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1 *Running head:* Using traits to model plant communities

2 Using functional traits to model annual plant community dynamics

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## 6 Abstract

7 Predicting the response of biological communities to changes in the environment or management  
8 is a fundamental pursuit of community ecology. Meeting this challenge requires the integration  
9 of multiple processes: habitat filtering, niche differentiation, biotic interactions, competitive ex-  
10 clusion, and stochastic demographic events. Most approaches to this long-standing problem focus  
11 either on the role of the environment, using trait-based filtering approaches, or on quantifying bi-  
12 otic interactions with process-based community dynamics models. We introduce a novel approach  
13 that uses functional traits to parametrise a process-based model. By combining the two approaches  
14 we make use of the extensive literature on traits and community filtering as a convenient means  
15 of reducing the parametrisation requirements of a complex population dynamics model whilst re-  
16 taining the power to capture the processes underlying community assembly. Using arable weed  
17 communities as a case study, we demonstrate that this approach results in predictions that show  
18 realistic distributions of traits and that trait selection predicted by our simulations is consistent  
19 with in-field observations. We demonstrate that trait-based filtering approaches can be combined  
20 with process-based models to derive the emergent distribution of traits. While initially developed  
21 to predict the impact of crop management on functional shifts in weed communities, our approach  
22 has the potential to be applied to other annual plant communities if the generality of relationships  
23 between traits and model parameters can be confirmed.

24 *Key words: arable weeds, annual plants, community dynamics, ecological function, environmental fil-*  
25 *tering, functional diversity, functional traits, population dynamics modelling*

## 26 Introduction

27 Predicting the assembly of biological communities and their resulting ecological function in dif-  
28 ferent environments is a fundamental pursuit of community ecologists and has been characterised  
29 as the Holy Grail of ecology (Lavorel & Garnier, 2002). As society increasingly recognises the  
30 ecosystem services the biosphere contributes to human survival and well-being (Carpenter *et al.*,  
31 2006) the need to understand the impact of changes in environment, land-use, or management  
32 on biological communities has become more urgent. Within this ecosystem service framework,  
33 it is more important to predict the impact of change on the functioning of the emergent biolog-  
34 ical community than on taxonomic composition (Fig. 1 A; Díaz *et al.*, 2007a). Meeting this  
35 challenge requires a unified approach that combines the theories of 1) habitat filtering and niche  
36 differentiation, 2) biotic interactions and competitive exclusion, and 3) stochastic demographic  
37 events (neutral theory). These processes, together with historical and evolutionary factors (which  
38 determine the regional species pool) all play a role in determining the local ecological community  
39 in a given environment (D'Amen *et al.*, 2017). Most approaches to this long-standing problem of  
40 predicting community composition at a given location focus either on the role of the environment,  
41 using trait-based filtering approaches (Fig. 1 B), or instead focus on quantifying biotic interactions  
42 with process-based community dynamics models (Fig. 1 C).

43 Trait-based filtering approaches that identify the abiotic and biotic filters acting on regionally  
44 available pools of species and determine those with favourable combinations of traits that can per-  
45 sist in a given habitat (Keddy, 1992) have now been applied across several taxa (e.g. plants (da  
46 Silveira Pontes *et al.*, 2010), arthropods (Braaker *et al.*, 2017), and bees (Hoiss *et al.*, 2012)),  
47 in a range of environments (e.g. tropics (Lebrija–Trejos *et al.*, 2010), streams (Poff, 1997), and  
48 rangelands (Bernard–Verdier *et al.*, 2012)) and across a number of different gradients (e.g. grazing  
49 (Díaz *et al.*, 2007b), geo-morphological (Gilardelli *et al.*, 2015), and aridity (Gross *et al.*, 2013)).  
50 However, all these studies rely on fitting statistical models to empirical relationships between envi-  
51 ronmental gradients and functional trait metrics and are, therefore, limited in their power to predict  
52 responses to environments with a novel combination of environmental variables. These models

53 typically predict a convergence of trait attributes, as only species which are functionally similar  
54 will pass through successive filters on plant traits.

55 An alternative approach that avoids these limitations is to build process-based models of the  
56 responses of multiple interacting species to the environment. This more mechanistic approach  
57 involves describing key life-cycle processes mathematically, often from first principles, and can  
58 include spatially explicit individual based modelling approaches. Such process-based community  
59 dynamics models have also been widely developed to predict the community composition of a  
60 number of taxa (e.g. fish (Shin and Cury, 2001), coral (Langmead and Sheppard, 2004. ), and  
61 trees (Purves *et al.*, 2008)), in a range of environments (e.g. tundra (Gilg *et al.*, 2003), freshwater  
62 lakes (van Nes *et al.*, 2002), and forests (Botkin, 1993)) and across a number of different envi-  
63 ronmental gradients (e.g. disturbance (Matsinos and Troumbis, 2002), fire (Thonickeet al., 2001),  
64 and nutrient limitation (Moore *et al.*, 2004)). In contrast to the trait-based filtering approach, these  
65 process-based community dynamics models often focus on biotic interactions which can be de-  
66 scribed mathematically and aim to predict relative species abundances in a more mechanistic way.  
67 By focussing on competitive processes, these models tend to select for species with divergent trait  
68 attributes in order to minimise overlapping resource use and competition, although practically this  
69 may not always be the observed outcome (Mayfield and Levine, 2010). Process-based community  
70 dynamics models often require extensive parametrisation to capture all the ecologically impor-  
71 tant processes. Each aspect of the life-cycle must be described mathematically for each simulated  
72 species, and where there is asymmetric competition for multiple resources this must also be quan-  
73 tified. As such, these models tend to be limited to a small pool of species and to a particular  
74 environment in which the parametrisation has been conducted (da Silveira Pontes *et al.*, 2010).

