

# Increased soil phosphorus availability induced by faba bean root exudation stimulates root growth and phosphorus uptake in neighbouring maize

Deshan Zhang<sup>1</sup>, Chaochun Zhang<sup>1</sup>, Xiaoyan Tang<sup>1</sup>, Haigang Li<sup>1</sup>, Fusuo Zhang<sup>1</sup>, Zed Rengel<sup>2</sup>, William R. Whalley<sup>3</sup>, William J. Davies<sup>4</sup> and Jianbo Shen<sup>1</sup>

<sup>1</sup>Key Laboratory of Plant-Soil Interactions, Department of Plant Nutrition, Ministry of Education, China Agricultural University, Beijing 100193, China; <sup>2</sup>Soil Science & Plant Nutrition, School of Earth and Environment, The UWA Institute of Agriculture, The University of Western Australia, Crawley, WA 6009, Australia; <sup>3</sup>Rothamsted Research, West Common, Harpenden, Hertfordshire, AL5 2JQ, UK; <sup>4</sup>Lancaster Environment Centre, University of Lancaster, Lancaster, LA1 4YQ, UK

## Summary

Author for correspondence:

Jianbo Shen

Tel: +86 10 62732406

Email: jbshen@cau.edu.cn

Received: 6 February 2015

Accepted: 22 July 2015

*New Phytologist* (2016) **209**: 823–831

doi: 10.1111/nph.13613

**Key words:** heterogeneous phosphorus supply, phosphorus (P) uptake, rhizosphere processes, root interactions, *Vicia faba* (faba bean), *Zea mays* (maize).

- Root growth is influenced by soil nutrients and neighbouring plants, but how these two drivers affect root interactions and regulate plant growth dynamics is poorly understood. Here, interactions between the roots of maize (*Zea mays*) and faba bean (*Vicia faba*) are characterized.
- Maize was grown alone (maize) or with maize (maize/maize) or faba bean (maize/faba bean) as competitors under five levels of phosphorus (P) supply, and with homogeneous or heterogeneous P distribution.
- Maize had longer root length and greater shoot biomass and P content when grown with faba bean than with maize. At each P supply rate, faba bean had a smaller root system than maize but greater exudation of citrate and acid phosphatase, suggesting a greater capacity to mobilize P in the rhizosphere. Heterogeneous P availability enhanced the root-length density of maize but not faba bean. Maize root proliferation in the P-rich patches was associated with increased shoot P uptake.
- Increased P availability by localized P application or by the presence of faba bean exudation stimulated root morphological plasticity and increased shoot growth in maize in the maize/faba bean mixture, suggesting that root interactions of neighbouring plants can be modified by increased P availability.

## Introduction

Root interactions among plants play an important role in determining the performance of individuals in natural communities as well as crop productivity in agroecological systems. The foraging capacity of roots is a key trait in belowground competition and is dependent on morphological (root architecture) and physiological plasticity (e.g. exudation of organic and inorganic compounds, nutrient uptake) in response to the soil environment. To compete for soil resources (water and nutrients), plants invest in root growth to maximize root length density and can outcompete neighbouring plants (Cahill *et al.*, 2010; Cahill & McNickle, 2011). For example, *Rumex palustris* increased root growth in nutrient-rich areas and as a result became the superior species in competition with *Agrostis stolonifera* (Mommer *et al.*, 2012). In addition to root growth, root physiological plasticity (i.e. activity) can be critical in obtaining a long-term competitive advantage (Fransen *et al.*, 2001). Modelling studies have suggested that physiological responses may be more important for the capture

of patchy nitrogen (N) than morphological responses (Jackson & Caldwell, 1996; Robinson, 1996; Dunbabin *et al.*, 2003).

Root morphological and physiological plasticity is determined by various environmental factors, such as availability of soil resources and their distribution (Jackson *et al.*, 1990; Hutchings & de Kroon, 1994; Hodge, 2004; Cahill *et al.*, 2010; Andersen *et al.*, 2014). Soil nutrient availability can profoundly affect root morphology and physiology. For example, cluster-root formation and citrate exudation in *Lupinus albus* can be induced by low phosphorus (P) conditions, and inhibited by increased P supply (Shen *et al.*, 2005; Lambers *et al.*, 2006; Li *et al.*, 2008). Faba bean roots have no significant growth response to localized nutrient supply (Li *et al.*, 2014a), suggesting relatively poor morphological plasticity to variable nutrient availability, and potentially a low capacity to take full advantage of nutrient-rich patches. However, in many species, root growth and distribution are affected by soil nutrient heterogeneity (Drew, 1975; Li *et al.*, 2008; Jing *et al.*, 2010). In response to nutrient-rich patches, plants tend to stimulate root growth and alter root distribution, with increased

root proliferation in the local nutrient-rich zone (Drew, 1975; Hodge *et al.*, 1999; Hodge, 2004). In addition, some species show root physiological responses to localized nutrient enrichment (e.g. an increase in the nutrient uptake capacity per unit root length; Jackson *et al.*, 1990). Hence, foraging for nutrients is determined by root morphological and physiological dynamics in response to soil environmental conditions, and may result in differences in competitive ability, which, to a large extent, affects plant productivity and nutrient uptake in a mixed system.

Growing mixtures of maize and faba bean are used widely to improve grain yield and P-use efficiency in cropping ecosystems in many parts of the world (intercropping; Li *et al.*, 2007; Zhang *et al.*, 2010; Shen *et al.*, 2011, 2013). Root interactions have a profound impact on P uptake and yield in the maize/faba bean system (Li *et al.*, 2003, 2014b), with rhizosphere acidification caused by faba bean increasing mobilization of organic/inorganic P sources and facilitating P uptake by target maize (Li *et al.*, 2007). The roots of maize and faba bean intermingle and appear to grow together; hence, complementarity of the spatial root distribution of intercropped species contributes to interspecies facilitation in maize/faba bean intercropping compared to the maize/maize monocropping system (Li *et al.*, 2006). However, the detailed mechanisms underlying root interactions in the maize/faba bean system are still unclear.

Much work on root traits in response to P supply has been conducted in maize or faba bean separately (Li *et al.*, 2014a), but how altered nutrient gradients and spatial distribution affect root–root interactions in the mixture, and thus regulate plant growth dynamics, is largely unknown. Maize and faba bean roots grow well together, but the root interactions based on complementary and niche differentiation in space might be enhanced with a better understanding of how spatial changes in soil nutrient availability affect root growth, distribution and hence competition.

In order to investigate the effects of heterogeneous nutrient availability on root growth, distribution and thus competition intensity in the maize/faba bean mixture system, we conducted experiments with varying P supplies and homogeneous or heterogeneous P distribution to test the hypothesis that neighbouring faba bean would not compete strongly with target maize because faba bean roots have low morphological plasticity that is compensated for by significant physiological plasticity in variable soil P supply (Hypothesis 1). We then demonstrate that increased maize growth in the maize/faba bean mixtures is consistent with the hypothesis that P availability induced by faba bean exudation stimulates root morphological plasticity in maize, resulting in improved maize shoot growth and nutrient uptake (Hypothesis 2).

