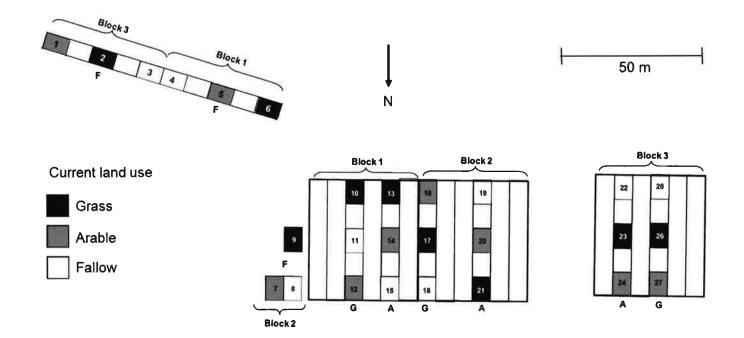
Table 2
Mean ± standard error of the mean of soil organic C (SOC), total N (N), and soil extracellular polymeric substances (EPS) concentrations (protein, polysaccharide and uronic acid; μg EPS g<sup>-1</sup> soil), and stable aggregate mean weight diameter (MWD), grouped by land uses (*n*=3), and their interaction (*n*=9). Where significant at *p*<0.05, the least significant differences (LSD) of means are given ('ns' indicates non-significance). Note that some variates were transformed by log<sub>10</sub> firstly to normalise the distribution of residuals.