75 Ecological communities lie on a continuum: from those with strong biotic interactions to those  
76 where local interactions between individuals are weak and few (Cornell and Lawton, 1992) and  
77 models that aim to predict community dynamics should ideally avoid making prior assumptions  
78 on the dominant processes shaping that community. Several attempts have been made to include  
79 biotic processes into trait-based filtering models in order to simulate both the convergence and

80 divergence of traits, and eliminate the need for *a-priori* knowledge of the dominant processes  
81 driving community dynamics at a given location. For example, Shipley *et al.* (Maxent, 2006) and  
82 later Laughlin *et al.* (Traitspace, 2012), developed generic models based on the trait-based filtering  
83 approach but limited convergence by selecting the community with the maximum Shannon index  
84 of all possible outcomes based on the environmental filtering step. Whilst these two models go  
85 some way to reconciling the role of trait-based filtering and competition in predicting community  
86 composition, they are both based on empirical relationships between observed trait distributions  
87 and environmental gradients. A valuable addition to these approaches would be to derive models  
88 that predict shifts in trait distributions in a changing environment from first principles (Laughlin &  
89 Laughlin, 2013).

90 Here we introduce a model which uses functional traits to parametrise a process-based model  
91 (Fig. 1 D), using arable weed communities as a case study. The immediate questions the model  
92 is designed to address are to do with an impact of a change in crop management on the functional  
93 composition of weed communities. However, the model structure is generic to any annual plant  
94 community. By combining the two approaches we make use of the extensive literature on traits  
95 and community filtering as a convenient means of reducing the parametrisation requirements of  
96 a complex population dynamics model whilst retaining the power to capture the processes under-  
97 lying community assembly. In so doing, we aimed for the optimal balance between complexity  
98 and tractability. Weeds are dominated by annual species making the generic life cycle model more  
99 tractable and, because of their economic importance, are highly studied with a rich literature of  
100 population dynamics models parametrised at the species level. The parameters of the system are  
101 also clearly defined by the management operations in the arena of a cropped field. The arable  
102 species pool is also sufficiently large to demonstrate the usefulness of a trait-based approach for  
103 model parametrisation (including a range of ecological strategies (Bourgeois *et al.*, 2019)), and,  
104 because it is dominated by annual species, responds to change on relatively short time scales. In ad-  
105 dition, the traits of arable weeds have been well-studied in recent years and trait-based approaches  
106 have quantified functional responses of weed communities to management filters (e.g. Fried *et al.*,

107 2009, Gardarin *et al.*, 2010, Gunton *et al.*, 2011, Fried *et al.*, 2012, Colbach *et al.*, 2014, Armengot  
108 *et al.*, 2016).

109 We used functional traits and groups to parametrise the species specific mechanistic processes  
110 within our model (Fig. 1 D). We wanted to keep the model parsimonious and so chose only four  
111 continuous traits (*sensu* Violle *et al.*, 2007): seed mass, maximum height, date of first flowering  
112 and specific leaf area. These four traits are readily available for many annual plants and have been  
113 shown to relate to many life-cycle process (Table 1). For example increasing seed mass is known  
114 to be associated with decreased seed production (Henery & Westoby, 2003). In addition we also  
115 assigned species to functional groups according to i) Ellenberg N number to model the impact  
116 of soil fertility on community dynamics, ii) emergence periodicity to model responses to changes  
117 in management timings, iii) seedbank type to model persistence in the soil, and iv) phylogeny:  
118 whether they were grasses or broadleaves as many of the relationships between other traits and the  
119 model parameters varied between these two groups.

120 We selected these traits based not only on their relationship with various life cycle processes,  
121 making them suitable predictors of our model parameters but also due to their availability within  
122 the literature. We chose to use only ‘soft traits’ (*sensu* Díaz *et al.*, 2004) which are more easily  
123 measured than ‘hard traits’ (which may be more directly related to the life-cycle process) and are  
124 well documented for a large range of annual plant species across a number of databases (e.g. TRY  
125 plant trait database (Kattge *et al.*, 2020), Seed Information Database (SID, 2018), Ecoflora (Fitter  
126 and Peat, 1994), and LEDA traitbase (Kleyer *et al.*, 2008)).

127 The quantification of relationships between functional groups, traits and model parameters is  
128 based on a series of experiments screening ecophysiological parameters for 21 annual weed species  
129 summarised in Storkey (2006).

## 130 **Methods**

131 We developed a model of the annual plant life cycle based on transitions between seedlings, ma-  
132 ture plants, fresh seed and seed in the seedbank (Fig. 1 C). Some of the processes governing the

133 transitions between these four life stages are influenced by biotic interactions as well as habitat fil-  
134 tering. For each transition (except for fresh seed to seedbank) there are one or more response traits  
135 that we anticipate will be selected for or against by environmental or management filters (Table  
136 1). These **response traits** (highlighted in bold throughout the methods section) are integrated into  
137 the simulation of mechanistic processes within the annual plant life cycle by quantifying relation-  
138 ships between traits and model parameters (see Appendix S1, Box S1 for a summary of the data  
139 sources used to parametrise our trait-response relationships). We fitted linear models to describe  
140 the relationships between the life cycle parameters of the simulation model and the weed traits  
141 (or functional groups) using GenSTAT. In each case this results in parameter estimates  $\{a, b\}$  and  
142 an associated covariance function  $C$  that captures the uncertainty in the estimates. The data we  
143 used to fit the models came from a series of experiments screening ecophysiological parameters  
144 for 21 annual weed species summarised in Storkey (2006). In our simulation model, we explic-  
145 itly account for the uncertainty in the relationships between the traits and life-cycle parameters by  
146 stochastically sampling the parameters values from multivariate normal distributions with mean  
147  $\{a, b\}$  and covariance  $C$ .

148 The weed life-cycle model proceeds as follows. For each weed species, the number of weed  
149 seedlings that emerge from the seedbank is calculated and this is converted to an initial estimate  
150 of green area. The green area increases as a function of thermal time up until the canopy reaches  
151 closure (which is defined as the total green area index, GAI, equalling 0.75). Thereafter the plants  
152 are assumed to grow in competition and both plant height and green area are monitored up until  
153 the crop matures to calculate partitioning of light in the canopy. At this stage, we calculate the  
154 total biomass for each species and use this to estimate seed production, a proportion of which is  
155 returned to the seedbank.

