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# Effects of plant pathogens on population dynamics and community composition in grassland ecosystems: two case studies

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**Abstract** Grassland ecosystems comprise a major portion of the earth's terrestrial surface, ranging from high-input cultivated monocultures or simple species-mixtures to relatively unmanaged but dynamic systems. Plant pathogens are a component of these systems with their impact dependent on many interacting factors, including grassland species population dynamics and community composition, the topics covered in this paper. Plant pathogens are affected by these interactions and also act reciprocally by modifying their nature. We briefly review these features of disease in grasslands and then introduce the 150-year long-term Park Grass Experiment (PGE) at Rothamsted Research in the UK. We then consider in detail two plant-pathogen systems present in the PGE, *Tragopogon pratensis*-*Puccinia hysterium* and *Holcus lanata*-*Puccinia coronata*. These two systems have very different life history characteristics: the first, a biennial member of the Asteraceae infected by its host-specific, systemic rust; the second, a perennial grass infected by a host-non-specific rust. We illustrate how observational, experimental and modelling studies can contribute to a better understanding of population dynamics, competitive interactions and evolutionary outcomes. With *Tragopogon pratensis*-*Puccinia hysterium*, characterised as an "outbreak" species in the PGE, we show that pathogen-induced mortality is unlikely to be involved in host population regulation; and that the presence of even a short-lived seed-bank can affect the qualitative outcomes of the host-pathogen dynamics. With *Holcus lanata*-*Puccinia coronata*, we show how nutrient conditions can affect adaptation in terms of host defence mechanisms, and that co-existence of competing species affected by a common generalist pathogen is unlikely.

**Keywords** grassland species dynamics, pathogen dynamics, pathogen-induced mortality, seed bank, nutrient supply, competitive interactions, evolution of host defence

## Introduction

Grassland comprises a major component of global terrestrial ecosystems: excluding Greenland and Antarctica, perhaps some 40% of the earth's terrestrial surface. Grassland evolution can be considered a biological force for global change through the co-evolution of grasses and their grazers over geological time (Retallack, 2013). Grassland ecosystems range from simple cultivated grasslands used as rotational or semi-permanent leys for grazing livestock to relatively undisturbed or intentionally-conserved systems. Biodiversity in grassland ecosystems can be low, as in monocultures or simple grass/legume mixtures, or high as in extensive pastoral range-lands or

1 protected reserves such as the US prairies. Throughout this spectrum of land use there are  
2 associated communities of organisms, with varying levels of diversity depending on geography and  
3 history. These communities include other flora, microbes, invertebrates, and vertebrates including  
4 mammals, reptiles, and birds, all of which interact in various ways and with surrounding patterns of  
5 land use, e.g., agricultural systems and forestry.  
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8 Much of the scientific literature on the impact of pathogens and pests on plant populations and  
9 communities and factors which mitigate the impact has come from work with forage production and  
10 grazing lands; for example, Nelson and Burns (2006) reviewed the advances in grassland science over  
11 the last 50 years and documented how management has benefitted from advances in disease  
12 resistance and an understanding the role of endophytic fungi. There has been little consideration of  
13 the impact of pathogens in relatively unmanaged grassland ecosystems, except in the context of  
14 global/climate change and biodiversity (Chakraborty 2001; Pautasso et al. 2010). Many reviews have  
15 appeared at regular intervals that document the impacts of pathogens on natural or unmanaged  
16 plant populations and communities in general (e.g., Dinooor and Eshed, 1984; Kranz, 1990; Alexander  
17 & Holt, 1998; Alexander, 2010; Burdon et al., 2013, amongst others) but, despite many specific  
18 experimental studies, there appears to be no major overview that integrates the body of knowledge  
19 on grassland diseases obtained from these studies. Such a comprehensive review is beyond the  
20 scope of the present paper but selected examples of experimental studies that illustrate the wealth  
21 of information available on different aspects are given in Table 1.  
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27 Plant pathogens (fungi, oomycetes, bacteria, and viruses) are present in all grassland ecosystems.  
28 They may directly alter population dynamics of individual plant species and the competitive effects  
29 affecting community composition in grasslands. We give examples of research carried out on these  
30 two topics and then look in more detail at two specific examples: the grass *Holcus lanatus* and the  
31 composite *Tragopogon pratensis*, as affected by rust pathogens, but with differing life history  
32 characteristics in both host and the rust species. We do this in the context of long-term experiments  
33 on nutrient regimes affecting the population dynamics of host species in the Park Grass Experiments  
34 (PGE) at Rothamsted Research, UK, and for each example we consider observational, modelling and  
35 experimental approaches which manipulate the interactions involved.  
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### 43 **Effects of pathogens on plant population dynamics**

#### 44 Disease development in relation to plant host characteristics

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46 Disease development in grassland species can be influenced by host reproduction characteristics,  
47 morphology, and vegetative development. Systemic disease spread has been suggested as a  
48 disadvantage for clonal growth patterns in grasses. A mathematical model has been constructed to  
49 determine whether systemic disease decreases the competitive ability of highly integrated clonal  
50 plants when compared with less integrated ones (Koubek and Herben 2008). The model was  
51 constructed to represent grass ramets in short-turf grasslands, such as *Festuca rubra*. Although the  
52 same infection rate caused more systemic disease in the highly integrated plants, the disease had  
53 only limited potential to select for a less integrated strategy due to the advantages in clonal  
54 integration, such as greater translocation ability and resource sharing.  
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1 Fungal disease affecting seed has rarely been quantified in grassland ecosystems. Seed mortality of  
2 four prairie species (two grasses and two forbs) was studied in relation to causal factors (Clark and  
3 Wilson 2003). "Natural" mortality not associated with fungal disease or vertebrate predation (44-  
4 80%) was the greatest factor, much greater than fungal disease (< 10%). Vertebrate and invertebrate  
5 seed predation (not directly measured) was important only in single species.  
6

7 The effects of insect herbivores (by insecticide) and fungal pathogens (by fungicide) exclusion on  
8 population-level seed production of four grass species was studied over two years in Minnesota  
9 (Dickson and Mitchell 2010). There were no interactive effects but seed production of all four  
10 species was affected by either herbivore or pathogen exclusion. Reduced seed production by  
11 dominant species may promote co-existence with sub-dominant species through competition-  
12 colonization trade-offs. Seed yield losses are an important consideration in managed grasslands  
13 (Pfender and Alderman, 2006).  
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17 Fungal endophyte interactions with host plants have a range of effects, both beneficial and  
18 detrimental, on host reproduction and development. The fungal endophyte *Epichloë bromicola*  
19 causes choke disease on *Bromus erectus*. A 3-year field experiment was conducted at a site in the  
20 Swiss Jura Mountains where, overall, 4.3% of tillers had diseased inflorescences under conditions of  
21 small-scale habitat fragmentation (Groppe et al. 2001). After fragmentation, disease incidence  
22 increased at the level of the individual plant (genet), probably due to increased horizontal  
23 transmission of disease, but at the level of the tiller disease incidence was decreased due to an  
24 increase in healthy tillers. Species of *Epichloë* infect *Holcus lanatus* and *H. mollis*, but insecticide-  
25 exclusion experiments in grassland sites at Silwood Park in the UK gave contrasting results for these  
26 two grass species (Clay and Brown 1997): insect herbivory seemed to increase infection in *H. mollis*,  
27 but decrease it in *H. lanatus*. Leaf wounds caused by insects were important sites for infection of  
28 orchard grass by *Epichloë typhina* (Alderman, 20013), and slugs are also thought to play a potential  
29 role in both stroma fertilisation and disease spread (Hoffman and Rao, 2013).  
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36 One of the longest studied grassland communities is the Park Grass experiment (PGE) at Rothamsted  
37 Research, Harpenden, UK (Silvertown et al. 2006) (Fig.1). Since 1856, surveys of the plant  
38 community structure have been undertaken regularly (also soil microbe responses: see Rousk et al.  
39 2011). As a consequence, long term assessments of the population dynamics of grassland species  
40 have been possible. Dodd et al (2005) characterised species as increasing (e.g. *Trifolium pratense*),  
41 decreasing (e.g. *Veronica chamaedrys*), fluctuating (e.g. *Conopodium majus*), or following a  
42 sporadic outbreak pattern (e.g. *Tragopogon pratensis*). Many of these patterns in dynamics were  
43 originally assumed to be a result of the fertiliser treatment regimes applied to the plots or to be  
44 general population changes. The effect of plant pathogens was not considered. Although the  
45 surveys provide good data relating to host plant species incidence, there are no records of disease.  
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51 Case study: *Tragopogon pratensis* – *Puccinia hysteriorum*  
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53 *Tragopogon pratensis* is a biennial (Qi et al. 1996) member of the Asteraceae that occasionally  
54 demonstrates perennial characteristics (Jorritsma-Wienk et al. 2007) and is host to the autecious  
55 demicyclic rust *Puccinium hysteriorum*. The life cycle is shown schematically in Fig. 2. Basidiospores  
56 infect first year seedlings but symptoms are not observed until the second year as the rust becomes  
57 systemic and overwinters in the underground stock. As the second-year plant emerges, aecia are  
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1 produced on the stem and leaves. Aeciospores are produced but play no role in the population  
2 dynamics, as the second year plants die following seed set. The sequence of events from teliospore  
3 to basidiospore and infection of seedlings has not been investigated.  
4