## Materials and Methods

### Experimental set-up

**Experiment 1** In order to investigate how maize roots respond to faba bean in soil with variable P supply, and test how soil P availability regulates root interactions between maize and faba bean, a pot experiment was conducted in a glasshouse with five soil P supply levels and three cropping treatments. Maize was grown alone

as a single species (single maize treatment), mixed with other maize plants (maize/maize treatment) or intercropped with faba bean (maize/faba bean treatment) in soil supplied with 0, 50, 100, 200 or 500 mg P kg<sup>-1</sup> soil, corresponding to soil Olsen-P values of 3.5, 36, 51, 123 and 270 mg kg<sup>-1</sup>, respectively. We choose the soil P supply levels based on a preliminary experiment with the same soil: (1) deficient, 0 and 50 mg P kg<sup>-1</sup> soil; (2) moderate, 100 mg P kg<sup>-1</sup> soil; (3) adequate, 200 mg P kg<sup>-1</sup> soil, and (4) high, 500 mg P kg<sup>-1</sup> soil (to mimic a high-input maize cropping system in intensive agriculture of China). There were 15 treatment combinations in the study, with four replicates per treatment.

The soil was collected from Shangzhuang experimental station in Beijing, China, air-dried and passed through a 2-mm sieve. Soil properties were as follows: Olsen-P 3.5 mg kg<sup>-1</sup>, organic C 11.5 g kg<sup>-1</sup>, total N 0.72 g kg<sup>-1</sup>, available N 8.5 mg kg<sup>-1</sup> (NO<sub>3</sub><sup>-</sup> and NH<sub>4</sub><sup>+</sup>), available K 32.3 mg kg<sup>-1</sup> and pH 8.2 (the ratio of soil to CaCl<sub>2</sub> solution was 1 : 2.5). The pot was filled with 1.5 kg of air-dried soil. To ensure that the nutrient supply was adequate for plant growth, soil was also fertilized with basal nutrients at the following rates (mg per pot): Ca(NO<sub>3</sub>)<sub>2</sub>·4H<sub>2</sub>O, 1687; K<sub>2</sub>SO<sub>4</sub>, 200; MgSO<sub>4</sub>·7H<sub>2</sub>O, 65; Fe-EDTA, 8.78; MnSO<sub>4</sub>·H<sub>2</sub>O, 10; ZnSO<sub>4</sub>·7H<sub>2</sub>O, 15; CuSO<sub>4</sub>·5H<sub>2</sub>O, 3; H<sub>3</sub>BO<sub>3</sub>, 1; and Na<sub>2</sub>MoO<sub>4</sub>·5H<sub>2</sub>O, 0.25. Phosphorus was applied as Ca (H<sub>2</sub>PO<sub>4</sub>)<sub>2</sub>·H<sub>2</sub>O.

The genotype of maize (*Zea mays* L.) was cv ZD958, and the genotype of faba bean (*Vicia faba* L.) was cv Linca5. Maize and faba bean seeds were surface-sterilized in 30% v/v H<sub>2</sub>O<sub>2</sub> for 20 min, washed with deionized water, soaked in CaSO<sub>4</sub> saturated solution for 12 h and then germinated in Petri dishes covered with wet filter papers for 1–2 d at 22°C. All of the pots were arranged in a completely randomized design, and were re-randomized weekly during the experiment. The plants were watered every day to maintain field capacity (18%, w/w).

Plants were harvested 38 d after sowing (DAS) and separated into shoots and roots. Following root excavation, the soil adhering to roots was defined as rhizosphere soil and was sub-sampled for carboxylate and acid phosphatase measurements. Roots were transferred to a tube containing 50 ml of 0.2 μM CaCl<sub>2</sub> and gently shaken to dislodge the rhizosphere soil, followed by shaking for 5–10 s to create homogeneous suspensions. A suspension volume of 10 ml was taken by pipette to a 10-ml centrifuge tube for carboxylate analysis by HPLC (see below, carboxylate analysis method), and a 0.5-μl aliquot of suspension was placed in a 2-ml centrifuge tube for acid phosphatase measurement (see below, acid phosphatase analysis method).

**Experiment 2** In order to address the question of how soil P heterogeneity affects root interactions between maize and faba bean, we set up a rhizo-box experiment comprising two P supply treatments and three cropping treatments (maize, maize/maize and maize/faba bean) with four replicates of each treatment. We chose the same soil, genotypes of maize and faba bean, and crop treatments as in Expt 1.

In order to record root growth and distribution, we constructed microcosm PVC-rhizo-boxes (20 × 1.5 × 35 cm, Fig. 1), containing irrigation holes, a viscose fleece for moisture

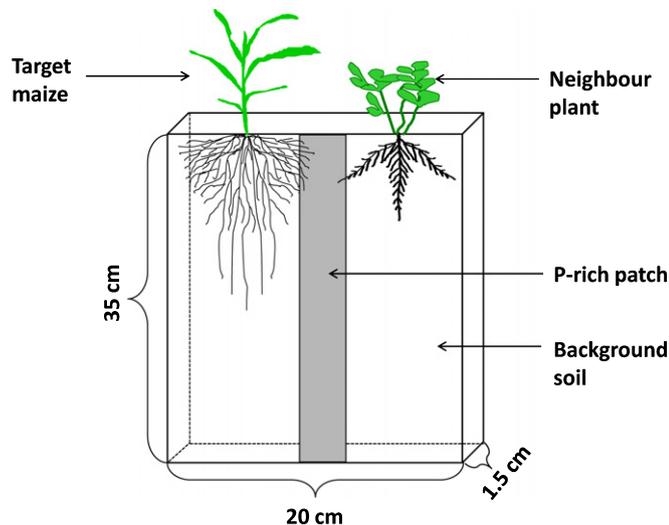


Fig. 1 A schematic diagram of the rhizo-box.

distribution, transparent plastic foil for soil-covering and a Perspex front lid with screws. All rhizo-boxes were filled with 1.3 kg air-dried soil. Phosphorus was supplied as  $\text{Ca}(\text{H}_2\text{PO}_4)_2 \cdot \text{H}_2\text{O}$  in an homogeneous or heterogeneous pattern. For the heterogeneous P treatment, a 3.5-cm P-rich layer (227.5 g soil) containing 46.2 mg P (200 mg P  $\text{kg}^{-1}$  soil) was manually mixed and placed at the centre of the rhizo-box (Fig. 1), referred to as the P-rich patch, and the remaining soil without P additions was the background soil (1072.5 g soil). For the homogeneous P treatment, the same total P (46.2 mg P) was spread evenly throughout the soil (35 mg P  $\text{kg}^{-1}$  soil).

Maize and faba bean seeds were handled as in Expt 1. To ensure nutrient supply for plant growth, the same amounts of basal nutrients as in Expt 1 were added to soil, followed by thorough mixing.

Before planting, all rhizo-boxes were irrigated through the bottom irrigation holes. After 15 d of growth, the rhizo-boxes were irrigated every 2 d until the final harvest. After 30 d of growth, the target species (maize) and neighbouring species were separated, and shoots of each species were collected for biomass measurements. Shoot P content was analysed in maize. In the heterogeneous treatments, roots grown in vs out of the P-rich patch were sampled separately for length measurement, and the corresponding roots at the same location were sampled in the homogeneous treatments.