156 We integrated our model within an existing model of the agricultural landscape — the Rotham-  
157 sted Landscape Model (RLM, Coleman *et al.*, 2017) to define the environmental and management  
158 context of the simulation arena. RLM simulates soil processes, including water and nutrient flows,  
159 and the growth of arable crops. We use the soil and crop variables generated by the RLM as inputs

160 into our life-cycle model. This allows us to simulate the response of the weed community to vari-  
161 ous environmental factors (such as light and nutrient availability) as well as management (timings  
162 of cultivation and application of herbicides). The model runs on a daily time step driven by daily  
163 weather variables.

#### 164 **Seedbank** → **Seedlings**

165 **Seedling Emergence** The model is initialised by ‘planting’ a number of seeds per species in  
166 each of two layers of the seedbank (a shallow layer from which seeds can readily emerge and  
167 a deeper layer from which emergence is reduced). On the day on which the crop is ‘sown’ in  
168 RLM the weed-seedling emergence function is triggered. This function calculates the number of  
169 seedlings that emerge for each species. Firstly, we calculate the proportion of the total seedbank  
170 that can potentially emerge  $r_t$ . We model this as a generic, stochastic process across all species  
171 by drawing from a censored Weibull distribution (Eq. 1 with parameters  $a = 1.52$  and  $b =$   
172  $0.21$ ). This distribution was chosen as it gave a good description of data on seedling emergence  
173 observed at 5 sites over three years for three contrasting weed species (Appendix S1 Fig. S1.). This  
174 is a pragmatic approach that deliberately avoids the need to model interactions between season,  
175 induced dormancy and soil microclimate in determining emergence in any given year.

$$f(r_t) = \min\left(\frac{a}{b}r_t^{a-1}\exp\left(-\frac{r_t^a}{b}\right), 1\right) \text{ for } r_t > 0$$
$$= 0 \quad \text{elsewhere.} \quad (1)$$

176 Weeds are adapted to emerge at different times of the year. We use an emergence calendar  
177 for each species to describe this, and select the proportion of seeds ( $r_e$ ) predicted to emerge in the  
178 time period between sowing and when germination is inhibited by the crop canopy (45 days and  
179 30 days after sowing for autumn and spring sown crops respectively). It would be extremely costly  
180 to parameterise an emergence calendar for each species in turn. Instead, we use the functional

181 groupings according to **emergence periodicity**. Here, each species is assigned to one of three  
 182 groups: spring emergers, autumn emergers or generalists. We fitted bimodal normal probability  
 183 distributions to each group using data from Storkey *et al.* (2015), see Fig. 2.

184 We assume that the seeds in the deep layer of the seedbank have a reduced probability of  
 185 emergence. This is described by scaling the emergence of seeds from the bottom later by

$$r_d = \frac{1}{2} \left( \frac{D - 2}{8} \right). \quad (2)$$

186 where  $D$  is the maximum depth from which seeds of that species can germinate. The maximum  
 187 depth from which seeds of a given species can emerge ( $D$ ) is estimated using the **seed mass** trait  
 188 ( $S_m$ ). The linear relationship

$$D = c \ln(S_m) + d \quad (3)$$

189 was derived using data from Storkey *et al.* (2015) for 18 weed species (see Appendix S1, Figure  
 190 S2).

191 The number of seedlings that emerge for each species  $S_{em}$  is then given by:

$$S_{em} = (S_B r_d + S_T) r_t r_e \quad (4)$$

192 where  $S_B$  and  $S_T$  are the seeds in the deep and shallow layers of the seedbank respectively.

**Seedling mortality and seedbank decay** The numbers of seeds that persist in the deep and  
 shallow layers of the seedbank from one year ( $k$ ) to the next ( $k + 1$ ) are given by

$$S_B(k + 1) = \Delta [S_B(k) - S_B(k) r_d r_t (r_m + r_e) - l_g] \quad (5)$$

$$S_T(k + 1) = \Delta [S_T(k) - S_T(k) r_t (r_m + r_e)], \quad (6)$$

193 where  $r_m$  is the proportion of seedlings that are removed by pre-emergence control methods (either  
 194 through herbicides or cultivation). Here we use the emergence calendar for each species according

195 to its **emergence periodicity** and assume  $r_m$  is the proportion of seeds emerging between the 1st  
196 of January (September) and the date the crop is sown in the Spring (Autumn). We assume that  
197 15% (Benvenuti *et al.*, 2001), of the seeds in the bottom layer that are above the maximum depth  
198 for emergence lethally germinate ( $l_g$ ):

$$l_g = 0.15S_B \left( \frac{D - 2}{8} \right). \quad (7)$$

199 We also account for the fact that a certain proportion of seeds  $1 - \Delta$  are lost due to seed-  
200 bank decay. The survival rate of seeds in the seedbank,  $\Delta$ , is associated with the **seedbank type**  
201 functional grouping. Following Thompson *et al* (1997) each species is assigned to one of three  
202 seedbank types: transient, short-term persistent, or long-term persistent. Using data from Lutman  
203 *et al* (2002) on the seedbank survival rates for 20 species (3 transient, 11 short-term persistent, 6  
204 long-term persistent) we calculated the average survival rate for each of the three groups:  $\Delta_{\text{transient}}$   
205  $= 0.3$ ,  $\Delta_{\text{short-term persistent}} = 0.6$ , and  $\Delta_{\text{long-term persistent}} = 0.8$ .

206 Following emergence, a proportion of the seedlings are removed by post-emergence control  
207 methods. There is currently no known association between herbicide efficacy and plant traits and  
208 the response to different herbicides is species specific. To determine the proportion of seedlings of  
209 each species removed under different post-emergence herbicide programs we followed the method  
210 used by Benjamin *et al.* (2009) and categorised post-emergence herbicide control as either low,  
211 moderate, moderately-high or high cost. Expert knowledge was used to estimate the percentage kill  
212 of each weed in each crop, given the costing band of the herbicide programme. Cheap programmes  
213 were assumed to control weeds, which are easy to kill, whereas more expensive programmes are  
214 needed to kill more resilient weeds.