5 Over the period 2005-2008, observations on the incidence of *P. hystereum* were made in the PGE.  
6 Rust infected plants produced fewer seeds and fewer of these germinated (Salama et al 2010).  
7 Overall, *P. hystereum* suppresses potential reproduction of *T. pratensis* by a factor of about 30: an  
8 infected individual will on average leave two germinated offspring, whereas a healthy individual may  
9 produce up to 75 seedlings. Additionally, infected plants produced shorter seeds with longer pappi.  
10 However, once germinated, pot experiments gave no evidence for additional seedling mortality due  
11 to pathogen infection (Salama et al 2012). A similar reduced seed production occurs in groundsel  
12 (*Senecio vulgaris*) infected by the rust, *P. lagenophorae* (Paul and Ayres 1987), and altered seed  
13 morphology has been observed in cases of *Lolium multiflorum* infected with *P. coronata* (Mattner  
14 and Parbery 2007) and *P. hordei* infecting *Hordeum vulgare* (Das et al. 2007).  
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20 A discrete time epidemiological model was developed (Salama et al 2012) which compartmentalised  
21 the host population as: susceptible (*S*) seedlings, exposed/latently infected (*E*) seedlings, infectious  
22 (*I*) second-year plants, and healthy second-year plants (*R*) which no longer contribute to the epidemic  
23 (even if infected within-year they do not develop symptoms or contribute to the infection of  
24 seedlings). The model captured the essential characteristics of a biennial host and systemic  
25 pathogen. The model contained parameters for: seedling recruitment (*a*); density dependence of  
26 seedling emergence ( $\lambda$ ); within (*b*) and between (*d*) year mortality of the host; a Poisson distribution  
27 function for pathogen transmission (*c*); and the probabilities of disease expression (*p*) and pathogen  
28 induced mortality ( $\beta$ ).  
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33 The model was also adapted to account for variable pathogen-induced mortality and a trade-off  
34 between transmissibility and pathogen-induced mortality. For many parameter combinations the  
35 frequency of the host varied in a way reminiscent of the observed irregular outbreak dynamics of *T.*  
36 *pratensis* in the PGE (Salama and Jeger, 2013). Other parameter combinations produced steady-  
37 state host populations free of disease, cyclical population dynamics of diseased and healthy hosts,  
38 and extinction of both the host and pathogen (Salama et al. 2012).  
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42 Disease persists in the model population if each infected individual produces at least one other  
43 infected individual: that is, if the basic reproductive number  $R_0 > 1$ .  $R_0$  is determined by a formula  
44 including parameters representing pathogen ecology (transmissibility, the probability of disease  
45 expression and pathogen-induced mortality) and host ecology (background mortality and host  
46 population size). Pathogen-induced mortality directly influences the host population dynamics. It  
47 was possible to derive a recurrence relationship between the number of healthy second-year plants  
48 and the number of healthy seedling plants two years previously. This relationship was fitted to  
49 observational data collected at the PGE (1995 – 2008) (Salama et al. 2012). However, disease-  
50 induced mortality hardly affected the goodness of fit. Furthermore, the model suggested that in the  
51 field, density-dependence in seedling emergence hardly affected the dynamics. Overall, the  
52 observational data, pot experiments and model analysis suggests that the dynamics of *T. pratensis* in  
53 the PGE results from a moderately performing host exposed to a highly transmissible strain of *P.*  
54 *hystereum* which has a low probability of surviving the systemic phase of its life-cycle, and causes  
55 little additional mortality in diseased compared with healthy plants.  
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1 The model above ignores the possible existence of a long-lived seed-bank. For example, in other  
 2 systems the species *Linanthus parryae* has a vast seed-bank where seeds remain viable up to six  
 3 years (Kaj et al. 2001) and *Brassica napus* up to ten years (Pekrun et al. 2005). This in turn can  
 4 increase population size, modulate changes in genetic variation (Nunney 2002) and also modify age  
 5 structure within the population (Evans and Dennehy 2005). Seed-banks have an impact on plant  
 6 populations by acting as reservoirs of genetic material that transfer across generations, so retaining  
 7 genes that would otherwise be lost by drift (Waples 2006). A seed-bank allows for the recruitment of  
 8 individuals irrespective of biotic and abiotic factors which may have affected the previous year's  
 9 adult stock (Waples 2006), so potentially altering host population dynamics.

12 There is some evidence for a small seed-bank for *T. pratensis* (Clements et al. 1999) remaining  
 13 viable for up to two years (Roberts 1986) but sensitive to anaerobic conditions and temperatures  
 14 above 25°C which reduce seed viability. A seed-bank exists for the related cultivated species *T.*  
 15 *porrifolius* (purple salsify) (Haubensak and Smyth (2002). A seed-bank was therefore included in  
 16 the model above (Salama et al. 2012, Salama and Jeger 2013) and the resulting dynamics  
 17 investigated. The seed-bank was added as an extra compartment (Fig. 3). This was considered to be  
 18 reasonable (rather than using a full age structured matrix: Caswell 1989) where the seed-bank is  
 19 likely to be short lived and have minimal cohort structure. The seed-bank phase ( $B$ ) receives a  
 20 constant number of seeds ( $a_1$ ) from each uninfected adult above ground. A proportion of the below-  
 21 ground seed-bank ( $a_0$ ) emerges as  $S$  individuals each season. Below-ground mortality occurs in the  
 22 seed-bank both within ( $b_0$ ) and between season ( $d_0$ ), whilst background mortality is also experienced  
 23 by above ground individuals within ( $b_1$ ) and between season ( $d_1$ ). Exposed and infected individuals  
 24 experience pathogen-induced mortality ( $\beta$ ) as before.

31 The revised model can be represented by the following equations which describe the population at  
 32 the beginning of a growing season. Note that, as in the basic model, there is no exposed class at the  
 33 start of a season (see Salama 2009 for derivation):

$$36 B_{t+1} = [(1-d_0)(1-a_0)] \left[ (1-b_0)B_t + \frac{a_1(1-b_1)R_t}{1+(1-b_1)\lambda R_t} \right]$$

$$37 S_{t+1} = a_0 \left[ (1-b_0)B_t + \frac{a_1(1-b_1)R_t}{1+(1-b_1)\lambda R_t} \right]$$

$$40 I_{t+1} = (1-b_1)(1-d_1)(1-\beta)pS_t[1 - e^{-c(1-b_1)(1-\beta)I_t}]$$

$$41 R_{t+1} = (1-b_1)(1-d_1)S_t \{1 - p[1 - e^{-c(1-b_1)(1-\beta)I_t}]\}$$

53 By rearranging the model equations in the absence of disease it is possible to estimate the steady  
 54 state populations for the seed-bank and non-infected plants. We can then derive an expression  
 55 describing the conditions required for disease to invade a healthy population:

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$$\frac{pc(1-\beta)^2}{\lambda} \left\{ \frac{a_0 a_1 (1-b_1)^2 (1-d_1)}{1-(1-d_0)(1-a_0)(1-b_0)} - 1 \right\} > 1$$

This condition involves all of the model parameters. An implicit steady-state equation for infectious individuals ( $I^*$ ) can be derived as in the basic model (Salama et al. 2012), but this is complex and is not shown.

Using the model framework simulations can be undertaken for a range of parameter values (Table 2) with and without seed-banks. In the case without a seed-bank, host populations may go extinct, tend to a stable population, or undergo continual population cycling with some of the population remaining diseased (Fig.4). Similar patterns also occur when there is a trade-off between transmissibility and pathogen-induced mortality. However, with a seed-bank it appears that extinction is avoided for parameter combinations that would otherwise cause the pathogen to make the host extinct. Fig. 5 demonstrates that under most conditions the host populations tend to a steady population size with disease endemic. Under conditions where a moderate or high performing host population is exposed to a highly transmissible strain which causes little pathogen-induced mortality, the host may persist while the disease goes extinct. In systems without a seed-bank, for the same parameter values the pathogen population will cycle indefinitely (Fig. 4). Overall, therefore, the seed-bank is a stabilising factor in the population dynamics of both host and pathogen.

The *T. pratensis* – *P. hysteriorum* system in the PGE demonstrates that it is possible for rust pathogens to significantly alter the recruitment of a host plant by reducing the production and viability of seed. The simulations highlight that, depending on conditions, host populations may be stable with or without a persistent pathogen population, may cycle, or may be driven extinct by the pathogen. However, a seed-bank makes it unlikely that the pathogen will drive the host extinct, despite its dramatic effect on fecundity. These findings are consistent with the episodic appearance of an outbreak host species above ground.

## Effect of pathogens on plant community composition

### Diversity, competition and co-existence in grassland communities

The evidence that plant species diversity decreases the severity of plant diseases is strengthening in agricultural systems, but is still sparse for complex plant communities. Roscher et al. (2007) varied the levels of species richness and functional group richness in experimental grassland communities. Different cultivars of *Lolium perenne* were used to identify effects of plant and cultivar diversity on the incidence and severity of *Puccinia coronata* and *P. graminis*. Cultivar monocultures showed strong differences in disease severity. Disease severity of both rusts decreased with increasing species richness and populations of both retained a high level of host specificity. Differences in cultivar susceptibility persisted in the polycultures independent of diversity treatment and host density, but there were cultivar-specific patterns of disease across the species richness gradient.