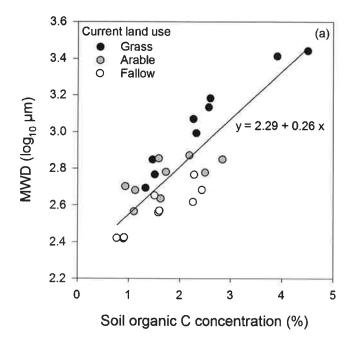
| Land use |         | SOC  |       |                       |      | N      |                       |       | EPS            | _     |                 |       |     | MW   | D    |                        |
|----------|---------|------|-------|-----------------------|------|--------|-----------------------|-------|----------------|-------|-----------------|-------|-----|------|------|------------------------|
|          |         |      |       |                       |      | protei |                       | ein   | polysaccharide |       | uronic acid     |       |     |      |      |                        |
|          |         | (0   | %)    | (log <sub>10</sub> )% | (    | %)     | (log <sub>10</sub> )% |       |                | (ին ն | <sup>-1</sup> ) |       |     | (µı  | m)   | (log <sub>10</sub> )µm |
| Previous |         |      |       |                       |      |        |                       |       |                |       |                 |       |     |      |      |                        |
| Grass    |         | 2.83 | ±0.27 | 0,439                 | 0.26 | ±0.02  | 0.598                 | 188   | ±12            | 327   | ±16             | 144   | ±7  | 1157 | ±307 | 2.96                   |
| Arable   |         | 1.86 | ±0.13 | 0.262                 | 0.18 | ±0.01  | -0.753                | 186   | ±10            | 376   | ±24             | 203   | ±16 | 718  | ±124 | 2.81                   |
| Fallow   |         | 1.12 | ±0.09 | 0.038                 | 0.11 | ±0.01  | -0.974                | 143   | ±15            | 301   | ±27             | 180   | ±17 | 437  | ±52  | 2.61                   |
| p        |         |      |       | 0.017                 |      |        | 0.035                 | 0.823 |                | 0.285 |                 | 0.086 |     |      |      | 0.137                  |
| LSD      |         |      |       | 0.100                 |      |        | 0.129                 | ns    |                | ns    |                 | ns    |     |      |      | ns                     |
| Current  |         |      |       |                       |      |        |                       |       |                |       |                 |       |     |      |      |                        |
| Grass    |         | 2.49 | ±0,36 | 0.362                 | 0.22 | ±0.03  | -0,681                | 201   | ±11            | 387   | ±16             | 209   | ±16 | 1354 | ±276 | 3.06                   |
| Arable   |         | 1.73 | ±0.22 | 0.211                 | 0.17 | ±0.02  | -0.804                | 163   | ±16            | 298   | ±22             | 152   | ±11 | 573  | ±45  | 2.75                   |
| Fallow   |         | 1.59 | ±0.21 | 0.166                 | 0.15 | ±0.02  | -0.840                | 153   | ±11            | 319   | ±25             | 166   | ±15 | 384  | ±37  | 2.57                   |
| р        |         |      |       | <0.001                |      |        | < 0.001               | 0.014 |                | 0.005 |                 | 0.002 |     |      |      | < 0.001                |
| LSD      |         |      |       | 0.053                 |      |        | 0.052                 | 31    |                | 50    |                 | 28    |     |      |      | 0.07                   |
| Previous | Current |      |       |                       |      |        |                       |       |                |       |                 |       |     |      |      |                        |
| Grass    | Grass   | 3.67 | ±0.56 | 0.553                 | 0.32 | ±0.05  | -0.503                | 213   | ±21            | 346   | ±9              | 152   | ±4  | 2293 | ±388 | 3.35                   |
|          | Arable  | 2.51 | ±0.19 | 0.397                 | 0.23 | ±0.02  | -0.632                | 195   | ±11            | 340   | ±24             | 152   | ±11 | 684  | ±43  | 2.83                   |
|          | Fallow  | 2.33 | ±0.06 | 0.367                 | 0.22 | ±0.01  | -0.659                | 157   | ±20            | 294   | ±41             | 129   | ±19 | 494  | ±49  | 2.69                   |
| Arable   | Grass   | 2.39 | ±0,09 | 0,377                 | 0.22 | ±0.01  | -0.653                | 198   | ±2             | 435   | ±13             | 248   | ±2  | 1174 | ±109 | 3.07                   |
|          | Arable  | 1.64 | ±0.04 | 0.215                 | 0.16 | ±0.00  | -0.787                | 185   | ±31            | 318   | ±45             | 167   | ±34 | 584  | ±82  | 2.76                   |
|          | Fallow  | 1.56 | ±0.03 | 0.194                 | 0.15 | ±0.00  | -0.818                | 176   | ±13            | 375   | ±37             | 194   | ±12 | 395  | ±27  | 2.60                   |
| Fallow   | Grass   | 1.43 | ±0.05 | 0.155                 | 0.13 | ±0.01  | -0.888                | 191   | ±29            | 382   | ±31             | 227   | ±19 | 595  | ±61  | 2.7                    |
|          | Arable  | 1.05 | ±0.06 | 0.021                 | 0.10 | ±0.00  | -0.992                | 111   | ±5             | 236   | ±3              | 136   | ±2  | 450  | ±42  | 2.65                   |
|          | Fallow  | 0.87 | ±0.05 | -0.063                | 0.09 | ±0.00  | -1.041                | 126   | ±14            | 287   | ±46             | 177   | ±30 | 264  | ±1   | 2.42                   |
| p        |         |      |       | 0.988                 |      |        | 0.988                 | 0.597 |                | 0.133 |                 | 0.073 |     |      |      | 0.035                  |
| LSD      |         |      |       | ns                    |      |        | ns                    | ns    |                | ns    |                 | ns    |     |      |      | 0.16                   |

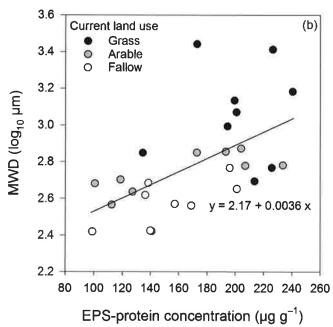
Table 3

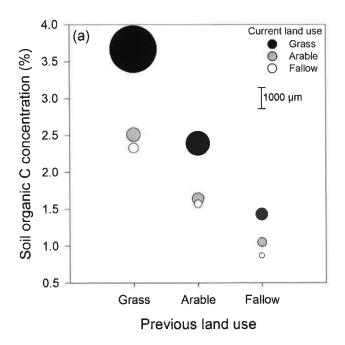
Stable aggregate mean weight diameter ( $(\log_{10}) \mu m$ ) as a linear function of soil organic C (SOC; %) and soil extracellular polymeric substances (EPS) concentrations (protein, polysaccharide and uronic acid;  $\mu g EPS g^{-1}$  soil). The table gives the constant (a) and coefficient (b) of the linear regression (y = a + bx), the probability level associated with the regression (p), and the adjusted proportion of the variance accounted for by the fit (Adjusted  $R_2$ ). Note that for uronic acid, the adjusted  $R_2$  is not calculable as the residual variance exceeded the variance of the response variate.