### 215 **Seedlings → Mature plants**

216 **Early growth** In the early part of the growing season, before the total green area index (GAI) of  
217 crop and weeds reaches 0.75, plant growth responds to thermal time ( $T$ ) and we assume there is

218 no competition between individuals. The GAI of a single plant grows according to

$$W_{\text{GAI}}(T(j)) = A \exp(RT(j)) \quad (8)$$

219 where  $A$  is the initial value of the GAI when  $T = 0$ , the  $R$  is the seedling relative growth rate  
220 and  $T(j)$  is the accumulated thermal time from sowing on day  $j$  (see Appendix S1 Box S2). The  
221 total GAI for a single species is obtained by multiplying the GAI of an individual by the number  
222 of seedlings of that species which emerged. This is calculated daily until canopy closure.

223 There is an allometric relationship between seed mass and relative growth rate (Shipley &  
224 Peters 1990) that we employ using the intermediate step of relating seed mass to initial green area.  
225 The initial value of the GAI for a single seedling ( $A$ ) is estimated from the **seed mass** trait by

$$A = \alpha \ln(S_m) + \beta \quad (9)$$

226 where the parameters  $\alpha$  and  $\beta$  vary according to two functional groupings; the **emergence pe-**  
227 **riodicity** and the **phylogeny (grass/broadleaf)**. These parameters were derived for each combi-  
228 nation of these functional groupings using data for 19 species (4 autumn-emerging grasses (AG),  
229 11 autumn-emerging broadleaved weeds (AB), and 4 spring emergers (SE)) from Storkey (2004)  
230 (Appendix S1 Fig. S3).

231 The seedling relative growth rate  $R$  is then estimated from the initial green area ( $A$ ):

$$R = \gamma A + \delta. \quad (10)$$

232 Here the parameters  $\gamma$  and  $\delta$  vary according to the functional groupings of **emergence periodicity**  
233 and the **phylogeny** as well as the season in which the function is called. These parameters were  
234 derived for each combination of these functional groupings using data for 19 species (4 autumn-  
235 emerging grasses (AG), 11-autumn emerging broadleaved weeds (AB), and 4 spring emergers  
236 (SE)) from Storkey (2004) (Appendix S1, Figure S4).

237 In RLM, crop relative growth rate is limited by nitrogen according to a scaling factor ( $N$ ).  
 238 We use this factor to also scale the growth rate of the weed species ( $R$ ). When the **Ellenberg N**  
 239 **number** is greater than or equal to that of the crop, the scaling factor  $N$  takes the same value used  
 240 in the crop model (this is output from RLM). If the weed species is more sensitive to nitrogen than  
 241 the crop (i.e. its **Ellenberg N number** is smaller than that of the crop) then  $N$  is scaled according  
 242 to

$$N(q) = N(p) \frac{B(p)}{B(q)} \quad (11)$$

243 where  $q$  refers to the weed species in question and  $p$  refers to the crop. Here,  $B$  is the reduction in  
 244 plant biomass (under nitrogen limitation) and is also related to **Ellenberg N**:

$$B = 6.5E_N - 14.4 \quad (12)$$

245 We derived this relationship using data (Storkey, 2010) on the difference in biomass for 7 weed  
 246 species grown with and without nitrogen limitation (Appendix S1 Figure S5)

247 **Growth Under Competition** Once canopy closure has been achieved plants will compete for  
 248 light. We used the method described in Kropff and van Laar (1993) to determine the share of  
 249 light for each species, and to calculate growth rates using an estimate of light use efficiency (see  
 250 Appendix S1 Box S3). The share of light ( $s$ ) for a plant of species  $q$  on day  $j$  is calculated  
 251 using information about its own height ( $W_H$ ) as well as the height ( $W_H$ ) and GAI ( $W_{GAI}$ ) of the  
 252 competing species ( $p$  of  $n$  species):

$$s(q) = \exp \left( - \sum_{p=1}^n \left[ \zeta(p) W_{GAI}(p, j) \frac{W_H(p, j) - 0.5W_H(q, j)}{W_H(p, j)} \right] \right) \quad (13)$$

when  $W_H(p, j) - 0.5W_H(q, j) \geq 0$

253 where  $\zeta$  is an extinction coefficient with a value of 0.9 for broadleaves and 0.6 for grasses (Kropff  
 254 and van Laar, 1993).

255 In order to determine  $s$  for each species we need to calculate the plant height. Crop height is  
256 provided by RLM. Weed height is assumed to grow according to

$$W_H(j) = H_I + \frac{H_M}{1 + \exp(-\zeta [P(j) - \tau])} \quad (14)$$

257 where  $P(j)$  is the accumulated photo-thermal time on day  $j$  (see Appendix S1 Box S2).  $H_I$  is  
258 the initial plant height and  $H_I + H_M$  is the **maximum height** for a plant of the given species.  
259 The initial plant height,  $H_I$ , depends on the **phylogenetic grouping** as in an analysis of initial  
260 plant heights ( $H_I$ ) for 16 species (Storkey, 2006) we found significant differences between grasses  
261 and broadleaves with mean values of ( $H_I$  (grasses) = 5.204 (SEM = 2.133),  $H_I$  (broadleaves) =  
262 0.89 (SEM = 0.738)) (Appendix S1, Figure S6). The  $\zeta$  parameter describes the rate of growth;  
263 a common value across species was determined from data for 16 species (Storkey, 2006) to be  
264 0.0106 (SEM = 0.0011).

265 The point of inflection,  $\tau$ , is related to the **day of first flowering** trait ( $W_F$ ):

$$\tau = \lambda W_F + \kappa \quad (15)$$

266 where  $\lambda = 1.354$  (SE = 0.573), and  $\kappa = 501.8$  (SE = 68.8) with a correlation between parameters  
267 of -0.955. These parameters were derived from plant height growth data for 16 species (Storkey,  
268 2006).