A long-term manipulative field experiment was used to investigate how environmental factors, fertilization and plant community diversity affect plant traits and herbivore/pathogen damage in the



1 common prairie legume *Lespedeza capitata* (Lau et al. 2008). Plants growing in polyculture were  
2 damaged considerably less by specialist herbivores and by pathogens, but had five times more  
3 damage from generalist herbivores than when growing in monocultures. Differences in plant traits  
4 partly explained the differences in herbivory in relation to diversity but did not explain the  
5 differences in disease incidence between monocultures and polycultures.  
6

7 In a prospective study of alpine grasslands being converted to agricultural use, observations were  
8 made of the damage caused by herbivory and fungal pathogen infection in relation to plant  
9 functional group (legumes, herbs and grasses), land use (mowed, grazed or abandoned) and altitude  
10 (Fischer et al. 2012). Damage by pathogens was independent of land use and altitude but pathogen  
11 diversity decreased with fertilization. There was no correlation between damage caused by  
12 herbivory or fungal disease with plant species diversity.  
13

14 Two introduced cultivated genotypes of *Lolium perenne* were introduced into semi-natural grassland  
15 to investigate whether a genotype selected for resistance to *Puccinia coronata* showed better  
16 establishment than a susceptible genotype (Erneberg et al. 2008). Introduced genotypes had  
17 greater shoot biomass in the semi-natural plant communities when exposed to pathogen attack, but  
18 showed the expected reduction in growth when inoculated onto monoculture pots in a controlled  
19 setting.  
20

21 Much has been learnt by a programme of research in Minnesota since the early 1990s on foliar  
22 fungal pathogens in grassland communities. This research has resulted in papers describing diversity-  
23 productivity relationships (HilleRis Lambers et al. 2004), CO<sub>2</sub> and nitrogen deposition (Mitchell et al,  
24 2003), net production (Mitchell, 2003), species diversity, abundance and composition (Mitchell et al.  
25 2002; Knops et al. 1999) and other topics which cannot be reviewed within the scope of this paper.  
26 However, it remains true that little is known about the role of foliar fungal pathogens as drivers of  
27 plant community composition and diversity in grasslands. Pesticides may have non-target effects.  
28 Most workers attempt to allow for these by testing their effects on yield in potted plants; clearly a  
29 significant result in such an experiment means the pesticide's effect on a community needs to be  
30 interpreted with care. However, a non-significant result does not mean important side-effects are  
31 absent: effects may be undetectable due to inter-replicate variation, may occur only under particular  
32 stresses, or may only come into play in competition with other species. Peters and Shaw (1996)  
33 manipulated rust severity on *Holcus lanatus* in a replicated plot experiment using early succession  
34 communities created by tillage of existing grassland. Although there were indirect effects of  
35 fungicide, it was also possible to show that more severe rust reduced the dominance of the grass  
36 and increased the abundance of forbs in the community. Similarly, Allan et al. (2010) used fungicide  
37 to exclude pathogens from perennial grassland, as well as vertebrate and invertebrate herbivores. It  
38 was possible to quantify their effects compared with the herbivores. Foliar fungal pathogens  
39 substantially reduced above-ground plant biomass and promoted plant diversity, similar in effect to  
40 the other herbivores but with differential responses of plant species.  
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42 A stochastic model was developed to predict the effect on competing species in plant communities  
43 of a shared natural enemy that limits recruitment (Schoolmaster 2008). The interactions between  
44 natural enemies and recruitment limitation led to some counter-intuitive results. For example, in a  
45 competitive interaction it is neither necessary nor sufficient for a shared natural enemy to reduce  
46 growth of the better competitor more than the poorer competitor for the presence of the enemy to  
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1 have an overall positive effect on the poorer competitor. This applies especially where there is short  
2 scale dispersal and fecundities are low, which emphasises the importance of considering spatial  
3 processes in models of plant community dynamics. Host range differences between populations of  
4 pathogen species may also be important, for example with *Puccinia graminis* subsp. *graminicola*  
5 populations from perennial ryegrass and tall fescue (Pfender, 2001).  
6

7 A spatiotemporal model of the transmission dynamics of cereal yellow dwarf viruses was used to  
8 examine the effects of seasonality and plant competition, and whether the suite of viruses can  
9 persist across a range of plant community spatial structures (Moore et al. 2011). The ability of a  
10 pathogen to invade a fragmented community with different levels of patch connectivity was  
11 examined, as was the extent to which a pathogen could facilitate the establishment of a newly  
12 arrived host species. Connectivity interacts with the arrival time and host “infection tolerance” in  
13 determining whether establishment will occur.  
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17 These examples are of foliar pathogens. Competitive effects affecting plant populations are claimed  
18 to maintain diversity in plant populations particularly strongly through “Janzen-Connell effects”,  
19 where these are widespread and sufficiently strong enough (Fitzsimons and Miller, 2010; Petermann  
20 et al., 2008) to promote species co-existence by preventing the continued occupation of a site by  
21 one species. The general mechanism proposed for this is the build up of soil-borne pathogens which  
22 reduces the competitive ability of single species and balances out differences in individual fitness  
23 among species. An example has been claimed in the sand-dune grass *Vulpia ciliata* (Carey et al.,  
24 1992); the phenomenon of “clover decline” or “clover sickness” (also a subject of a long-term  
25 experiment in the early days of Rothamsted) is due to a variety of root pathogens and pests and  
26 would occur in dense stands in natural as well as planted grassland.  
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32 Grassland ecosystems function as dynamic plant communities, even in the simplest case of binary  
33 grass-legume mixtures. In these communities the competitive outcomes between species can be  
34 modified, even reversed, in the presence of pathogens affecting one or more of the component  
35 species. Plant species may adapt in their resistance levels to pathogens with a trade-off involving a  
36 reduction in growth rate (van den Berg and van den Bosch 2009). This has implications for co-  
37 existence of competing plant species, as discussed in the case study of *Holcus lanata-Puccinia*  
38 *coronata* .  
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#### 43 Case study : *Holcus lanatus* – *Puccinia coronata*

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45 The evolution in grasses of traits due to selection by pathogen attack is likely to alter competitive  
46 outcomes. Apical leaf necrosis or leaf shedding has been modelled (van den Berg and van den Bosch,  
47 2004) as an adaptive response to disease pressure. Infection-driven leaf shedding was shown to be  
48 a more efficient response to pathogen attack than constant leaf shedding. Both strategies imply  
49 increased shedding rates if nutrients are more available. Which strategy will evolve depends on the  
50 balance of autoinfection (infections on the same plant) and alloinfection (infection of other plants).  
51 Study of these strategies was further developed by van den Berg et al. (2007). When nutrient  
52 availability is high, and assuming plants are more susceptible as a consequence, there will be a  
53 higher N drain by the pathogen and hence faster apical leaf necrosis. For a biotrophic pathogen this  
54 will reduce the disease load as the pathogen cannot grow on necrotic tissue. For apical leaf necrosis  
55 to be a successful defence mechanism the benefits of disease reduction by early leaf necrosis must  
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1 be greater than the costs of losing healthy leaf tissue, the source of assimilates for plant growth.  
2 These costs should be lower if exogenous nutrients are more freely available.

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4 Experimental work was then conducted with the grass *Holcus lanatus*, a common species in the PGE  
5 (van den Berg et al. 2008). Seeds of the grass and urediniospores of *Puccinia coronata*, a rust with  
6 many graminaceous hosts, were collected from adjacent plots in the PGE, similar in habitat but with  
7 differing N-fertilization regimes that had been applied for 150 years. In a factorial design with all  
8 combinations of treatments present, seeds from each plot were grown in pots under regimes similar  
9 to one of the original nutrient conditions and inoculated with urediniospores collected from one of  
10 the original plots. The time of disease onset was noted and the population growth rate on leaves  
11 calculated. The experiment was repeated in two years. With higher nutrient availability, disease  
12 onset was earlier and the population growth rate was higher, but the size of these effects depended  
13 on both pathogen and seed origin. Higher nutrient availability also led to a reduced time between  
14 disease onset and apical leaf necrosis onset (assumed to result from pathogen attack), with some  
15 indication that necrosis mechanisms were influenced by the origin of the seed and pathogen. The  
16 findings were consistent with apical leaf necrosis being a defence mechanism.

17  
18 The implications of the evolution of adaptive traits such as defence mechanisms for co-existence of  
19 plant species were further analysed by van den Berg and van den Bosch (2009). They studied a  
20 model, shown schematically in Fig.6, for the dynamics of two plant species competing for space that  
21 are both infected with a generalist pathogen. The model is governed by the equations

$$22 \frac{dX}{dt} = r_X X - \alpha_X X^2 - \alpha_Y XY - \beta_X IX$$

$$23 \frac{dY}{dt} = r_Y Y - \alpha_Y Y^2 - \alpha_X XY - \beta_Y IY$$

$$24 \frac{dI}{dt} = \beta_X IX + \beta_Y IY - \omega I$$

25 Here  $X$  and  $Y$  are the densities of the two plant species and  $I$  is the total density of infected plants.  
26 The intrinsic rate of increase of the densities  $X$  and  $Y$  of the plant species are denoted by  $r_X$  and  $r_Y$ ,  
27 respectively. The competitive ability of plant species  $X$  and  $Y$  are measured by  $\alpha_X$  and  $\alpha_Y$ ,  
28 respectively. The generalist plant pathogen has infection parameters  $\beta_X$  and  $\beta_Y$  on species  $X$  and  $Y$ ,  
29 respectively. Plants remain infectious for, on average,  $1/\omega$  time units. The model was analysed first  
30 with fixed parameters, ignoring the evolution of defences. It predicts that:

- 31  
32 (i) In the absence of the pathogen the two plant species do not co-exist and the plant  
33 species with the largest rate of increase outcompetes the other plant species. This is a  
34 well known result for competition for space.  
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36 (ii) In the presence of disease there is a wide range of parameters for which the two plant  
37 species can co-exist.