Both experiments were conducted in a glasshouse at China Agricultural University, Beijing (latitude: 40°01'N, longitude: 116°16'E). In Expt 1, the temperature in the glasshouse was maintained at 21–25°C during the day and 15–18°C at night, with 12–14 h daytime throughout the growth period. In Expt 2, temperature was maintained at 24–28°C during the day and 18–20°C at night, with 14–19 h daytime.

## Measurements

**Plant biomass and phosphorus uptake** Shoots were oven-dried at 105°C for 30 min and then at 65°C for 3 d before weighing for dry biomass determination. Phosphorus concentration in

shoots was determined after digestion with a mixture of 5 ml of concentrated sulphuric acid and 8 ml of 30% v/v  $\text{H}_2\text{O}_2$ . Shoot P was analysed by the molybdovanadophosphate method at 440 nm by spectrophotometry (Varian Vista-Pro CCD; Johnson & Ulrich, 1959).

**Root parameter measurement** Roots were washed in deionized water and then scanned with an EPSON root scanner at 400 dots-per-inch resolution (Epson Expression 1600 pro, Model EU-35, Tokyo, Japan). The total root length was analysed with software Win-RHIZO (Regent Instruments Inc., Quebec, QC, Canada).

**Carboxylate and acid phosphatase exudation** Carboxylates in the rhizosphere soil were analysed using a reversed phase high-performance liquid chromatography (HPLC) system according to a previous report (modified from Shen *et al.*, 2003, and Wang *et al.*, 2010). The chromatographic separation was conducted on a 250 × 4.6 mm reversed-phase column (Alltima C18, 5 Micrometers; Alltech Associates Inc., Deerfield, IL, USA). The mobile phase was 25 mM  $\text{KH}_2\text{PO}_4$  (pH 2.25) with a flow rate of 1 ml  $\text{min}^{-1}$  at 31°C. Detection of carboxylates was carried out at 214 nm.

In order to determine the activity of acid phosphatase in the rhizosphere soil, 0.5-ml aliquots of soil suspensions were transferred into a 2-ml Eppendorf vial with 0.4 ml sodium acetate buffer and 0.1 ml *p*-nitrophenyl phosphate (NPP) substrate added. Vials were incubated at 30°C for 60 min, and the reaction was terminated by adding 0.5 ml of 0.5 M NaOH. Absorption was measured at 405 nm (Alvey *et al.*, 2000).

## Statistical analyses

In order to investigate the effect of faba bean on target maize, the relative interaction intensity (RII) was calculated in Expt 1. RII can be used to compare the performance of plants growing in mixtures or alone in order to estimate the magnitude of competitive responses of the target plant species (Markham & Chanway, 1996; Howard & Goldberg, 2001; Weigelt & Jolliffe, 2003). Calculation of RII was based on the method proposed by Wilson & Keddy (1986) and modified by Markham & Chanway (1996). A formula of Markham & Chanway (1996) was adopted to calculate RII (modified from Valladares *et al.*, 2006).

$$\text{RII} = \frac{\sum \left( \frac{xi - xi'}{X} \right)}{n}$$

( $xi$ , shoot biomass of target maize in the presence of neighbours;  $xi'$ , biomass of target maize in the absence of neighbours).  $X$  is either  $xi$  or  $xi'$ , whichever was larger. RII is symmetrical around zero and constrained between +1 and -1 (Markham & Chanway, 1996), so that magnitudes of competition or facilitation can be compared. If maize is unaffected by its neighbour, RII will be zero. Negative RII values represent the relative competition between neighbouring plants; positive values indicate that target maize can benefit from neighbour presence.

The  $i$  and  $i'$  are two randomly selected individuals (in the present study,  $i = 1, \dots, 4$ , because each treatment had four replicates) of the same species belonging to two different cropping treatments. The  $n$  is the number of  $(x_i - x_{i'})$  values. In the present study,  $n$  is equal to 16 because four replicates (four random individuals) were set in each pair of cropping treatments (single maize and maize/maize, or single maize and maize/faba bean).

Analysis of variance (ANOVA) was conducted using SPSS statistical software (SPSS version 19.0, IBM SPSS Inc., Chicago, IL, USA). Significant differences among means were separated by LSD at the  $P \leq 0.05$  probability level. Plant growth and root length were subjected to two-way ANOVA to assess the effects of plant species, soil P supply, and their interaction in Expt 1 as well as the effects of heterogeneous/homogeneous P supply and competition in Expt 2.

## Results

Root interactions between maize and faba bean grown with variable soil P supply (Expt 1)

Shoot biomass and P content in target maize increased with soil P supply (Fig. 2). Compared with the single maize treatment at five levels of P supply, the shoot biomass of target maize in the mixtures was unaffected by the presence of faba bean, but was significantly reduced when maize was grown with maize (Fig. 2a). The shoot P content of target maize showed a similar response to shoot biomass when grown either as single or with neighbours (Fig. 2b). In the maize/maize mixture, a decrease in maize shoot P content was affected by soil P availability and the presence of neighbouring maize, although there was no significant interaction (Fig. 2b; Supporting Information Table S1). In the maize/faba bean mixture, shoot P content in target maize increased with soil P supply, but not with the neighbour presence; the interactive effect was not significant (Fig. 2b; Table S1). The phosphorus content in shoots of target maize was higher in the maize/faba bean than maize/maize mixtures at 36 and 270 mg kg<sup>-1</sup> (soil Olsen-P levels). When comparing shoot P content in target maize between the maize/maize and maize/faba bean treatments, we found a significant main effect of P supply and neighbour, as well as their interaction (Fig. 2b; Table S1). The interaction was significant because the effect of faba bean on increasing maize shoot P content (compared to competing maize plants) became more pronounced as soil P supply increased (Fig. 3).

The RII of maize grown with maize decreased with soil P supply, whereas in the maize/faba bean mixture, RII of target maize increased from 3.5 to 36 mg Olsen P kg<sup>-1</sup> and remained unchanged with further increases in soil P (Fig. 3). RII of maize in maize/maize was significantly lower than that of maize grown with faba bean, except in the lowest P treatment (3.5 mg Olsen P kg<sup>-1</sup>; Fig. 3). Compared with the maize/maize mixture, the RII in maize/faba bean was significantly greater in treatments with Olsen P at 36, 51, 123 and 270 mg kg<sup>-1</sup>.