269 Once we have calculated the share of light for a given species ( $s_q$ ) and that of the competing  
270 species ( $s_p$ ) we can calculate the proportion of intercepted light that each species ( $q$ ) receives on a  
271 given day ( $j$ )

$$g(q) = \frac{\varsigma_q W_{GAI_q}(j) s_q}{\sum_{p=1}^n \varsigma_p W_{GAI_p}(j) s_p} \left[ 1 - \exp \left( - \sum_{p=1}^n \varsigma_p W_{GAI_p}(j) \right) \right]. \quad (16)$$

272 In the case of the crop, we returned this parameter to RLM to adjust the PAR available for crop  
273 growth. Growth continues in this way until the weed species reaches maturity, the Julian date of

274 which is predicted using the **day of first flowering** trait ( $W_F$ ):

$$W_M = 0.314W_F + 121.8 \quad (17)$$

275 We derived this relationship using data on weed maturation times for 15 weed species (Storkey,  
276 2006). As data were only available for early flowering species we assumed a constant difference of  
277 10 days between flowering and maturity for all later flowering species (flowering after Julian day  
278 163). We would expect this relationship to be sigmoidal rather than linear however due to the lack  
279 of data and the fact that these species will often flower very close to harvest or even after harvest  
280 the additional biomass accumulation between flowering and maturity would be unimportant for  
281 our model.

282 During growth under competition, GAI also accumulates. The increase in GAI of a weed  
283 species from day  $j$  to day  $j + 1$  is given by

$$W_{\text{GAI}}(j + 1) = W_{\text{GAI}}(j) + gI(j) E_m (1 - \rho) \quad (18)$$

284 where  $I$  is the amount of incoming photosynthetically active radiation (PAR, given by RLM),  $E_m$  is  
285 the average light use efficacy ( $\text{m}^2$  dry matter  $\text{MJ}^{-1}$ , see Appendix S1, Box S3) and  $\rho$  is a reflection  
286 coefficient (0.08 based on an average solar elevation of  $45^\circ$ ; Kropff and van Laar, 1993).

### 287 **Mature plants → Fresh seed**

288 **Seed Production** The number of seeds produced ( $S_d$ ) by a given species are related to the plant  
289 biomass at maturity ( $W_{\text{BM}}$ , Lutman *et al.*, 2002). We assume this size dependency of reproductive  
290 allocation remains constant such that the slope of the relationship = 1 (Sugiyama and Bazzaz,  
291 1998)

$$\ln(S_d) = v + \ln(W_{\text{BM}}). \quad (19)$$

292 The species dependent parameter  $v$  is estimated from the **seed mass** trait:

$$v = -0.1177 \ln(S_m)^2 - 0.672 \ln S_m + 5.789. \quad (20)$$

293 We fitted this relationship to data on 14 weed species (Storkey *et al.*, 2015) (Appendix S1 Figure  
294 S7).

295 The weed biomass at maturity ( $W_{\text{BM}}(j)$ ) is related to the GAI on the day of maturation  
296 ( $W_{\text{GAI}}(j)$ ) and the **specific leaf area** trait ( $W_{\text{SLA}}(j)$ ):

$$W_{\text{BM}}(j) = \varepsilon \frac{W_{\text{GAI}}(j)}{W_{\text{SLA}}(j)} \quad (21)$$

297 where  $\varepsilon = 6.121$ ,  $\text{SE} = 0.363$  and relates the leaf biomass (GAI/SLA) to total plant biomass on day  
298  $j$ . We used data on measured green area and dry weights from Storkey (2006) to determine this  
299 relationship (Appendix S1, Figure S8).

### 300 **Fresh seed → Seedbank**

301 **Seed Losses** If the weed has not reached maturity on the Julian day when the crop is "harvested"  
302 in RLM then no seed is shed. A maximum of 100% seed shed is reached 38 days after matu-  
303 rity (mean of observed data from the UK for *Avena* spp (Barosso *et al.*, 2006) and *Alopecurus*  
304 *myosuroides* (R Hull unpublished data) and estimates for *Galium aparine* (Lutman, 2002) and we  
305 assume the response is linear. Any unshed seed is lost and not subsequently added to the seedbank.

306 Following a meta-analysis of post-harvest seed losses by Davis (2011), seed predation is ran-  
307 domly sampled from a normal distribution with mean 0.52 and standard deviation 0.05. This  
308 portion of the seed shed is not subsequently added to the seedbank.

309 **Vertical Movement of Seed in the Soil** Seeds are moved vertically between the shallow and  
310 deep soil layers following data described by Moss (1990). In years when the cultivation type is  
311 "plough" a proportion of seeds from the shallow soil layer are buried into the deep soil layer drawn

312 from a log-normal distribution with mean =  $-0.0515$  and standard deviation =  $0.0191$ , conversely  
313 some seeds are brought up to the shallow soil layer — this proportion is drawn from a log-normal  
314 distribution with mean =  $-1.0570$  and standard deviation =  $0.1199$ . For all other cultivation types  
315 there is no upward movement of seed (from the deep soil layer to the shallow soil layer). For “min  
316 till” data on cultivations at 10 cm were used to give the proportion of seeds that are buried taken  
317 from the distribution  $\mathcal{N}(0.2, 0.051)$ . In years where “direct drill” is chosen (data from  $<5$  cm tine)  
318 no seeds move vertically.

319 If the seedbank for a species (in either the top or bottom soil layer) falls below 1 seed ( $\text{m}^{-2}$ )  
320 then that species is assumed to have gone extinct locally and is not included in subsequent years  
321 simulation.

## 322 **Model Testing**

323 To evaluate the performance of our trait-based community model we compared the community  
324 predicted by our model with the observed weed community (see Appendix S1 Box S4 for methods  
325 of data collection) in an arable field (Brome Pin, Brooms Barn, Suffolk, UK), for which the weather  
326 (e-RA, 2018), crops, tillage, and fertiliser input history was available for 30 years (1987–2016).  
327 We initialised our model with 100 weed seeds in each soil layer of each species in the regional pool  
328 (101 annual arable weeds - see Appendix S1 Box S5). We simulated the 30 years prior to seedbank  
329 collection (1987–2016) using the known management information for those years. As we did not  
330 know the level of herbicide input used in the field we ran the model 20 times for each level of  
331 herbicide input (none, low, medium, high, and very high) to determine whether this significantly  
332 altered the number of plants, seeds in the top layer of the seedbank, or seeds per plant in the final  
333 simulated community (One-Way ANOVA).