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39 These are well-established properties for species that compete and are infected by a generalist  
40 disease, as discussed above. Next, the consequences of variation, allowing the defensive capacity of  
41 the plant species to evolve, were investigated. It is reasonably well established (e.g. Ruuhola and  
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1 Julkunen-Tiitto 2003; Way et al. 2002) that there is often a trade-off between defence against  
2 pathogen infection and the growth rate of plants. This trade-off is incorporated into the model using  
3 a simple energy budget calculation (Fig. 7) resulting in a hyperbolic relation between plant growth  
4 rate and infection efficiency of the pathogen:  
5

$$6 \quad \beta_i = \beta_{0i} e^{-A_i (1-B_i r_i)}$$

7  
8 Here the subscript  $i$  denotes plant species  $X$  or  $Y$ .  $A_i$ ,  $B_i$  and  $\beta_{0i}$  are parameters in the energy budget  
9 model. In the model, the defence capacity of both species evolves over time subject to the  
10 condition that increasing defence decreases the plant species growth rate, and so its capacity to  
11 compete for space. Surprisingly, for any set of parameter values for which the plant species can co-  
12 exist in the static-trait ecological model, plant defences evolve such that the species can no longer  
13 co-exist.  
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17 The conclusion from these modelling studies is therefore that - despite frequency dependent  
18 infection by generalist pathogens playing an important role in promoting plant species coexistence  
19 on an ecological time scale - long term species coexistence as observed in many natural plant  
20 communities cannot be explained by such a mechanism (van den Berg and van den Bosch, 2009).  
21 The question is then what mechanisms may lead to so many plant species coexisting. In practice,  
22 completely generalist pathogens are unusual: in many cases cryptic or partial specialisation on  
23 particular hosts exists, even below species level. This is true, for example, in the *Holcus – Puccinia*  
24 *coronata coronata* system (Dawkins, 1988). It might therefore be useful to extend these models to  
25 include the possibility of evolution in the pathogens, driven by trade-offs between pathogenicity on  
26 the two hosts.  
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## 32 33 34 **Conclusions**

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36 This paper does not survey the full range of grassland ecosystems across a range of temporal and  
37 spatial scales. However the selected examples and case studies presented in this paper allow for  
38 some conclusions to be drawn about the relationship between plant pathogens, host population  
39 dynamics and diversity, and disease dynamics.  
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- 42 1. Studies on grassland diseases benefit from combining observational, experimental and  
43 modelling approaches.
- 44 2. Disease-induced mortality is not a necessary condition for host population regulation by plant  
45 pathogens.
- 46 3. The existence of a seed bank affects the qualitative as well as quantitative dynamics of natural  
47 plant populations. This aspect has been relatively understudied in field investigations of the role  
48 of disease in such populations.
- 49 4. Adaptation and evolutionary change in response to disease alter the ecology and population  
50 dynamics of individual species over longer time-scales, and therefore alter community  
51 composition and levels of biodiversity in grasslands.  
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5. Most attempts to improve grasslands for human purposes involve fertilization. This alters immediate disease levels, and the trajectory of longer-term evolution in response. However, the effects of fertilisation on disease dynamics, plant adaptive responses and grassland species diversity are poorly understood and deserve more study.
  6. Co-existence of plant species in natural plant populations on an evolutionary time scale appears not to be wholly explained by mechanisms such as frequency-dependent infection and competitive interactions , although these can be shown to be effective on ecological time scales.
  7. A greater dialogue between plant pathologists concerned mostly with disease in managed plant populations and ecologists concerned mostly with disease in natural unmanaged systems would be to the mutual benefit of both. Grasslands, because of the continuum in a broad range of management inputs, make an ideal system for this interchange to take place.

## 18 References

- 19 Alderman, S.C. (2013). Survival, germination, and growth of *Epichloë typhina* and significance of leaf wounds and insects in infection of orchardgrass. *Plant Disease*, 97, 323-328.
- 22  
23 Alexander, H.M. (2010). Disease in natural plant populations, communities, and ecosystems: insights into ecological and evolutionary processes. *Plant Disease*, 94, 492-503.
- 26  
27 Alexander, H. M. & Holt, R. D. (1998). The interaction between plant competition and disease. *Perspectives in Plant Ecology, Evolution and Systematics*, 1, 206-220.
- 29  
30 Allan, E., van Ruijven, J., & Crawley, M.J. (2010). Foliar fungal pathogens and grassland biodiversity. *Ecology*, 91, 2572-2582.
- 33  
34 Barnes, C.W. Kinkel, L.L., & Groth, J.V. (2005). Spatial and temporal dynamics of *Puccinia andropogonis* on *Comandra umbellata* and *Andropogon gerardii* in a native prairie. *Canadian Journal of Botany*, 83, 1159-2581.
- 38  
39 Burdon J.J., Thrall, P.H., & Ericson, L. (2013). Genes, communities & invasive species: understanding the ecological and evolutionary dynamics of host-pathogen interactions. *Current Opinion in Plant Biology*, in press.
- 43  
44 Booth, R.E., & Grime, J.P. (2003). Effects of genetic impoverishment on plant community diversity. *Journal of Ecology*, 91, 721-730.
- 47  
48 Borer, E.T., Hosseini, P.R., Seabloom, E.W., & Dobson, A.P. (2007). Pathogen-induced reversal of native dominance in a grassland community. *Proceedings of the National Academy of Sciences USA*, 104, 5473-5478.
- 51  
52 Borer, E.T., Seabloom, E.W., Mitchell, C.E., & Power, A.G. (2010). Local context drives infection of grasses by vector-borne generalist viruses. *Ecology Letters*, 13, 810-818.
- 55  
56 Borer, E.T., Adams, V.T., Engler, G.A., Adams, A.L., Schumann, C.B., & Seabloom, E.W. (2009). Aphid fecundity and grassland invasion: Invader life history is the key. *Ecological Applications*, 19, 1187-1196.
- 60  
61  
62  
63  
64  
65

1 Caswell, H. (1989) Matrix population models: construction, analysis, and interpretation. Sinauer  
2 Associates, Sunderland

3 Chakraborty, S. (2001). Grassland plant diseases: management and control. In Proceedings of the XIX  
4 International Grassland Congress: Grassland ecosystems: an outlook into the 21st century (Eds  
5 Gomide JA, Mattos WRS, DaSilva SC) San Pedro, Brazil, Feb 11-21 2001, pp 223-230.

6  
7  
8 Clarke, D.L., & Wilson, M.V. (2003). Post-dispersal seed fates of four prairie species. *American*  
9 *Journal of Botany*, 90, 730-735.

10  
11  
12 Clay, K., & Brown, V.K. (1997). Infection of *Holcus lanatus* and *H. mollis* by *Epichloe* in experimental  
13 grasslands. *Oikos*, 79, 363-370.

14  
15  
16 Clements, D. R., Upadhyaya, M. K., & Bos, S. J. (1999). The biology of Canadian weeds. 110.  
17 *Tragopogon dubius* Scop., *Tragopogon pratensis* L., and *Tragopogon porrifolius* L. *Canadian Journal*  
18 *of Plant Science*, 79, 153-163.

19  
20  
21 Das, M., Griffey, C. A., Baldwin, R. E., Waldenmaler, C. M., Vaghn, M. E., Price, A. M., & Brooks, W. S.  
22 (2007). Host resistance and fungicide control of leaf rust (*Puccinia hordei*) in barley (*Hordeum*  
23 *vulgare*) and effects on grain yield and yield components. *Crop Protection*, 26, 1422-1430.

24  
25  
26 Dawkins GHM, 1988. *Plant pathogens and ecological succession*. PhD thesis: University of London,  
27 Imperial College.

28  
29 de Jong, M.D., Bourdot ,G.W., & Powell, J. (2002). A model of the escape of *Sclerotinia sclerotiorum*  
30 ascospores from pasture. *Ecological Modelling*, 150, 83-105.

31  
32  
33 Delmiglio, C., Pearson, M.N., Lister, R.A., & Guy, P.L. (2010). Incidence of cereal and pasture viruses  
34 in New Zealand's native grasses. *Annals of Applied Biology*, 157, 25-36.

35  
36  
37 Dickson, T.L., & Mitchell, C.E. (2010). Herbivore and fungal pathogen exclusion affects the seed  
38 production of four common grassland species. *PLoS One*, 5, e12022.

39  
40  
41 Dinoor, A. & Eshed, N. (1984). The role and importance of pathogens in natural plant communities.  
42 *Annual Review of Phytopathology*, 22, 443-466.

43  
44  
45 Dodd, M., Silvertown, J., McConway, K., Potts, J., & Crawley, M. (1995) Community stability - a 60-  
46 year record of trends and outbreaks in the occurrence of species in the Park Grass Experiment.  
47 *Journal of Ecology*, 83, 277-285.

48  
49  
50 Erneberg, M., Strandberg, B., Strandberg, M., Jensen, B.D., & Weiner, J. (2008). Effects of soil  
51 disturbance and disease on the growth and reproduction of *Lolium perenne* (Poaceae) introduced to  
52 semi-natural grasslands. *Polish Journal of Ecology*, 56, 593-604.