The root length of target maize in the maize/faba bean or maize/maize mixture significantly increased with an increase in P supply (Fig. 4; Table S1). In the maize/faba bean mixture,

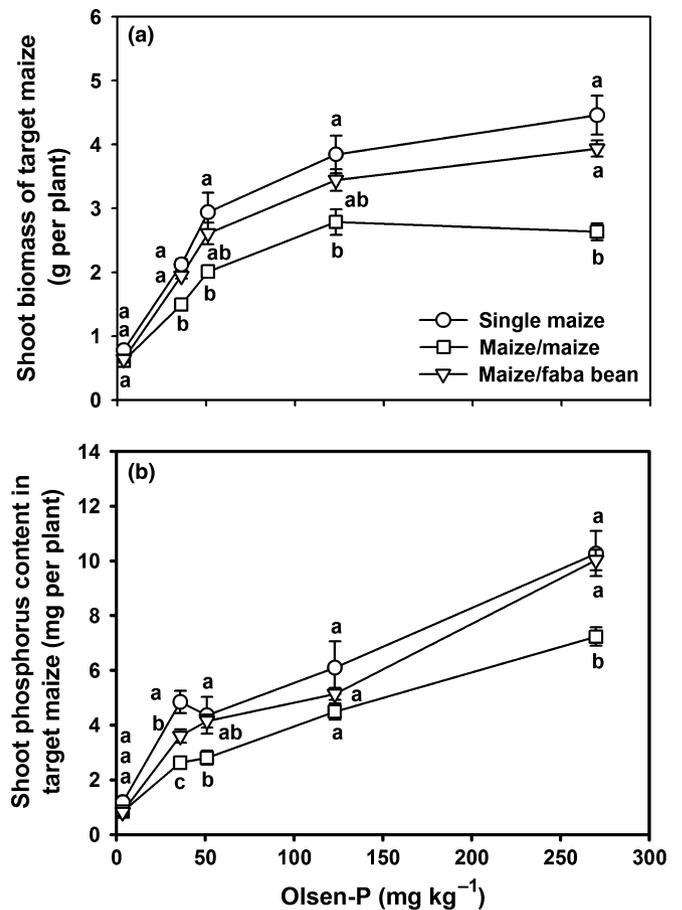
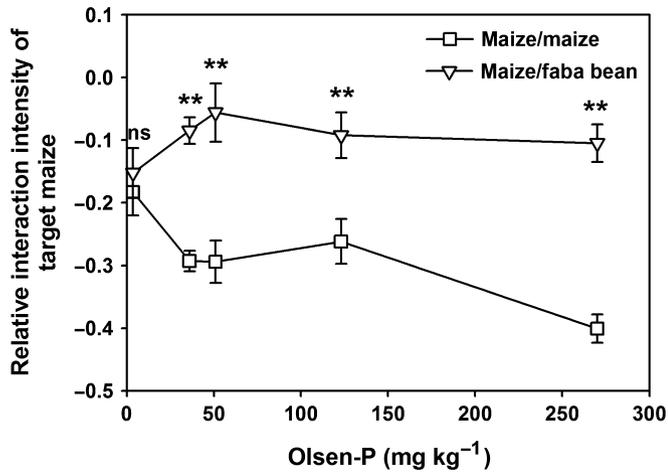


Fig. 2 Expt 1: effects of neighbour on shoot biomass (a) and shoot phosphorus (P) content (b) of target maize grown at five levels of P supply. Error bars,  $\pm$  SE. Within a P supply, different letters denote significant differences among target maize data in the single maize, maize/maize and maize/faba bean treatments ( $P \leq 0.05$ ).

root length reached the maximum at 51 mg Olsen P kg<sup>-1</sup> and remained similar at higher P supplies (Fig. 4), with both the neighbour main effect and the interaction being nonsignificant (Fig. 4; Table S1). The root length of maize when competing with maize attained the highest value at 123 mg Olsen P kg<sup>-1</sup>, and was significantly influenced by soil P supply and neighbour, but there was no interactive effect (Fig. 4; Table S1). Compared with the maize/maize mixture, the root length of maize in maize/faba bean was higher (Fig. 4). The difference in root length of target maize between maize/faba bean and maize/maize mixtures depended on soil P supply, competing plant species and the interaction (Fig. 4; Table S1), with the effect of faba bean on increasing maize root length (relative to competing maize plants) being largest at intermediate P supply (36–51 mg Olsen P kg<sup>-1</sup>; Fig. 4).

For the neighbouring species (faba bean or maize), root length was significantly lower in faba bean than maize, except at 3.5 mg Olsen P kg<sup>-1</sup> (Fig. 5a). There was no effect of P supply level on root length of neighbouring faba bean, but root length of neighbouring maize significantly increased with increasing P supply (Fig. 5a).

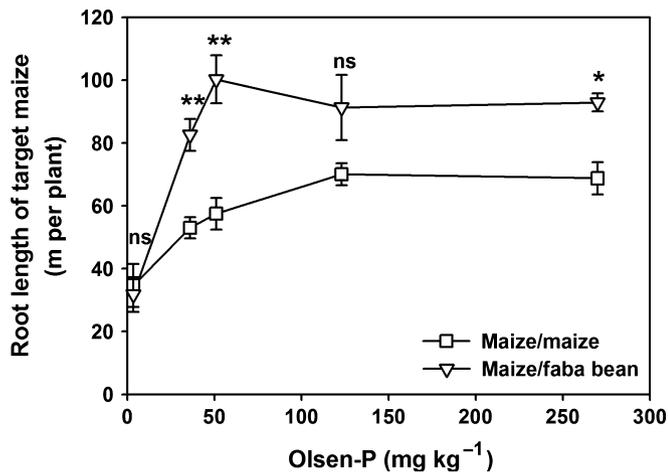


**Fig. 3** Expt 1: relative interaction intensity (RII) of target maize with its neighbours in the maize/maize and maize/faba bean mixtures. Error bars,  $\pm$  SE. Within a phosphorus (P) supply, asterisks denote significant differences between the maize/maize and maize/faba bean mixtures: \*\*,  $P < 0.01$ ; ns, not significant.

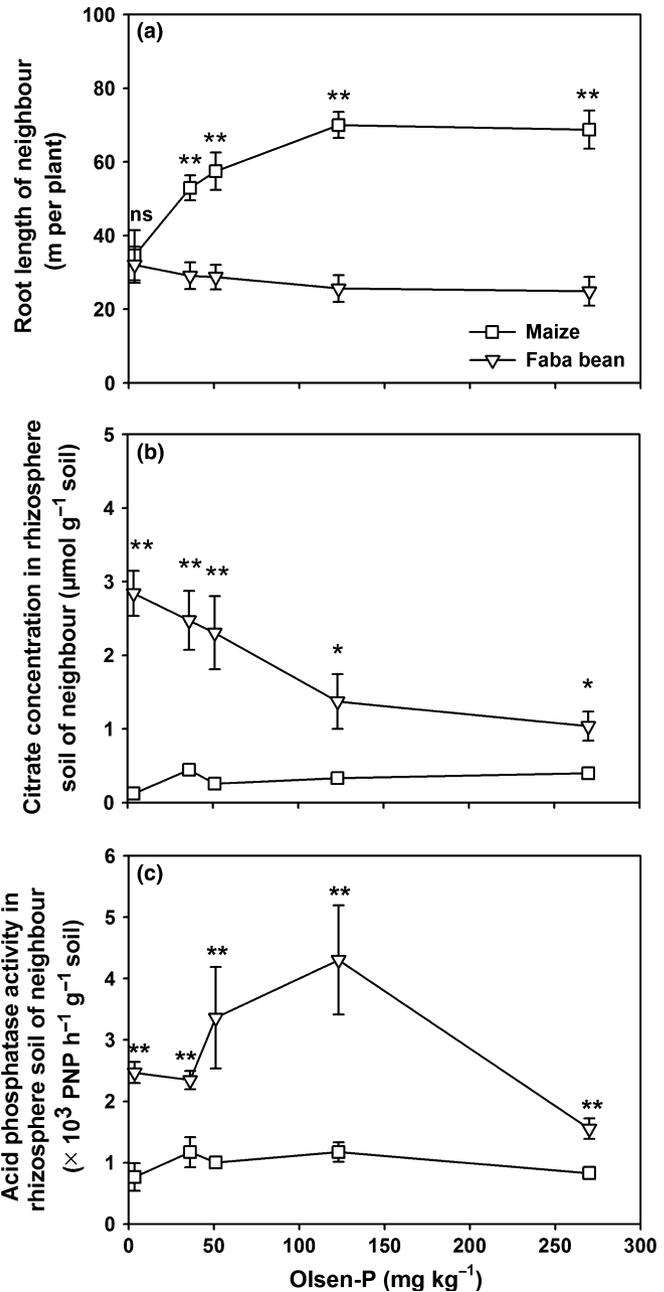
Citrate concentration and the acid phosphatase activity were significantly greater in the rhizosphere of faba bean than maize (Fig. 5b,c). Increased P supply depressed citrate exudation by faba bean. By contrast, the activity of acid phosphatase in the faba bean rhizosphere was increased from the lowest P treatment ( $3.5 \text{ mg Olsen P kg}^{-1}$ ) to  $123 \text{ mg Olsen P kg}^{-1}$ , and then decreased with a further increase in P supply. There were no evident effects of P treatments on citrate exudation and secretion of acid phosphatase in maize, which were at relatively low values compared with faba bean (Fig. 5b,c).