| Statistic               | SOC       | EPS           |                |                |  |  |  |  |
|-------------------------|-----------|---------------|----------------|----------------|--|--|--|--|
|                         |           | protein       | polysaccharide | uronic acid    |  |  |  |  |
| Constant                | 2.29±0.07 | 2.17±0.18     | 2.26±0.23      | 2.64±0.21      |  |  |  |  |
| Coefficient             | 0.26±0.03 | 0.0036±0.0010 | 0.0016±0.0007  | 0.0008±0.0011  |  |  |  |  |
| ρ                       | <0.001    | 0.002         | 0.027          | 0.465          |  |  |  |  |
| Adjusted R <sup>2</sup> | 0.719     | 0.301         | 0.149          | not calculable |  |  |  |  |









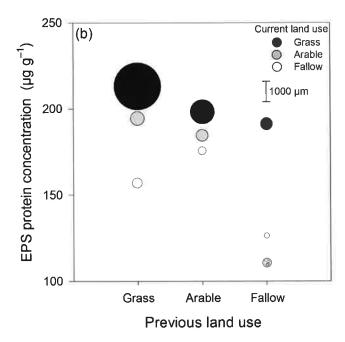


Table S1

Full dataset of soil organic C (SOC), total N (N) and extracellular polymeric substances (EPS) concentrations (protein, polysaccharide and uronic acid), and stable aggregate mean weight diameter (MWD) variates under previous and current land uses and their interaction.