53  
54  
55 Evans, M. E. K., & Dennehy, J. J. (2005). Germ banking: Bet-hedging and variable release from egg  
56 and seed dormancy. *Quarterly Review of Biology*, 80, 431-451

57  
58  
59 Finch, D.D., & Alexander, H.M. (2011). Variation in plant distributions, plant traits and disease levels  
60 across a woodland /grassland ecotone. *American Midland Naturalist*, 166, 309-324.

61  
62  
63  
64  
65

1 Firbank, L.G., Smart, S.M., Crabb, J., Critchley, C.N.R., Fowbert, J.W., Fuller, R.J., et al. (2003).  
2 Agronomic and ecological costs and benefits of set-aside in England. *Agriculture Ecosystems &*  
3 *Environment*, 95, 73-85.

4 Fischer, M., Weyand, A., Rudmann-Maurer, K., & Stocklin, J. (2012). Omnipresence of leaf herbivory  
5 and leaf infection by fungal pathogens in agriculturally used grasslands of the Swiss Alps, but low  
6 plant damage. *Alpine Botany*, 122, 95-107.

7  
8  
9  
10 Fitzsimons, M.S. & Miller, M. (2010). The importance of soil microorganisms for maintaining diverse  
11 plant communities in tallgrass prairie. *American Journal of Botany*, 97, 1937-1943.

12  
13 Frantzen, J. (1994). The role of clonal growth in the pathosystem *Cirsium arvense* – *Puccinia*  
14 *punctiformis*. *Canadian Journal of Botany*, 72, 832-836.

15  
16  
17 Goodall, J., Witkowski, E.T.F., McConnachie, A.J., & Keen, C. (2012). Altered growth, population  
18 structure, and realised niche of the weed *Campuloclinium macrocephalum* (Asteraceae) after  
19 exposure to the naturalised rust *Puccinia eupatorii* (Pucciniaceae). *Biological Invasions*, 14, 1947-  
20 1962.

21  
22  
23 Groppe, K., Steinger, T., Schmid, B., Baur, B., & Boller, T. (2001). Effects of habitat fragmentation on  
24 choke disease (*Epichloe bromicola*) in the grass *Bromus erectus*. *Journal of Ecology*, 89, 247-255.

25  
26  
27 Han, X.M., Dendy, S.P., Garrett, K.A., Fang, L., & Smith, M.D. (2008). Comparison of damage to native  
28 and exotic tallgrass prairie plants by natural enemies. *Plant Ecology*, 198, 197-210.

29  
30  
31 Haubensak, K. & Smyth, A. (2002). *Tragopogon porrifolius*. University of California, Berkley.

32  
33 Hille Ris Lambers, J., Harpole, W.S., Tilman, D., Knops, J., & Reich, P.B. (2004). Mechanisms  
34 responsible for the positive diversity-productivity relationship in Minnesota grasslands. *Ecology*  
35 *Letters*, 7, 661-668.

36  
37  
38 Hoffman, G.D. & Rao, S. (2013). Association of slugs with the fungal pathogen *Epichloë typhina*  
39 (Ascomycetina: Clavicipitaceae): potential role in stroma fertilisation and disease spread. *Annals of*  
40 *Applied Biology*, 162, 324-334.

41  
42  
43 Jenkins, S.N., Waite, I.S., Blackburn, A., Husband, R., Rushton, S.P., Manning, D.C., et al. (2009).  
44 Actinobacterial community dynamics in long term managed grasslands. *Antonie van Leeuwenhoek*  
45 *International Journal of General and Molecular Microbiology*, 95, 319-334.

46  
47  
48 Jorritsma-Wienk, L.D., Ameloot, E., Lenssen, J.P.M., & de Kroon, H. (2007). Differential responses of  
49 germination and seedling establishment in populations of *Tragopogon pratensis* (Asteraceae). *Plant*  
50 *Biology*, 9, 109-115.

51  
52  
53 Kaj, I., Krone, S. M. & Lascoux, M. L. (2001). Coalescent theory for seed bank models. *Journal of*  
54 *Applied Probability*, 38, 285-300.

55  
56  
57 Koubek, T., & Herben, T. (2008). Effect of systemic diseases on clonal integration: modelling  
58 approach. *Evolutionary Ecology*, 22, 449-460.

59  
60  
61  
62  
63  
64  
65

1 Knops, J.M.H., Tilman, D., Haddad, N.M., Naeem, S., Mitchell, C.E., Haarstad, J., et al. (1999). Effects  
2 of plant species richness on invasion dynamics, disease outbreaks, insect abundances and diversity.  
3 *Ecology Letters*, 2, 286-293.

4 Kranz, J. (1990). Fungal disease in multispecies plant communities. *New Phytologist*, 116, 383-405.

5 Kremer, R.J., Caesar, A.J., & Souissi, T. (2006). Soilborne microorganisms of *Euphorbia* are potential  
6 biological control agents of the invasive weed leafy spurge. *Applied Soil Ecology*, 32, 27-37.

7 Latch, G.C.M. (1998). Grass endophytes as a model. *Sydowia*, 50, 213-228.

8 Lau, J.A., Strengbom, J., Stone, L.R., Reich, P.B., & Tiffin, 2008. Direct and indirect effects of CO<sub>2</sub>,  
9 nitrogen, and community diversity on plant-enemy interactions. *Ecology*, 89, 226-236.

10 Linders, E.G.A., van Damme, J.M.M., & Zadoks, J.C. (1996). Epidemics of *Diaporthe adunca* in  
11 experimental and in natural populations of *Plantago lanceolata* and the effect of partial resistance  
12 on disease development. *Plant Pathology* 45, 70-83.

13 Malmstrom, C.M., McCullough, A.J., Johnson, H.A., Newton, L.A., & Borer, E.T. (2005). Invasive  
14 annual grasses indirectly increase virus incidence in California native perennial bunchgrasses.  
15 *Oecologia*, 145, 153-164.

16 Malmstrom, C.M., Shu, R., Linton, E.W., Newton, L.A., & Cook, M.A. (2007). Barley yellow dwarf  
17 viruses (BYDVs) preserved in herbarium specimens illuminate historical disease ecology of invasive  
18 and native grasses. *Journal of Ecology*, 95, 1153-1166.

19 Mattner, S. W. & Parbery, D. G. (2007). Crown rust affects plant performance and interference ability  
20 of Italian ryegrass in the post-epidemic generation. *Grass and Forage Science*, 62, 437-444.

21 Mitchell, C.E. (2003). Trophic control of grassland production and biomass by pathogens. *Ecology*  
22 *Letters*, 6, 147-155.

23 Mitchell, C.E., Tilman, D., & Groth, J.V. (2002). Effects of grassland plant species diversity,  
24 abundance, and composition on foliar fungal disease. *Ecology*, 83, 1713-1726.

25 Mitchell, C.E., Reich, P.B., Tilman, D., & Groth, J.V. (2003). Effects of elevated CO<sub>2</sub>, nitrogen  
26 deposition, and decreased species diversity on foliar fungal plant disease. *Global Change Biology*, 9,  
27 438-451.

28 Moore, S.M., Manore, C.A., Bokil, V.A., Borer, E.T., & Hosseini, P.R. (2011). Spatiotemporal model of  
29 barley and cereal yellow dwarf virus transmission dynamics with seasonality and plant competition.  
30 *Bulletin of Mathematical Biology*, 73, 2707-2730.

31 Nelson, C.J., & Burns, J.C. (2006). Fifty years of grassland science leading to change. *Crop Science*, 46,  
32 2204-2217.

33 Nunney, L. (2002). The effective size of annual plant populations: The interactions of a seed bank  
34 with fluctuation size in maintaining genetic variation. *American Naturalist*, 60, 195-204.

35  
36  
37  
38  
39  
40  
41  
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43  
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56  
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58  
59  
60  
61  
62  
63  
64  
65



1 Paul, N. D., & Ayres, P. G. (1987) Survival, growth and reproduction of groundsel (*Senecio vulgaris*)  
2 infected by rust (*Puccinia lagenophorae*) in the field during summer. *Journal of Ecology*, 75, 61-71.

3 Pautasso, M., Dehnen-Schmutz, K., Holdenrieder, O., Pietravalle, S., Salama, N., Jeger, M.J., et al.  
4 (2010). Plant health and global change – some implications for landscape management. *Biological*  
5 *Reviews*, 85, 729-755.

6  
7  
8 Pekrun, C., Lane, P. W., & Lutman, P. J. W. (2005). Modelling seedbank dynamics of volunteer oilseed  
9 rape (*Brassica napus*). *Agricultural Systems*, 84, 1-20.

10  
11 Petermann, J.S. Fergus, A.J.F., Turnbull, L.A., & Schmid B. (2008). Janzen-Connell effects are  
12 widespread and strong enough to maintain diversity in grasslands. *Ecology*, 89, 2399-2406.

13  
14 Peters, J.C., & Shaw, M.W. (1996). Effect of artificial exclusion and augmentation of fungal plant  
15 pathogens on a regenerating grassland. *New Phytologist*, 134, 295-307.

16  
17 Pfender, W.F. (2001). Host range differences between populations of *Puccinia graminis* subsp.  
18 *graminicola* obtained from perennial ryegrass and tall fescue. *Plant Disease*, 85, 993-998.

19  
20 Pfender, W.F. & Alderman, S.C. (2006). Regional development of orchardgrass choke and estimation  
21 of seed yield loss. *Plant Disease*, 90, 240-244.