The effect of spatial variability in P supply on root interactions between maize and faba bean (Expt 2)

The shoot biomass of target maize in the maize/maize and maize/faba bean mixtures was significantly stimulated by heterogeneous P supply, although the total amount of P applied was the same



**Fig. 4** Expt 1: effects of neighbour on root length of target maize grown at five levels of phosphorus (P) supply. Error bars,  $\pm$  SE. Within a P supply, asterisks denote significant differences between the maize/maize and maize/faba bean mixtures: \*,  $P \leq 0.05$ ; \*\*,  $P < 0.01$ ; ns, not significant.



**Fig. 5** Expt 1: root length (a), and citrate concentration (b) and acid phosphatase activity (c) in the rhizosphere of neighbours (maize and faba bean). Error bars,  $\pm$  SE. Within a phosphorus (P) supply, asterisks denote significant differences between neighbouring species (maize vs faba bean): \*,  $P \leq 0.05$ ; \*\*,  $P < 0.01$ ; ns, not significant.

for the homogeneous and heterogeneous treatments. A similar trend was found in the single maize treatment, but the differences were not significant (Fig. 6a). When soil P was supplied heterogeneously, the biomass of target maize was 100% higher when grown with faba bean than with maize.

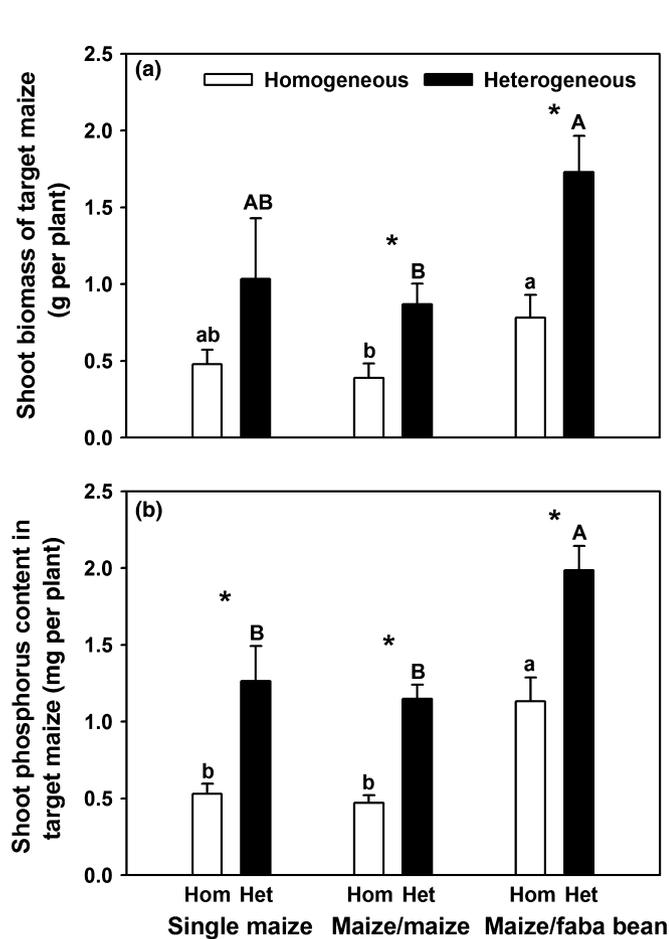
Soil P heterogeneity and neighbouring species both influenced shoot P content in target maize (Fig. 6b; Table S2). Compared to homogeneous P supply, the P content in shoots of target maize in the heterogeneous environment increased by 139%, 144% and 75% in single maize, maize/maize and maize/faba bean,

respectively (Fig. 6b). In the maize/maize mixture, the shoot P content of target maize was influenced by P heterogeneity only (Fig. 6b; Table S2). However, in mixture with faba bean, it was significantly affected by both P heterogeneity and neighbouring species (faba bean), but there was no interactive effect (Fig. 6b; Table S2). When the two mixtures were compared, shoot P content in target maize was higher in the maize/faba bean than maize/maize mixtures, which was influenced by soil P supply pattern and neighbouring species, but the interaction was not significant (Fig. 6b; Table S2). Thus, the shoot P content of maize was significantly increased by heterogeneous P supply, as well as the presence of faba bean, compared with the maize/maize system.

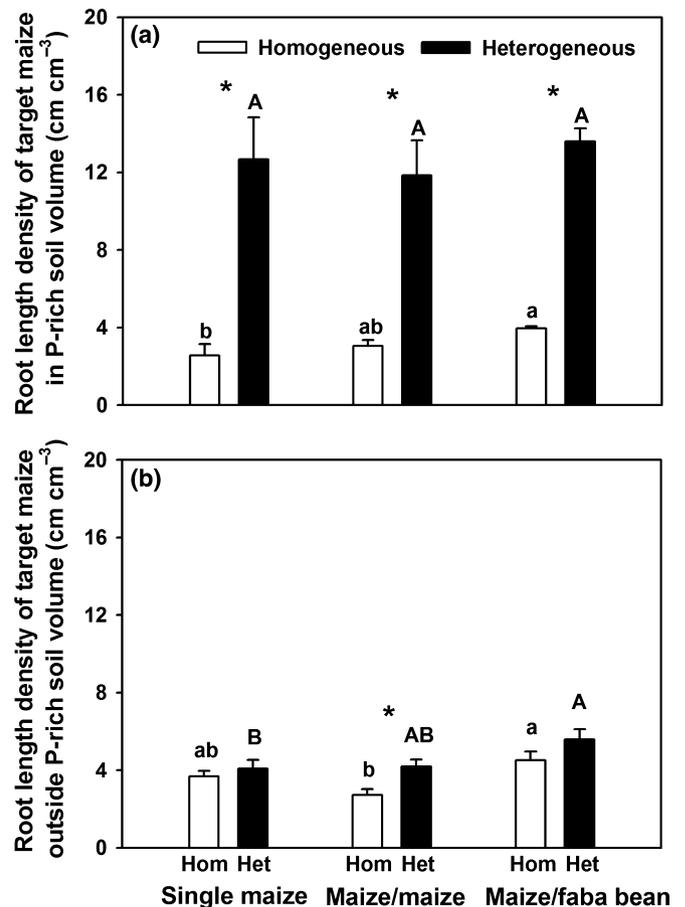
Root length density (RLD) of target maize was greater in the P-rich soil volume in the heterogeneous treatment compared with the same position in the homogeneous treatment (Fig. 7a). In the heterogeneous environment, RLD of target maize did not differ among the single maize, maize/maize and

maize/faba bean treatments, whereas RLD of maize grown in homogeneous environment was higher in the maize/faba bean than single maize treatment. The root length density (RLD) of target maize in the P-rich soil volume was significantly influenced by heterogeneous P supply and not by neighbour competition in either maize/maize or maize/faba bean systems (Fig. 7a; Table S2).