| Land use |         | Block | Plot | Split-plot | SOC   | N                     |         | EPS            |             | MWD     |
|----------|---------|-------|------|------------|-------|-----------------------|---------|----------------|-------------|---------|
| Previous | Current |       |      |            |       |                       | protein | polysaccharide | uronic acid |         |
|          |         |       | (9   | %)         |       | (μg g <sup>-1</sup> ) |         | (µm)           |             |         |
| Grass    | Grass   | 1     | 4    | 10         | 4.500 | 0.386                 | 172.54  | 338.84         | 145.46      | 2768.57 |
| Grass    | Grass   | 2     | 6    | 17         | 3.900 | 0.349                 | 226.46  | 364.06         | 151.83      | 2587.72 |
| Grass    | Grass   | 3     | 9    | 26         | 2.600 | 0.230                 | 240.50  | 335.17         | 157.66      | 1524.0  |
|          |         |       |      | Mean       | 3.667 | 0.322                 | 213.17  | 346.02         | 151.65      | 2293.45 |
| Grass    | Arable  | 1     | 4    | 12         | 2.186 | 0.204                 | 204.10  | 366.75         | 158.35      | 744.63  |
| Grass    | Arable  | 2     | 6    | 16         | 2.840 | 0.266                 | 172.58  | 292.39         | 130.74      | 707.66  |
| Grass    | Arable  | 3     | 9    | 27         | 2.497 | 0.234                 | 207.00  | 359.51         | 165.75      | 599.74  |
|          |         |       |      | Mean       | 2.508 | 0.235                 | 194.56  | 339.55         | 151.61      | 684.00  |
| Grass    | Fallow  | 1     | 4    | 11         | 2.285 | 0.211                 | 196.00  | 366.98         | 163.26      | 584.45  |
| Grass    | Fallow  | 2     | 6    | 18         | 2.439 | 0.230                 | 138.51  | 290.60         | 124.38      | 482.65  |
| Grass    | Fallow  | 3     | 9    | 25         | 2.260 | 0.217                 | 136.36  | 224.40         | 98.01       | 414.90  |
|          |         |       |      | Mean       | 2.328 | 0.219                 | 156.96  | 293.99         | 128.55      | 494.00  |
| Arable   | Grass   | 1     | 5    | 13         | 2.570 | 0.240                 | 199.42  | 455.70         | 244.38      | 1362.67 |
| Arable   | Grass   | 2     | 7    | 21         | 2.264 | 0.211                 | 200.85  | 438.84         | 246.90      | 1175.54 |
| Arable   | Grass   | 3     | 8    | 23         | 2.323 | 0.217                 | 194.46  | 410.06         | 251.60      | 983.7   |
|          |         |       |      | Mean       | 2.386 | 0.223                 | 198.24  | 434.87         | 247.63      | 1173.98 |
| Arable   | Arable  | 1     | 5    | 14         | 1.723 | 0.166                 | 233.58  | 291.52         | 140.03      | 603.67  |
| Arable   | Arable  | 2     | 7    | 20         | 1.621 | 0.155                 | 127.19  | 256.28         | 126.29      | 432.25  |
| Arable   | Arable  | 3     | 8    | 24         | 1.584 | 0.169                 | 193.15  | 405.56         | 235.28      | 715.86  |
|          |         |       |      | Mean       | 1.643 | 0.163                 | 184.64  | 317.79         | 167.20      | 583.93  |
| Arable   | Fallow  | 1     | 5    | 15         | 1.585 | 0.154                 | 168.71  | 325.95         | 174.81      | 364.27  |
| Arable   | Fallow  | 2     | 7    | 19         | 1.604 | 0.152                 | 157.13  | 350.64         | 190.59      | 371.89  |
| Arable   | Fallow  | 3     | 8    | 22         | 1.505 | 0.150                 | 201.16  | 447.40         | 216.15      | 449.17  |
|          |         |       |      | Mean       | 1.565 | 0.152                 | 175.67  | 374.66         | 193.85      | 395.1   |
| Fallow   | Grass   | 1     | 1    | 2          | 1.463 | 0.128                 | 134.58  | 331.83         | 209.57      | 705.15  |
| Fallow   | Grass   | 2     | 2    | 6          | 1.507 | 0.141                 | 225.71  | 437.72         | 264.38      | 585.13  |
| Fallow   | Grass   | 3     | 3    | 9          | 1.326 | 0.120                 | 213.51  | 374.96         | 206.77      | 494.2   |
|          |         |       |      | Mean       | 1.432 | 0.130                 | 191.27  | 381.50         | 226.91      | 594.84  |
| Fallow   | Arable  | 1     | 1    | 1          | 1.125 | 0.108                 | 100.68  | 229.42         | 139.56      | 479.84  |
| Fallow   | Arable  | 2     | 2    | 5          | 0.934 | 0.097                 | 118.74  | 237.02         | 135.56      | 503.87  |
| Fallow   | Arable  | 3     | 3    | 7          | 1.101 | 0.101                 | 112.59  | 240.20         | 133.63      | 367.60  |
|          |         |       |      | Mean       | 1.053 | 0.102                 | 110.67  | 235.55         | 136.25      | 450.44  |
| Fallow   | Fallow  | 1     | 1    | 3          | 0.908 | 0.094                 | 98.82   | 195.51         | 124.95      | 262.5   |
| Fallow   | Fallow  | 2     | 2    | 4          | 0.777 | 0.084                 | 140.71  | 335.48         | 229.02      | 263.9   |
| Fallow   | Fallow  | 3     | 3    | 8          | 0.919 | 0.095                 | 139.76  | 329.90         | 175.98      | 265.82  |
|          |         |       |      | Mean       | 0.868 | 0.091                 | 126.43  | 286.96         | 176.65      | 264.10  |

## 7 Table S2

The analysis of variance (ANOVA) table for the soil organic C (SOC), total N (N) and extracellular polymeric substances (EPS) concentrations (protein, polysaccharide and uronic acid), and stable aggregate mean weight diameter (MWD) variates with the structures outlined in equations 1-2, showing the effect of previous (P) and current (C) land uses and their interaction. The table gives the degrees of freedom (df) associated with the factor (first number) and its residual (comma separated), together with the variance ratio statistic (F) and its probability level (p). Where significant at p<0.05, the standard error of differences (SED) and the least significant differences (LSD) of means are given. Note that some variates were firstly transformed by  $\log_{10}$  to normalise the distribution of residuals.