22  
23 Pico, F. X., Ouborg, N. J., & van Groenendael, J. M. (2003). Fitness traits and dispersal ability in the  
24 herb *Tragopogon pratensis* (Asteraceae): Decoupling the role of inbreeding depression and maternal  
25 effects. *Plant Biology*, 5, 522-530.

26  
27 Power, A.G., Borer, E.T., Hosseini, P., Mitchell, C.E., & Seebloom, E.W. (2011). The community  
28 ecology of barley/cereal yellow dwarf viruses in Western US grasslands. *Virus Research*, 159, 95-100.

29  
30 Qi, M. Q., Upadhyaya, M. K., & Turkington, R. (1996). Reproductive behaviour of natural populations  
31 of meadow salsify (*Tragopogon pratensis*). *Weed Science*, 44, 68-73.

32  
33 Retallack, G.J. (2013). Global cooling by grassland soils of the geological past and near future. *Annual*  
34 *Review of Earth and Planetary Sciences*, 41, 69-86.

35  
36 Roberts, H. A. (1986). Seed persistence in soil and seasonal emergence in plant species from  
37 different habitats. *Journal of Applied Ecology*, 23, 639-656.

38  
39 Roscher, C., Schumacher, J., Foitzik, O., & Schultze, E.D. (2007). Resistance to rust fungi in *Lolium*  
40 *perenne* depends on within-species variation and performance of the host species in grasslands of  
41 different plant density. *Oecologia*, 153, 173-183.

42  
43 Rousk, J., Brookes, P.C., & Bååth E. (2011). Fungal and bacterial growth responses to N fertilization  
44 and pH in the 150-year 'Park Grass' UK grassland experiment.

45  
46 Rudgers, J.A. & Orr, S. (2009). Non-native grass alters growth of native tree species via leaf and soil  
47 microbes. *Journal of Ecology*, 97, 247-255.

48  
49 Salama, N.K.G., Edwards, G.R., Heard, M.S., & Jeger, M.J. (2011). The suppression of reproduction of  
50 *Tragopogon pratensis* infected by the rust fungus *Puccinia hysterium*. *Fungal Ecology*, 3, 406-408.

51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1 Salama, N.K.G. (2009). Regulation of a Biennial Host Plant Population by an Autoecious, Demicyclic  
2 Rust Fungus: *Puccinia hysterium* on *Tragopogon pratensis* in the Park Grass Experiment. Unpublished  
3 PhD thesis. Imperial College, University of London.

4 Salama, N.K.G., van den Bosch, F., Edwards, G.R., Heard, M.S., & Jeger, M.J. (2012). Population  
5 dynamics of a non-cultivated biennial plant *Tragopogon pratensis* infected by the autecious  
6 demicyclic rust fungus *Puccinia hysterium*. *Fungal Ecology*, 5, 530-542.

7  
8  
9 Salama, N.K.G., Jeger, M.J. (in press). Pathogen induced mortality and its role in natural plant-rust  
10 systems. In Exercises in Plant Disease Epidemiology, 2nd Edition (eds. Stevenson, K, Jeger, MJ) APS  
11 Press, Minneapolis.

12  
13  
14 Schoolmaster, D.R. (2008). Recruitment limitation modifies the net effects of shared enemies on  
15 competitively inferior plants. *Journal of Ecology* 96, 114-121.

16  
17  
18 Shiba, T., & Sugawara, K. (2005). Resistance to the rice leaf bug, *Trigonotylus caelestialium*, is  
19 conferred by *Neotyphodium endophyte* infection of perennial ryegrass, *Lolium perenne*. *Entomologia*  
20 *Experimentalis et Applicata*, 115, 387-392.

21  
22  
23 Schnitzer, S.A., Klironomos, J.N., HilleRisLambers, J., Kinkel, L.L., Reich, P.B., Xiao, K., et al. (2011). Soil  
24 microbes drive the classic plant diversity-productivity pattern. *Ecology*, 92, 296-303.

25  
26  
27 Seabloom, E.W., Borer, E.T., Jolles, A., & Mitchell, C.E. (2009). Direct and indirect effects of viral  
28 pathogens and the environment on invasive grass fecundity in Pacific Coast grasslands. *Journal of*  
29 *Ecology*, 97, 1264-1273.

30  
31  
32 Seabloom, E.W., Borer, E.T., Mitchell, C.E., & Power, A.G. (2010). Viral diversity and prevalence  
33 gradients in North American Pacific coast grasslands. *Ecology*, 91, 721-732.

34  
35  
36 Seabloom, E.W., Borer, E.T., Lacroix, C., Mitchell, C.E., & Power, A.G. (2013). Richness and  
37 composition of niche assembled viral pathogen communities. *PLoS One*, 8, e55675.

38  
39  
40 Silvertown, J., Poulton, P., Johnston, E., Edwards, G., Heard, M., & Biss, P. M. (2006). The Park Grass  
41 Experiment 1856-2006: Its contribution to ecology. *Journal of Ecology*, 94, 801-814.

42  
43  
44 Sjoberg, J., Martensson, A., & Persson, P. (2007). Are field populations of arbuscular mycorrhizal  
45 fungi able to suppress the transmission of seed-borne *Bipolaris sorokiniana* to aerial plant parts.  
46 *European Journal of Plant Pathology*, 117, 45-55.

47  
48  
49 Strengbom, J., Nordin, A., Nasholm, T., Ericson, L. (2002). Parasitic fungus mediates change in  
50 nitrogen-exposed boreal forest vegetation. *Journal of Ecology*, 90, 61-67.

51  
52  
53 Taye, T., Einhorn, G., & Metz, R. (2004). *Parthenium hysterophorus*, an invasive species in Ethiopia –  
54 investigations on the occurrence and on its pathogens. *Zeitschrift fur Pflanzenkrankheiten und*  
55 *Pflanzenschutz*, 19, 271-278.

56  
57  
58 van den Berg, F., & van den Bosch, F. (2004). A model for the evolution of pathogen-induced leaf  
59 shedding. *Oikos*, 107, 36-49.

1 van den Berg, F., Robert, C., Shaw, M.W., & van den Bosch, F. (2007). Apical leaf necrosis and leaf  
2 nitrogen dynamics in diseased leaves: a model study. *Plant Pathology*, 56, 424-436.

3 van den Berg, F., van den Bosch, F., Powers, S.J., & Shaw, M.W. (2008). Apical leaf necrosis as a  
4 defence mechanism against pathogen attack: effects of high nutrient availability on onset and rate  
5 of necrosis. *Plant Pathology*, 57, 1009-1016.

6  
7  
8 van den Berg, F., & van den Bosch, F. (2009). The presence of generalist plant pathogens might not  
9 explain the long-term coexistence of plant species. *Journal of Theoretical Biology*, 257, 446-453.

10  
11 van Elsas, J.D., Garbeva, P., & Salles, J. (2002). Effects of agronomical measures on the microbial  
12 diversity of soils as related to the suppression of soil-borne plant pathogens. *Biodegradation*, 13, 29-  
13 40.

14  
15  
16 Vignale M.V. et al (2013). Epichloid endophytes confer resistance to the smut *Ustilago bullata* in the  
17 wild grass *Bromus auleticus* (Trin.). Biological control (in press) DOI 10.1016/j.biocontrol.2013.06.002

18  
19  
20  
21 Vozenilkova, B., Markova, J., Klimes, F., Kvet J, & Maskova, Z. (2008). The influence of mountain  
22 meadow management on the occurrence of *Puccinia perplexans* Plow. *Journal of Plant Diseases and*  
23 *Protection*, 115, 167-171.

24  
25  
26 Vosenilkova, B., Kobes, M., Klimes, F., Cermak, B., & Ptacnikova, V. (2007). The aim of small-site  
27 experiments carried out in the area of the foothills of the Sumava Mts. was to determine the  
28 etiological agent of the disease of meadow peavine (*Lathyrus pratensis* L.). *Gesunde Pflanzen*, 59,  
29 179-182.

30  
31  
32 Waples, R. S. (2006). Seed banks, Salmon and sleeping genes: effective population size in  
33 semelparous, age structures species with fluctuating abundance. *American Naturalist*, 167, 118-135.

34  
35  
36 Wenström, A. (1999). The effect of systemic rusts and smuts on clonal plants in natural systems.  
37 *Plant Ecology*, 141, 93-97.

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Fig.1 The Park Grass Experiment at Rothamsted Research, Harpenden, UK.