Outside the P-rich soil volume, RLD of target maize was increased significantly with heterogeneous compared with homogeneous P supply in the maize/maize treatment, but the effect was not significant in the single maize and maize/faba bean treatments (Fig. 7b). The increased RLD outside the P-rich soil volume of maize in maize/maize was caused by heterogeneity, but there was no effect of neighbour presence (Fig. 7b; Table S2). The heterogeneous P supply increased maize RLD outside the P-rich soil volume in maize/faba bean in comparison with single maize, but there was no significant difference between the maize/



**Fig. 6** Expt 2: effects of homogeneous vs heterogeneous phosphorus (P) supply on shoot biomass (a) and shoot P content (b) of target maize. Hom, homogeneous P supply; Het, heterogeneous P supply. Error bars, + SE. Different lowercase letters denote significant differences among target maize ( $P \leq 0.05$ ) data in the single maize, maize/maize and maize/faba bean treatments in the homogeneous treatments, and capitals denote significant differences in the heterogeneous treatments. For each parameter and species treatment, *t*-tests were run separately to detect difference between homogeneous and heterogeneous treatments; asterisks indicate significant differences: \*,  $P \leq 0.05$ .



**Fig. 7** Expt 2: effects of homogeneous/heterogeneous phosphorus (P) supply on target maize root length density within (a) and outside P-enriched soil volume (b). Hom, homogeneous P supply; Het, heterogeneous P supply. Error bars, + SE. Different lowercase letters denote significant differences among target maize ( $P \leq 0.05$ ) data in single maize, maize/maize and maize/faba bean mixtures in the homogeneous treatments, and uppercase letters denote significant differences in the heterogeneous treatments. For each soil volume and each species treatment, *t*-tests were run separately to detect significant difference between homogeneous and heterogeneous treatments; asterisks indicate significant differences: \*,  $P \leq 0.05$ .

faba bean and maize/maize mixtures (Fig. 7b). In the maize/faba bean system, maize RLD outside the P-rich soil volume was enhanced by neighbour presence, but not by heterogeneity (Fig. 7b; Table S2). Compared with the maize/maize and maize/faba bean treatments, the RLD of target maize outside the P-rich soil volume was influenced by soil P heterogeneity and neighbouring species (Fig. 7b; Table S2).

## Discussion

In the present study, the maize/faba bean treatment was compared to maize alone or maize/maize treatment. Any neighbours with maize would potentially represent competition, but our findings indicated that neighbouring faba bean was less competitive than neighbouring maize (Fig. 3). With increasing P supply level, the relative interaction intensity (RII) of target maize in maize/maize mixtures significantly decreased (Fig. 3), suggesting an increase in competition intensity with soil P supply increasing from 3.5 to 270 mg Olsen-P kg<sup>-1</sup>. According to Hess & de Kroon (2007), plants can sense and coordinate root growth based on available soil volume and a nutrient supply pattern. In the present study, maize shoot growth was P-limited at low P supply (Fig. 2a), which was also consistent with short root length (Fig. 4), resulting in low competition for soil P in the maize/maize mixtures. However, as root growth significantly increased with an increase in soil P supply level, the intensity of maize competition in the maize/maize mixtures for the same available soil volume was strengthened, resulting in a more negative RII and thus high competition at higher soil P supply.

In the maize/faba bean mixture, the competition intensity remained unimodal and was lower in the maize/faba bean than maize/maize system at the same soil P supply (Fig. 3). The effect of soil P availability on shoot P content and root length of target maize was significantly dependent on neighbouring species, and was greater in the maize/faba bean than maize/maize mixtures (Table S1). Target maize shoot P content was significantly decreased by neighbouring maize ( $P < 0.001$ ) but not faba bean ( $P = 0.071$ ; Table S1), suggesting that the presence of faba bean resulted in lower competition intensity than the presence of maize. Previous studies have indicated that plant species win competition for a shared resource by using the resources available in hotspots more rapidly as a result of greater root growth plasticity (Grime, 1994; Craine *et al.*, 2005), or by depleting the resource to lower concentrations than their competitors (Tilman, 1982). In the present study, the difference between root competition in the maize/faba bean and maize/maize mixtures could be explained in two ways. First, the lower root length of faba bean compared to neighbouring maize could provide a greater available soil volume for target maize roots to exploit, indicating the competitive importance of morphological root responses to the abiotic environment (similarly, plasticity in root morphology of *Pseudoroegneria spicata* improved its capacity to withstand competition from the more vigorous, but less plastic, *Agropyron desertorum*; Huber-Sannwald *et al.*, 1996). Second, a larger amount of citrate and acid phosphatase in the faba bean rhizosphere soil would have improved mobilization of sparingly-

soluble soil P, thus effectively increasing the amount of accessible P for target maize (see Hinsinger, 2001; Jones *et al.*, 2003). Hence, nutrient mobilization by root exudation could lead to increased acquisition of limiting resources by decreasing the intensity of interspecies competition through niche complementarity (i.e. maize and faba bean accessing different P fractions: it was found that faba bean accessed sparingly soluble P (unavailable to maize) through root exudation, but maize mainly used soluble soil P, indicating a niche complementarity; Hinsinger *et al.*, 2011; Shen *et al.*, 2011, 2013; Brooker *et al.*, 2014; Li *et al.*, 2014b). This result supports Hypothesis 1 – that neighbouring faba bean provides low competition intensity to target maize by its relatively low root morphological plasticity, which was compensated for by high physiological plasticity (strong exudation) to alter soil P availability in the maize/faba bean system.

In response to heterogeneous P supply in the present study, maize showed significant root proliferation in the nutrient-rich soil volume (Fig. 7a), which conferred competitive advantage to maize in heterogeneous compared to homogeneous nutrient supply (see also Robinson *et al.*, 1999); hence, shoot P content in maize was greater in the heterogeneous than homogeneous P treatments in the maize/faba bean mixture (Fig. 6b). The reasons why faba bean contributed to increased shoot P content in maize in heterogeneous P supply could be explained as follows. First, heterogeneous P supply increased maize root proliferation (but not that of faba bean, data not shown) in P-enriched soil (Fig. 7), as in the earlier study (Li *et al.*, 2014a) in which heterogeneous nutrient supply did not influence faba bean root growth in the single faba bean treatments. Increased maize root proliferation in the localized fertilizer zone could cause higher competition intensity in maize/maize compared with maize/faba bean. Second, faba bean roots were located mostly outside P-enriched zones (i.e. in soil not fertilized with P), and shoot biomass and P content in faba bean were similar in heterogeneous and homogeneous P supply treatments (data not shown), which is consistent with the previous findings that shoot growth and nutrient uptake of single faba bean were not influenced by heterogeneous P availability (Li *et al.*, 2014a).