334 We calculated the functional diversity (*sensu* Petchey and Gaston, 2002) of each simulated  
335 community at the end of the 30 year simulation in R 3.5.0 (R Core Team, 2018). We first standard-  
336 ised the traits data and computed a dissimilarity matrix using the *vegan* package (Oksanen *et al.*,  
337 2019). We then used hierarchical clustering to create a dendrogram of the relations between species

338 and computed the functional diversity (total branch length) using the *picante* package (Kembel  
339 *et al.*, 2010). We tested to see if the selected communities were functionally different under the  
340 different herbicide regimes (One-Way ANOVA) and also whether the functional diversity of the  
341 selected communities differed significantly from the regional species pool (One-Way ANOVA).

342 For each model realisation we also compared the resulting density distribution of each trait  
343 in the simulated community with the initial trait distribution of the regional pool and that of the  
344 observed weed community in Brome Pin.

## 345 **Results**

346 The weed community in Brome Pin comprised 23 species. The two most abundant species were  
347 volunteer crops of oats and oilseed rape. Of the remaining 21 weed species, 6 were perennials  
348 and 15 were annuals (Appendix S2 Table S1). In our subsequent analyses we only considered the  
349 community of 15 annuals to align with the scope of our model and excluded crop volunteers as  
350 their population dynamics are driven by repeated reintroduction.

351 In our simulations, the abundance of plants varied significantly at different levels of herbicide  
352 ( $P < 0.001$ , One-way ANOVA). Plant abundance was highest when herbicide input was low and  
353 decreased with increasing herbicide input (Fig. 3a). The number of seeds in the top layer of  
354 the seedbank followed a different pattern. Following 30 years of simulation with no herbicide  
355 there were few seeds in the seedbank, yet significantly higher seed numbers were simulated at  
356 all levels of herbicide application ( $P < 0.001$ , One-way ANOVA). Seed abundance also increased  
357 with increasing herbicide input (Fig. 3b). This had the interesting effect that the number of seeds  
358 per plant was significantly altered under different herbicide regimes ( $P < 0.001$ , One-way ANOVA)  
359 with an exponential-like increase in seed production at increasing levels of herbicide input (Fig.  
360 3c). We suggest this is a result of communities being dominated by species with high fecundity,  
361 allowing them to buffer the effects of herbicide, and a reduction in competition between weed  
362 individuals.

363 The community of weed species selected for by the model was fairly consistent across simula-

364 tions. In the majority of our simulations *Sonchus asper* was the most abundant species but across  
365 all simulations there were only nine different species which were ever predicted to be the most  
366 abundant ( $P < 0.001$  compared to random selection of species; Appendix S2 Table S2). The species  
367 predicted to be the most abundant remained fairly consistent across higher levels of herbicide in-  
368 put, yet as herbicide input was reduced we saw a greater variety in the most abundant species  
369 predicted by the simulations (Appendix S2 Table S2). Our model very rarely predicted the local  
370 extinction of species, however, the abundances of most species remained very low. The species  
371 which did maintain high abundance were often similar across simulations, with eight species con-  
372 sistent ranking among the 20 most abundant species (across all 20 simulations for each herbicide  
373 scenario, see Appendix S2 Table S3). *Atriplex patula*, *Conyza cadensis*, *Fumaria officinalis* and  
374 *Veronica persica* were often found amongst the 20 most abundant species when no herbicide was  
375 applied but markedly less so at any level of herbicide application. Our model had mixed success  
376 at predicting the species found in Brome Pin with only 8 of the 15 annuals observed in Brome  
377 Pin ranking among the 20 most abundant species in any simulation. However, as it is not possible  
378 to separate out environmental filtering from founder effects, and these species were only found in  
379 small numbers in Brome Pin it could be that these species would not always be abundant at this  
380 field site given the environmental and management conditions. The model was more successful at  
381 predicting the emergent distribution of functional traits.

382 There was a significant difference in functional diversity of the resulting simulated communi-  
383 ties compared to the regional pool (One-way ANOVA,  $P < 0.001$ ), indicating that there has been  
384 directional selection of functional traits. However, the functional diversity of the simulated com-  
385 munities were not significantly different under different herbicide regimes (One-Way ANOVA,  
386  $P < 0.05$ ) indicating that herbicide input is not a key driver of functional diversity in our model, so  
387 we only show the trait distributions for the medium herbicide level here.

388 For the continuous traits included in our model the distribution of traits observed in Brome Pin  
389 (yellow distributions in Fig. 4) was a subset of the full regional pool (blue distributions in Fig.  
390 4). In our model simulations (black distributions in Fig. 4) we saw different levels of selection for

391 the various traits. In our simulations there was a strong selection according to seed mass (Fig. 4a)  
392 with the simulated communities all showing similar seedmass trait distributions to that observed in  
393 Brome Pin, whereas for maximum height the trait distribution of the simulated communities was  
394 not very dissimilar to the regional pool indicating that there is not a strong selection for maximum  
395 height in our model (Fig. 4b). The distribution of flowering times (Fig. 4c) observed in our  
396 simulations centres on later flowering species than we observed at Brome Pin, however the latest  
397 flowering species from our species pool, (first day of flowering in August, Julian day  $\geq 213$ ), are  
398 excluded following our simulations and so there is some limited evidence for directional selection  
399 based on flowering times in our model. There is little evidence for selection based on SLA in our  
400 model (Fig. 4d). However, this lack of selection based on SLA is also reflected in the observed  
401 community in Brome Pin.