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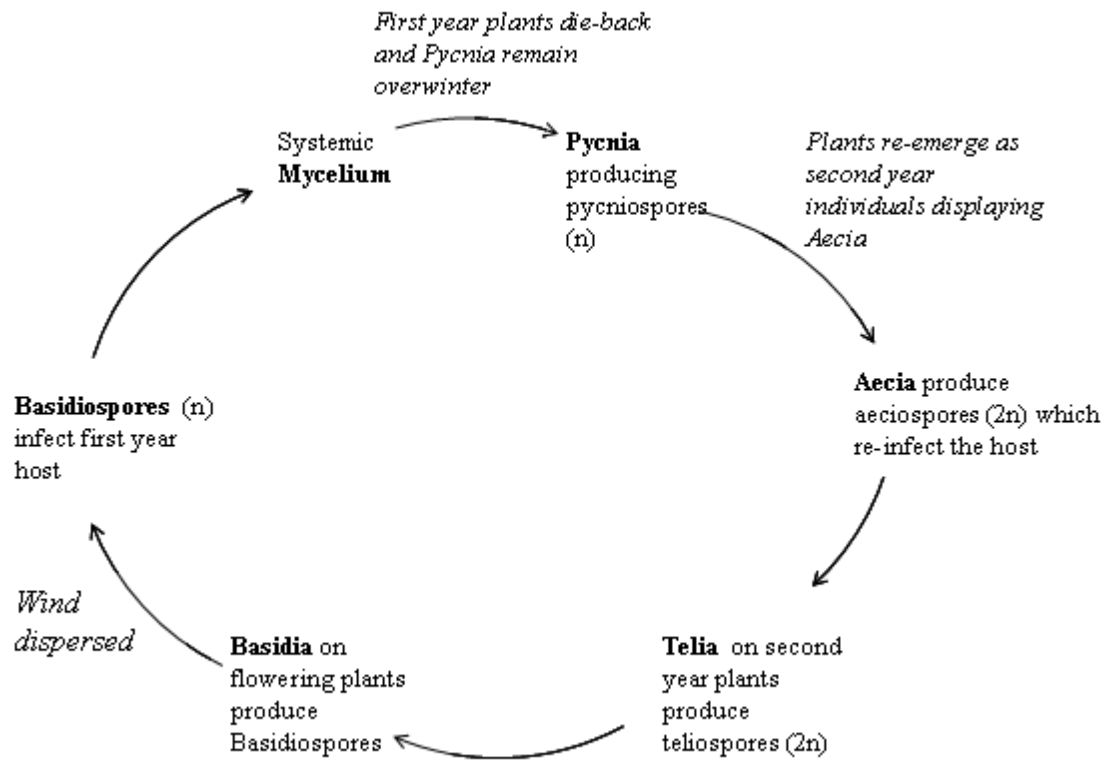


Fig. 2. The life-cycle of *P. hysterium* (n=haploid, 2n=diploid).

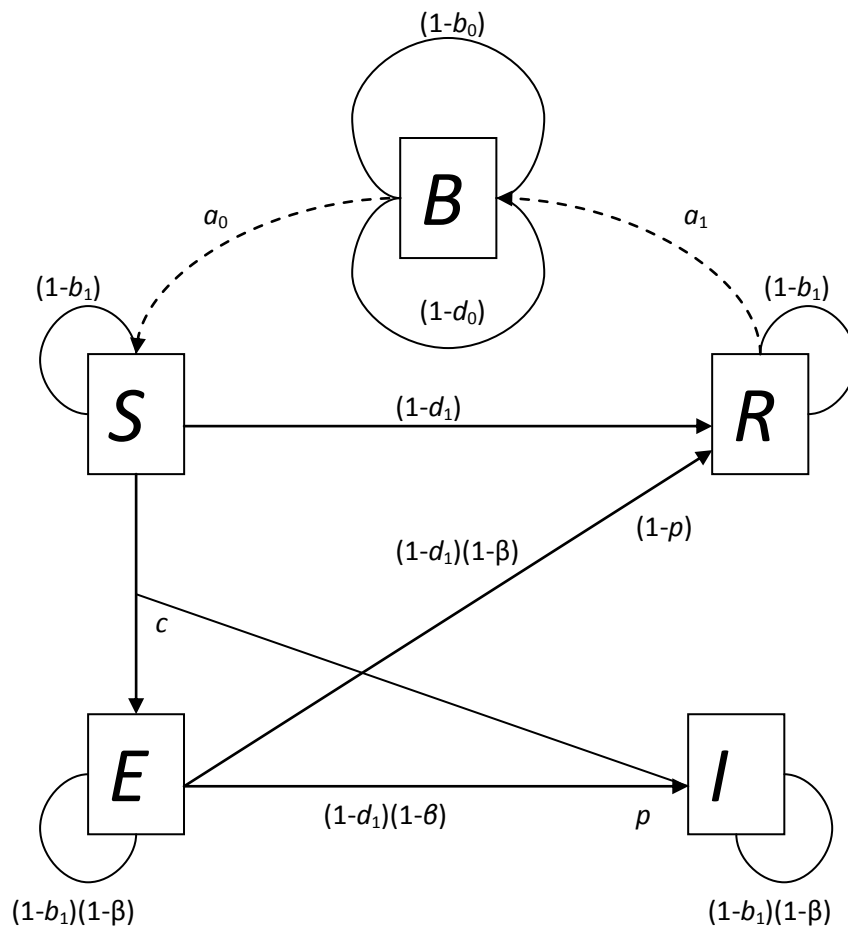


Fig. 3. A schematic representation of the *Tragopogon pratensis*-*Puccinia hysterium* model structure with a seedbank phase ( $B$ ) in addition to the susceptible ( $S$ ), latently infected ( $E$ ), infectious ( $I$ ) and the removed ( $R$ ) components of the above-ground host population.

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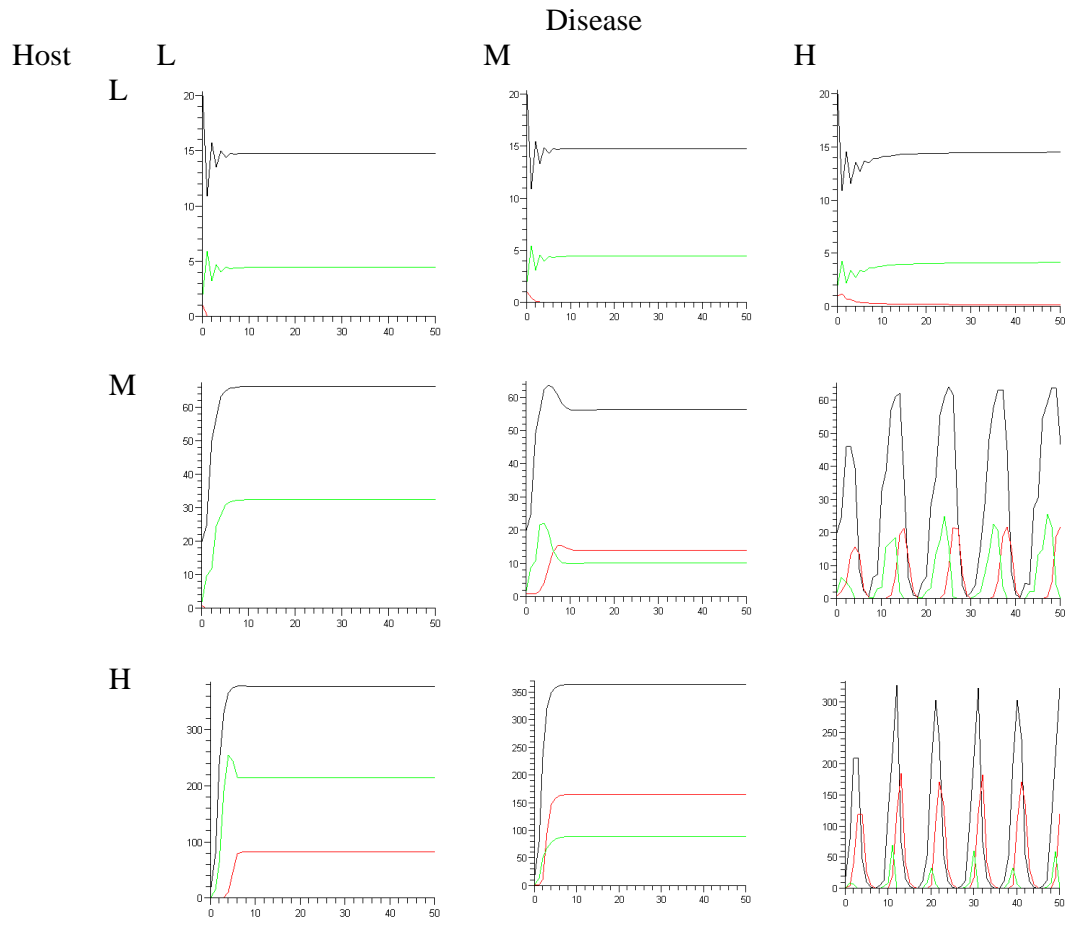


Fig. 4. Simulations produced under conditions of high, medium and low host performance and pathogenicity in the *Tragopogon pratensis*-*Puccinia hysteriorum* system. Each simulation represents — S, — I, — R dynamics.