The high physiological plasticity of faba bean in terms of exudation of carboxylates and acid phosphatase could mobilize soil P and increase P availability to provide adequate P supply for its own growth and could even contribute to satisfying demand of neighbouring plants (see Expt 1). A shallow root system of faba bean (Li *et al.*, 2006) provided a niche for maize roots to increase root length density (Fig. 7b) and P uptake (Fig. 6b) compared with maize in the maize/maize treatment. Hence, increased P uptake by maize may be attributed to root interactions between maize and faba bean being facilitated in the heterogeneous P supply treatment. The results indicated that the increased soil P availability induced by physiological root plasticity of faba bean stimulated morphological root plasticity in maize, resulting in improved maize growth and P uptake. By contrast, in the maize/maize mixture, high morphological root plasticity caused strong competition, further intensified in the heterogeneous P treatment by greater root proliferation in the P-rich patches, and resulting in lower shoot growth and nutrient uptake compared with maize

grown with faba bean. The results provide support for Hypothesis 2 – that maize would benefit from the root physiological traits of neighbouring faba bean as well as from heterogeneous P supply.

Even though the present study showed that root interactions could be significantly influenced by heterogeneous P supply and root traits of neighbouring species in the maize/faba bean and maize/maize mixtures, we do not know yet how common such interactions might be in other agroecological systems. In maize/faba bean intercropping, a more asymmetric root distribution between maize and faba bean would occur under heterogeneous than homogeneous nutrient supply (cf. Schenk, 2006), probably resulting in an increasing potential for spatial niche complementarity as mentioned above. Interspecies facilitation based on nutrient-mobilizing mechanisms (Shen *et al.*, 2011, 2013; Li *et al.*, 2014b) may be used to help explore potential P resources in soils through selecting neighbouring species or optimizing plant interactions based on root proliferation and the rhizosphere processes to alter competition. Alternatively, a plant species can achieve competitive superiority by depleting nutrient resources to lower concentrations than its competitors (Tilman, 1982). In the present study, maize had competitive superiority for nutrient acquisition not only by stimulating root growth in the presence of faba bean, but also by exploiting additional nutrient resources made available in the rhizosphere soil of faba bean. Nutrient availability (increased by neighbouring faba bean plants) modified maize root growth and thus changed root interactions in the maize/faba bean mixture. This study provided novel support for the nutrient-driven root interactions regulating growth dynamics of plant species in the mixed stand. Furthermore, heterogeneous nutrient supply could be considered a useful strategy for modifying root/rhizosphere interactions to optimize plant combinations and underpin improved crop productivity and nutrient uptake in agroecological systems.

## Conclusions

Both nutrient supply and neighbouring species significantly modified root interactions between maize and faba bean, influencing P uptake and biomass production. At different soil P supply, faba bean exhibited relatively poor morphological, but strong physiological root plasticity. This contributed to relatively low competition intensity in the maize/faba bean compared with the maize/maize mixture. Localized P supply enhanced root proliferation of target maize (but not faba bean) in the maize/faba bean mixture. Enhanced root exudation of citrate and acid phosphatase by faba bean could facilitate soil P availability to benefit maize growth in the maize/faba bean mixture. The study provided new insights into root/rhizosphere interactions in the maize/faba bean intercropping in the systems with variable P supply, which is important in developing strategies for rhizosphere management through optimizing plant combinations and soil nutrient supply to increase crop productivity and nutrient-use efficiency. The challenge of enhancing food supply without using extra nutrients could be accomplished by maximizing the efficacy of

intercropping as a means of delivering more crop production per unit of fertilizer applied.

## Acknowledgements

This study was supported by the National Natural Science Foundation of China (31330070, 30925024, 31210103906), the National Basic Research Program (973-2015CB150405) and the Innovative Group Grant of the National Science Foundation of China (31421092). Z.R. is supported by Australian Research Council (DP130104825). W.R.W. is supported at Rothamsted Research by the 20 : 20 Wheat<sup>®</sup> project.

## References

- Alvey S, Bagayoko M, Neumann G, Buerkert A. 2000. Cereal/legume rotation effects in two West African soils under controlled conditions. *Plant and Soil* 231: 45–54.
- Andersen SN, Dresbøll DB, Thorup-Kristensen K. 2014. Root interactions between intercropped legumes and non-legumes – a competition study of red clover and red beet at different nitrogen levels. *Plant and Soil* 378: 59–72.
- Brooker RW, Bennett AE, Cong WF, Daniell TJ, George TS, Hallett PD, Hawes C, Iannetta PPM, Jones HG, Karley AJ *et al.* 2014. Improving intercropping: a synthesis of research in agronomy, plant physiology and ecology. *New Phytologist* 3: 1–11.
- Cahill JF, McNickle GG. 2011. The behavioral ecology of nutrient foraging by plants. *Annual Review of Ecology, Evolution, and Systematics* 42: 289–311.
- Cahill JF, McNickle GG, Haag JJ, Lamb EG, Nyanumba SM, Clair CCS. 2010. Plants integrate information about nutrients and neighbors. *Science* 328: 1657.
- Craine JM, Fargione J, Sugita S. 2005. Supply preemption, not concentration reduction, is the mechanism of competition for nutrients. *New Phytologist* 166: 933–940.
- Drew MC. 1975. Comparison of the effects of localized supply of phosphorus, nitrate, ammonium and potassium on the growth of the seminal root system, and the shoot, in barley. *New Phytologist* 75: 479–490.
- Dunbabin V, Diggle A, Rengel Z. 2003. Is there an optimal root architecture for nitrate capture in leaching environments? *Plant, Cell & Environment* 26: 835–844.
- Fransen B, de Kroon H, Berendse F. 2001. Soil nutrient heterogeneity alters competition between two perennial grass species. *Ecology* 82: 2534–2546.
- Grime JP. 1994. The role of plasticity in exploiting environmental heterogeneity. In: Caldwell MM, Pearcy RW, eds. *Exploitation of environmental heterogeneity by plants: ecophysiological processes above- and below-ground*. San Diego, CA, USA: Academic Press, 1–19.
- Hess L, de Kroon H. 2007. Effects of rooting volume and nutrient availability as an alternative explanation for root self/non-self discrimination. *Journal of Ecology* 95: 241–251.
- Hinsinger P. 2001. Bioavailability of soil inorganic P in the rhizosphere as affected by root-induced chemical changes: a review. *Plant and Soil* 237: 173–195.
- Hinsinger P, Betencourt E, Bernard L, Brauman A, Plassard C, Shen JB, Tang XY, Zhang FS. 2011. P for two, sharing a scarce resource: soil phosphorus acquisition in the rhizosphere of intercropped species. *Plant Physiology* 156: 1078–1086.
- Hodge A. 2004. The plastic plant: root responses to heterogeneous supplies of nutrients. *New Phytologist* 162: 9–24.
- Hodge A, Robinson D, Griffiths BS, Fitter AH. 1999. Why plants bother: root proliferation results in increased nitrogen capture from an organic patch when two grasses compete. *Plant, Cell & Environment* 22: 811–820.
- Howard TG, Goldberg DE. 2001. Competitive response hierarchies for germination, growth, and survival and their influence on abundance. *Ecology* 82: 979–990.