402 For the discrete functional groups used in our model there was also a distinction between the  
403 composition of the regional pool (blue bars in Fig. 5) and the community in Brome Pin (yellow bars  
404 in Fig. 5). Again, the model simulations (black lines in Fig. 5) showed varying levels of selection  
405 for the different factors. The observed species in Brome Pin all had Ellenberg N values between 6  
406 and 8 with most individuals having an Ellenberg N of 6. The regional pool instead shows a peak at  
407 Ellenberg N =7. Many of our simulated communities show a broad spectrum of Ellenberg N values  
408 taken from the full range present within the regional pool, however, in some simulations there is  
409 selection towards a peak at Ellenberg N = 6, although this is not consistent. In the regional pool  
410 there are a similar number of species with each type of emergence calendar (Fig. 5b). However, in  
411 Brome Pin we found very few Autumn-emergers and most individuals were generalist emergers.  
412 Our simulated weed communities reflected this with strong selection against autumn-emerging  
413 species. Whilst most species in our regional pool are broadleaves with fewer grasses (Fig. 5c)  
414 there is an even stronger bias toward broadleaves in the weed community observed in Brome  
415 Pin, with very few grasses found in the sampled seedbank. Our model simulations reflected this  
416 selection pressure and in all simulations the frequency of grasses was reduced compared to the  
417 regional pool. In Brome Pin we saw a prevalence of short-term persistent seedbank types and an

418 absence of species with a transient seedbank. Our model selects for both short-term and long-term  
419 persistent seedbank species but we also saw a removal of species with a transient seedbank in line  
420 with our observations from Brome Pin.

## 421 **Discussion**

422 Predicting the relative abundance of species along environmental gradients or following changes  
423 in management practices is a fundamental goal in community ecology. Our approach, which links  
424 trait-based environmental filtering with a process-based community model, allows both the di-  
425 vergent and convergent selection pressures of environmental filters and biotic interactions to be  
426 considered in combination. The observed data on functional traits from the study field generally  
427 reflected a convergence of traits, especially for seed mass and this was captured by the model. How-  
428 ever two distinct peaks were observed in the density plots of observed data for maximum height  
429 and specific leaf area reflecting a divergence of traits in response to crop competition. The sim-  
430 ulation output for maximum height also had two peaks, although underestimating the dominance  
431 of shorter species. As the functions modelling competition incorporate height, it is encouraging  
432 that biological interactions result in a degree of trait divergence. However, the effect of variation  
433 in SLA on competition for light is not currently included in the model and the results indicate that  
434 further development is required to reflect the observed divergence in this trait.

435 We demonstrated that by parameterising a process-based model using data from well-studied  
436 plant traits that we can effectively model the effect of environmental filters on plant communities  
437 at the level of functional traits. In all of our simulations the direction of selection was consistent  
438 with in-field observations. Although there was stronger selection for some traits than for others.  
439 We predicted different plant communities under different levels of herbicide indicating that this  
440 simple management filter does exert selection pressure at the trait level. We also demonstrated  
441 that stochasticity can play a role in community assembly as the inclusion of stochastic processes  
442 in our model resulted in different realisations of the final plant community, although the functional  
443 diversity of those communities remained similar.

444 By combining the trait filtering approach with a process-based community model we revealed  
445 a number of emergent properties of the model which were not anticipated by the inputs alone.  
446 Our model predictions under varying levels of herbicide input predict the largest number of plants  
447 when herbicide input is low. This phenomenon has been observed in the field for a number of  
448 weed species (e.g. Buhler, 1999, Boström & Fogelfors, 2002) and is consistent with the interme-  
449 diate disturbance hypothesis which states that at intermediate levels of disturbance (low herbicide)  
450 coexistence is more likely (Catford *et al.*, 2012). This unanticipated emergent property highlights  
451 the importance of including mechanistic processes in the model in addition to empirical relation-  
452 ships between traits and environmental filters as the synergistic effect of these processes may reveal  
453 interesting aspects of community dynamics such as these which can only be revealed when there  
454 is both convergent and divergent selection acting simultaneously.

455 Our model takes mean trait values as input for each species, yet within a species the value for  
456 that trait may vary along environmental gradients or change through time (Violle *et al.*, 2007). It is  
457 important that we recognise this intraspecific variation in models of this kind. For example, plant  
458 height is very dynamic, and depends strongly on disturbance regime (Garnier & Navas, 2012)  
459 meaning that the mean values reported in the literature and used here in our model may not be  
460 very accurate for plants of the same species growing in a highly disturbed arable field. A similar  
461 argument can be made for flowering time. However, as yet, these data are not readily available  
462 for arable systems in the UK and as such this may be a source of error in our model. Despite  
463 not explicitly incorporating intraspecific trait variation in our model we do include it implicitly  
464 by accounting for the uncertainty in each trait–parameter relationship and so by including this  
465 stochasticity within the model we account, to some extent, for variation between individuals.

466 The discrepancies between the ability of our model to successfully predict the correct species  
467 list for our studied field (limited success) and the ability to predict the correct distribution of func-  
468 tional traits (greater success) highlights an important question surrounding its utility in predicting  
469 community composition. If, as we state in the introduction to this paper, the primary objective of  
470 community ecology is to predict the impact of change (environment, land-use, and management)

471 on the *function* of the emergent community then our model succeeds. This will be particularly  
472 pertinent where there are associations between the response traits included within the model and  
473 any effect traits that determine ecosystem function (Díaz *et al.*, 2007a). However, if the objective  
474 is to simply predict the composition of species then our model is of more limited use.

475 By demonstrating the ability of our model to predict changes in both weed abundance, and the  
476 distribution of functional traits we have shown that it will have utility in assessing the viability  
477 of various management scenarios. For example, if the aim of weed management is to reduce  
478 overall weed abundance we could use our model to assess the success of a number of hypothetical  
479 management regimes in achieving this for a given field. Similarly, if the aim of weed management  
480 is to provide a functionally diverse weed community which can support the provision of ecosystem  
481 services then this too could be assessed through simulation of various management options to  
482 determine the best approach for achieving this.