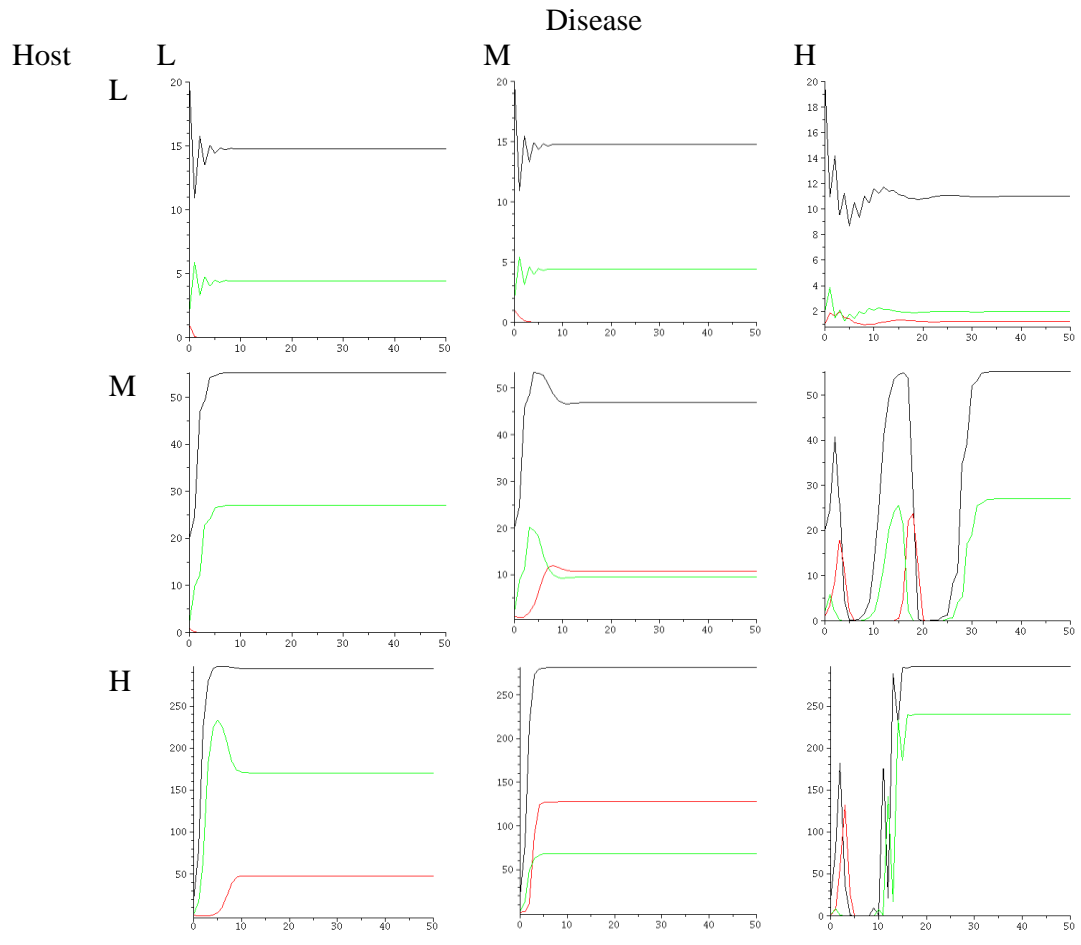


Fig. 5. Simulations representing, high, medium and low host performance and pathogenicity in the *Tragopogon pratensis-Puccinia hystrium* system where there is a trade-off between transmissibility and disease-induced mortality where mortality in the seedbank is high ( $b_0=0.97$ ). Each simulation represents — S, — I, — R dynamics.



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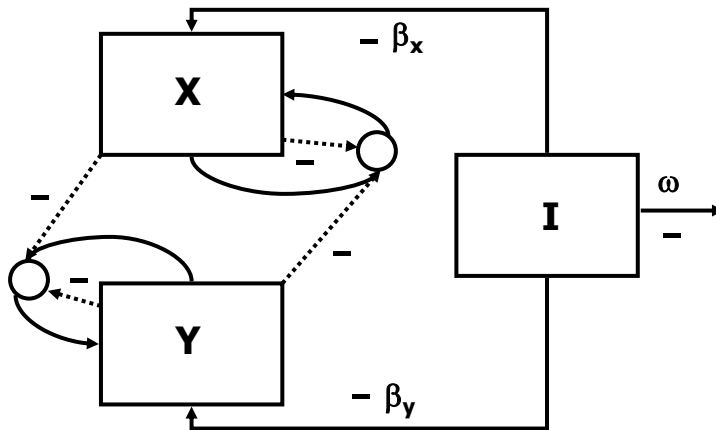


Fig. 6. Schematic representation of competition between two host species ( $X$  and  $Y$ ) in the presence of a generalist pathogen ( $I$ ). Infection parameters are  $\beta_x$  and  $\beta_y$  on species  $X$  and  $Y$  respectively. See text and van den Berg & van den Bosch (2009; Fig.1a) for incorporation of plant intrinsic rates of increase and competition coefficients into the model.

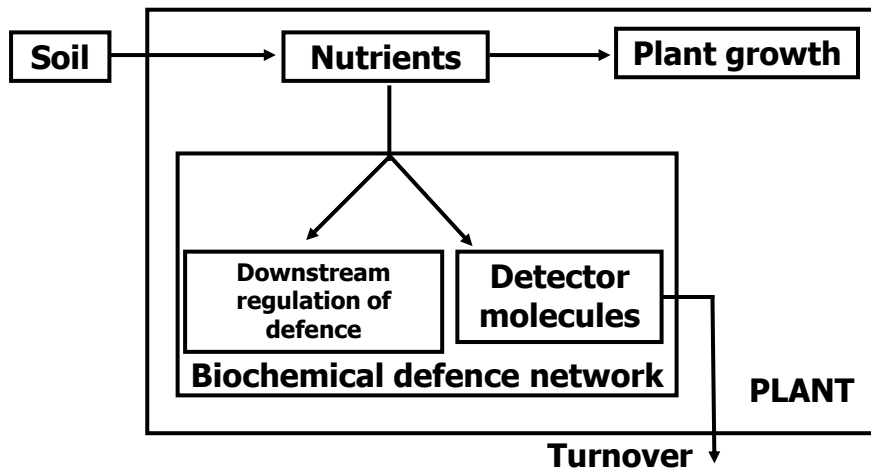


Fig.7. Mechanistic interpretation of the trade-off relationship between plant defence against pathogens and plant growth (van den Berg & van den Bosch, 2009; Fig. 1b) in relation to nutrient supply.

Table 1: Selected experimental studies<sup>1</sup> investigating a range of topics<sup>2</sup> concerned with the impact of disease in managed and natural/semi-natural grassland

Topic studied	Plant/pathogen system	References
Disease and annual management strategies (mowing/mulching/fallow/grazing)	Permanent grass stands with <i>Alopecurus pratensis</i> / <i>Puccinia perplexans</i> and <i>Lathyrus pratensis</i> /powdery mildew	Vozenilkova et al., 2007; 2008
Interactions with other habitats	<i>Carex blanda</i> / <i>Anthracoidiae blanda</i> and <i>Puccinia carici</i> in woodland and grassland; <i>Deschampsia flexuosa</i> with <i>Vaccinium myrtillus</i> / <i>Valdensia heterodoxa</i> in forest vegetation; non-sown grassland species/ <i>Blumeria graminis</i> and <i>Zymoseptoria tritici</i> in cereal fields; <i>Lolium arundinaceum</i> / <i>Neotyphodium coenophialum</i> and grassland to woodland succession	Finch & Alexander, 2011; Strengbom et al., 2002; Firbank et al., 2003; Rutgers & Orr, 2009
Manipulative experimentation in relatively unmanaged grassland ecosystems	Native and introduced grasses /barley (cereal) yellow dwarf viruses in California and N American Pacific coast; New Zealand native grasses and exotic cereal and pasture viruses	Malstrom et al., 2005, 2007; Borer et al., 2007, 2009, 2010; Seabloom et al., 2009, 2010, 2013; Power et al., 2011; Delmiglio et al., 2010
Invasion of pathogens into native grasslands	Invasive and non-invasive exotic grass species/leaf rust and herbivory; <i>Commandra umbellata</i> and <i>Andropogon gerardii</i> / <i>Puccinia andropogonis</i> in native prairie; <i>Plantago lanceolata</i> / <i>Diaporthe adunca</i> and partial resistance	Han et al., 2008; Barnes et al., 2005; Linders et al., 1996
Biological control of introduced plant species considered as weeds in managed grassland	<i>Campuloclinium macrocephalum</i> / <i>Puccinia eupatorii</i> ; <i>Euphorbia esula</i> /soil-borne pathogens and insect root feeding; <i>Parthenium hysterophorus</i> /facultative and	Goodall et al., 2012; Kremer et al., 2006; Taye et al., 2004; de Jong et al, 2002; Franzen et al., 1994

	obligate fungal pathogens and a phytoplasma; <i>Circium arvense/Sclerotinia sclerotiorum</i> and <i>Puccinia punctiformis</i>	
Soil microbes and plant diversity/disease suppression	Herbaceous plant communities/mycorrhizal and non-mycorrhizal infection; long-term managed grasslands/actinobacterial communities; barley and grassland/arbuscular mycorrhizal fungi and transmission of <i>Bipolaris sorokiniana</i> ; permanent grassland and arable rotations/ <i>Rhizoctonia solani</i>	Schnitzer al., 2011; Jenkins et al., 2009; Sjoberg et al., 2007; van Elsas et al., 2002
Fungal endophytes	Rice pests/ <i>Neotyphodium</i> in grasslands; <i>Bromus auleticus/Ustilago bullata</i>	Shiba & Sugawara, 2005, Vignale et al., 2013

<sup>1</sup> the examples of experimental studies for each topic are representative, not exhaustive.

<sup>2</sup> the range of topics is illustrative for grassland ecosystems, not exhaustive

Table 2. Parameter ranges for low, medium and high host performance and pathogenicity used in simulations of *Tragopogon pratensis*-*Puccinia hysterium* dynamics.

Disease	Low	Medium	High
$c$	0.1	0.3	0.8
$p$	0.3	0.7	1.0
$\beta$	0.1	0.2	0.3
<i>Host</i>			
$a_0$	0.2	0.3	0.5
$a_1$	100	100	120
$b_0$	0.97	0.7	0.5
$b_1$	0.4	0.3	0.1
$d_0$	0.3	0.2	0.1
$d_1$	0.5	0.3	0.1
$\lambda$	1.0	0.5	0.2

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