| Variate             | Unit                  | Factor       | df   | F      | р      | SED   | LSD   |
|---------------------|-----------------------|--------------|------|--------|--------|-------|-------|
| SOC                 | (log <sub>10</sub> )% | Р            | 1,2  | 57.42  | 0.017  | 0.023 | 0.100 |
|                     |                       | С            | 2,12 | 35.47  | <0.001 | 0.024 | 0.053 |
|                     |                       | $P \times C$ | 2,12 | 0.01   | 0.988  |       |       |
| N                   | (log <sub>10</sub> )% | Р            | 1,2  | 26.76  | 0.035  | 0.030 | 0.129 |
|                     |                       | С            | 2,12 | 24.66  | <0.001 | 0.024 | 0.052 |
|                     |                       | $P \times C$ | 2,12 | 0.01   | 0.988  |       |       |
| EPS- protein        | µg g <sup>−1</sup>    | Р            | 1,2  | 0.06   | 0.823  |       |       |
|                     |                       | С            | 2,12 | 6.23   | 0.014  | 14.3  | 31.1  |
|                     |                       | $P \times C$ | 2,12 | 0.54   | 0.597  |       |       |
| EPS- polysaccharide | µg g <sup>−1</sup>    | P            | 1,2  | 2.09   | 0.285  |       |       |
|                     |                       | С            | 2,12 | 8.38   | 0.005  | 23.0  | 50.0  |
|                     |                       | $P \times C$ | 2,12 | 2.40   | 0.133  |       |       |
| EPS- uronic acid    | µg g <sup>−1</sup>    | Р            | 1,2  | 10.12  | 0.086  |       |       |
|                     |                       | С            | 2,12 | 10.51  | 0.002  | 12.9  | 28.2  |
|                     |                       | $P \times C$ | 2,12 | 3.29   | 0.073  |       |       |
| MWD                 | (log₁₀)µm             | Р            | 1,2  | 5.81   | 0.137  |       |       |
|                     |                       | С            | 2,12 | 131.49 | <0.001 | 0.031 | 0.067 |
|                     |                       | P×C          | 2,12 | 4.50   | 0.035  | 0.069 | 0.155 |

## Table S3

Regression statistics of stable aggregate mean weight diameter ( $(\log_{10})$  µm) as a linear function of soil organic C (SOC; %) and soil extracellular polymeric substances (EPS) concentrations (protein, polysaccharide and uronic acid; µg EPS  $g^{-1}$  soil). The table gives the constant (a) and coefficient (b) of the linear regression (y = a + bx), the variance ratio statistic (F), the probability level associated with the regression (p), the degrees of freedom (df) and mean square error (MSE) of the residual, and the adjusted proportion of the variance accounted for by the fit (Adjusted  $R_2$ ). Note that for uronic acid, the adjusted  $R_2$  is not calculable as the residual variance exceeded the variance of the response variate.

| Statistic               | SOC       | EPS           |                |                |  |  |  |  |
|-------------------------|-----------|---------------|----------------|----------------|--|--|--|--|
|                         |           | protein       | polysaccharide | uronic acid    |  |  |  |  |
| Constant                | 2.29±0.07 | 2.17±0.18     | 2.26±0.23      | 2.64±0.21      |  |  |  |  |
| Coefficient             | 0.26±0.03 | 0.0036±0.0010 | 0.0016±0.0007  | 0.0008±0.0011  |  |  |  |  |
| F                       | 67.49     | 12.21         | 5.56           | 0.55           |  |  |  |  |
| p                       | < 0.001   | 0.002         | 0.027          | 0.465          |  |  |  |  |
| df                      | 25        | 25            | 25             | 25             |  |  |  |  |
| MSE                     | 0.020     | 0.051         | 0.062          | 0.074          |  |  |  |  |
| Adjusted R <sup>2</sup> | 0.719     | 0.301         | 0.149          | not calculable |  |  |  |  |

## **Declarations of interest statement**

Soil organic carbon, extracellular polymeric substances (EPS), and soil structural stability as affected by previous and current land-use

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