483 Whilst our model is intrinsically linked to the arable production system, the principle of com-  
484 bining trait-based filtering with process-based models could be easily extended to any ecosystem  
485 where the community composition of annual plants is of interest. The generic model of the annual  
486 plant life-cycle is broadly applicable and questions surrounding changes in management or envi-  
487 ronment can be easily addressed as demonstrated by our inclusion of different herbicide programs.  
488 For example, the effect of post-emergence herbicide on seedling mortality included in our model  
489 could be easily mapped to other management practices such as grazing or even natural disturbances  
490 such as burning — provided details are known about the proportion of the population removed by  
491 such disturbances. The main factor limiting the application of our modelling framework to habitats  
492 other than cultivated fields is the level of specificity of the relationships between functional traits  
493 and model parameters that have been quantified for a subset of arable weeds. There is evidence in  
494 the literature that some of these allometric relationships follow ecological rules and are conserved  
495 across functional groups (for example seed weight and seedling growth rate (Shipley & Peters,  
496 1990). However, the extent to which the model can be applied to other annual plant communities  
497 without additional experimental parameterisation remains to be determined.

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Table 1: Each transition from one stage of the annual plant life cycle to the next is the result of one or more mechanistic processes. The success of each of these processes is determined by the strength of certain environmental filters which act on various response traits.

Transition	Process	Filters	Response Traits
Seedbank → Seedlings	Seedling emergence	Date of crop sowing Depth of cultivation	Emergence periodicity* Seed mass
	Seedling mortality Seedbank Decline	Herbicide timing Frequency and depth of cultivation	Emergence periodicity* Seedbank persistence*
Seedlings → Mature plants	Early Growth	Temperature	Seed mass, emergence periodicity*, grass / broad-leaf*
	Competition	Fertilization Crop canopy architecture Fertilisation	Ellenberg N number* Maximum height, flowering time Ellenberg N number*
Mature plants → Fresh seed	Seedbank decline	Timing of harvest	Flowering time, SLA, Seed mass

## Figure Captions

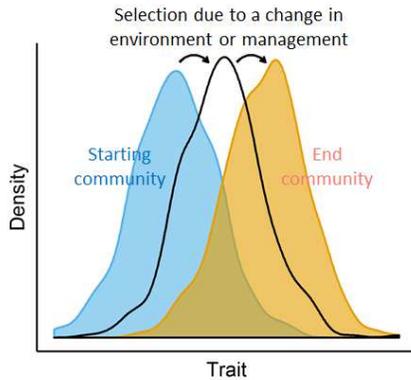
Fig.1: Combining trait filtering and community dynamics modelling approaches allows us to predict changes in community composition. We use relationships between functional traits published in trait databases and parameters in the annual plant life cycle to parameterise a mechanistic model for multiple species.

Fig.2: Emergence calendars for Spring emergers, autumn emergers and generalist emergers. Bimodal normal probability distributions are fitted to each group using data from Storkey *et al.* (2015), (5 spring emergers, 2 autumn emergers, 3 generalist emergers).

Fig. 3: Summary at all levels of herbicide input of the total a) plants, b) seeds in the top layer of the seedbank, and c) seeds per plant in the end community after 30 years of simulation. Bar height represent the means from 20 simulations at each level of herbicide input and error bars show the standard error of the mean.

Fig. 4: Density plots showing the frequency of the continuous traits a) seed mass, b) maximum height, c) flowering day, and d) specific leaf area. The green distribution shows the density function fitted to the observed data from the Brooms Barn field site, the blue distribution shows the full range of trait data included in the model and represents a density function fitted to an even community consisting of all species, the purple lines are the density functions fitted to each realisation of the field following the simulation of 30 years of management history at the brooms barn site.

Fig. 5: Density plots showing the frequency of the discrete grouping factors a) Ellenberg N, b) emergence group, c) phylogeny, and d) seedbank longevity. The green bars show the density of the observed data from the Brooms Barn field site, the blue bars show the full range of trait data included in the model and represents the density of an even community consisting of all species, the purple lines are the density values of the group for each realisation of the field following the simulation of 30 years of management history at the brooms barn site.



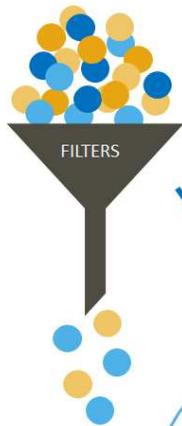
A. Predicting community composition and the effects of environmental change on functional diversity is a key goal of community ecology.

There are two main approaches to this.

**B. Trait-based filtering approach**

Species possessing traits within certain thresholds determined by filters are selected.

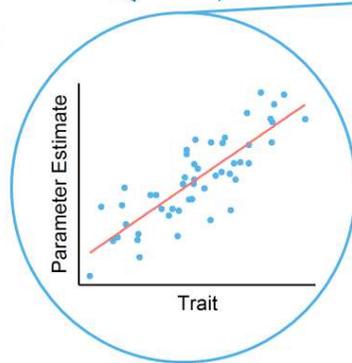
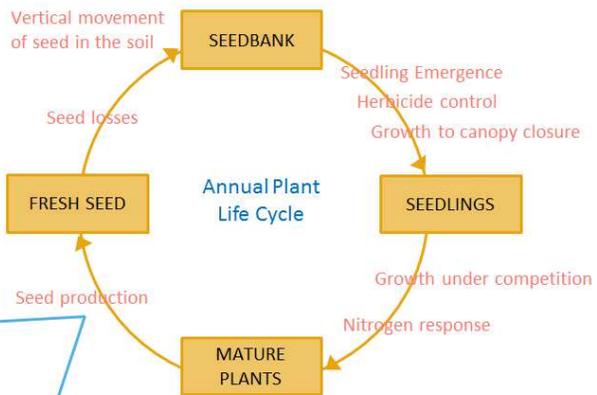
This leads to a *convergence* in traits



**C. Community dynamics modelling approach**

The life cycle of individuals is described mechanistically and species compete with one another for resources.

This leads to *divergence* in traits



Combining these two approaches allows us to predict changes in community composition

D. We can parameterise each mechanistic process within a community dynamics models for multiple species by describing trade-offs and correlations among plant traits.

This reduces the parameterisation requirements of the community dynamics modelling approach and resolves the issue of functional convergence/divergence.