- Huber-Sannwald E, Pyke DA, Caldwell MM. 1996. Morphological plasticity following species-specific recognition and competition in two perennial grasses. *Annals of Botany* 83: 919–931.
- Hutchings MJ, de Kroon H. 1994. Foraging in plants: the role of morphological plasticity in resource acquisition. *Advances in Ecological Research* 25: 159–238.
- Jackson RB, Caldwell MM. 1996. Integrating resource heterogeneity and plant plasticity: modelling nitrate and phosphate uptake in a patchy soil environment. *Journal of Ecology* 84: 891–903.
- Jackson RB, Manwaring JH, Caldwell MM. 1990. Rapid physiological adjustment of roots to localized soil enrichment. *Nature* 344: 58–60.
- Jing JY, Rui Y, Zhang FS, Rengel Z, Shen JB. 2010. Localized application of phosphorus and ammonium improves growth of maize seedlings by stimulating root proliferation and rhizosphere acidification. *Field Crops Research* 119: 335–364.
- Johnson CM, Ulrich A. 1959. *Analytical methods for use in plant analysis*. Berkeley, CA, USA: University of California, Agricultural Experiment Station.
- Jones DL, Dennis PG, Owen AG, van Hees PAW. 2003. Organic acid behaviour in soils: misconceptions and knowledge gaps. *Plant and Soil* 248: 31–41.
- Lambers H, Shane MW, Cramer MD, Pearse SJ, Veneklaas EJ. 2006. Root structure and functioning for efficient acquisition of phosphorus: matching morphological and physiological traits. *Annals of Botany* 98: 693–713.
- Li HB, Ma QH, Li HG, Zhang FS, Rengel Z, Shen JB. 2014a. Root morphological responses to localized nutrient supply differ among crop species with contrasting root traits. *Plant and Soil* 376: 151–163.
- Li HG, Shen JB, Zhang FS, Tang CX, Lambers H. 2008. Is there a critical level of shoot phosphorus concentration for cluster-root formation in *Lupinus albus*? *Functional Plant Biology* 35: 328–336.
- Li L, Li SM, Sun JH, Zhou LL, Bao XG, Zhang HG, Zhang FS. 2007. Diversity enhances agricultural productivity via rhizosphere phosphorus facilitation on phosphorus-deficient soils. *Proceedings of the National Academy of Sciences, USA* 104: 11 192–11 196.
- Li L, Sun JH, Zhang FS, Guo TW, Bao XG, Smith FA, Smith SE. 2006. Root distribution and interactions between intercropped species. *Oecologia* 147: 280–290.
- Li L, Tilman D, Lambers H, Zhang FS. 2014b. Biodiversity and overyielding: insights from below-ground facilitation of intercropping in agriculture. *New Phytologist* 203: 63–69.
- Li L, Zhang FS, Li XL, Christie P, Yang SC, Tang CX. 2003. Interspecific facilitation of nutrient uptakes by intercropped maize and faba bean. *Nutrient Cycling in Agroecosystems* 65: 61–71.
- Markham JH, Chanway CP. 1996. Measuring plant neighbour effects. *Functional Ecology* 10: 548–549.
- Mommer L, Van Ruijven J, Jansen C, Van de Steeg HM, de Kroon H. 2012. Interactive effects of nutrient heterogeneity and competition: implications for root foraging theory? *Functional Ecology* 26: 66–73.
- Robinson D. 1996. Resource capture by localized root proliferation: why do plants bother? *Annals of Botany* 77: 179–185.
- Robinson D, Hodge A, Griffiths BS, Fitter AH. 1999. Plant root proliferation in nitrogen-rich patches confers competitive advantage. *Proceedings of the Royal Society of London* 266: 431–435.
- Schenk HJ. 2006. Root competition: beyond resource depletion. *Journal of Ecology* 94: 725–739.
- Shen JB, Li CJ, Mi GH, Li L, Yuan LX, Jiang RF, Zhang FS. 2013. Maximizing root/rhizosphere efficiency to improve crop productivity and nutrient use efficiency in intensive agriculture of China. *Journal of Experimental Botany* 64: 1181–1192.
- Shen JB, Li HG, Neumann G, Zhang FS. 2005. Nutrient uptake, cluster root formation and exudation of protons and citrate in *Lupinus albus* as affected by localized supply of phosphorus in a split-root system. *Plant Science* 168: 837–845.
- Shen JB, Rengel Z, Tang CX, Zhang FS. 2003. Role of phosphorus nutrition in development of cluster roots and release of carboxylates in soil-grown *Lupinus albus*. *Plant and Soil* 248: 199–206.
- Shen JB, Yuan LX, Zhang JL, Li HG, Bai ZH, Chen XP, Zhang WF, Zhang FS. 2011. Phosphorus dynamics: from soil to plant. *Plant Physiology* 156: 997–1005.
- Tilman D. 1982. *Resource competition and community structure*. Monographs in population biology. Princeton, NJ, USA: Princeton University Press.
- Valladares F, Sanchez D, Zavala MA. 2006. Quantitative estimation of phenotypic plasticity: bridging the gap between the evolutionary concept and its ecological applications. *Journal of Ecology* 94: 1103–1116.
- Wang BL, Tang XY, Cheng LY, Zhang AZ, Zhang WH, Zhang FS, Liu JQ, Cao Y, Allan DL, Vance CP *et al.* 2010. Nitric oxide is involved in phosphorus deficiency-induced cluster-root development and citrate exudation in white lupin. *New Phytologist* 187: 1112–1123.
- Weigelt A, Jolliffe P. 2003. Indices of plant competition. *Journal of Ecology* 91: 707–720.
- Wilson SD, Keddy PA. 1986. Measuring diffuse competition along an environmental gradient: results from a shoreline plant community. *The American Naturalist* 127: 862–869.
- Zhang FS, Shen JB, Zhang JL, Zuo YM, Li L, Chen XP. 2010. Rhizosphere processes and management for improving nutrient use efficiency and crop productivity: implications for China. In: Sparks DL, ed. *Advances in agronomy*. Burlington, MA, USA: Academic Press, 107: 1–32.

## Supporting Information

Additional supporting information may be found in the online version of this article.

**Table S1** Effects of soil P availability and the neighbour (maize or faba bean) presence on shoot P content and total root length of target maize (*Zea mays*) (Expt 1)

**Table S2** Effects of soil P heterogeneity and the neighbour (maize or faba bean) presence on target maize shoot P content and root length density inside and outside the P-rich soil volume (Expt 2)

Please note: Wiley Blackwell are not responsible for the content or functionality of any supporting information supplied by the authors. Any queries (other than missing material) should be directed to the *New Phytologist* Central Office.