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Research



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An overlooked mechanism underlying the attenuated temperature response of soil heterotrophic respiration

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Biogeochemical reactions occurring in soil pore space underpin gaseous emissions measured at macroscopic scales but are difficult to quantify due to their complexity and heterogeneity. We develop a volumetric-average method to calculate aerobic respiration rates analytically from soil with microscopic soil structure represented explicitly. Soil water content in the model is the result of the volumetric-average of the microscopic processes, and it is nonlinearly coupled with temperature and other factors. Since many biogeochemical reactions are driven by oxygen (O₂) which must overcome various resistances before reaching reactive microsites from the atmosphere, the volumetric-average results in negative feedback between temperature and soil respiration, with the magnitude of the feedback increasing with soil water content and substrate quality. Comparisons with various experiments show the model reproduces the variation of carbon dioxide emission from soils under different water content and temperature gradients, indicating that it captures the key microscopic processes underpinning soil respiration. We show that alongside thermal microbial adaptation, substrate heterogeneity and microbial turnover and carbon use efficiency, O₂ dissolution and diffusion in water associated with soil pore space is another key explanation for the attenuated temperature response of soil respiration and should be considered in developing soil organic carbon models.

1. Introduction

Soil contains more organic carbon (1580 billion t C) than plant biomass (610 billion t C) and atmospheric carbon dioxide (CO₂) (750 billion t C) combined. As a consequence, small shifts in soil organic carbon (SOC) could have significant consequences for global warming [1]. An understanding of the mechanisms underlying SOC dynamics and representing them in SOC models adequately is crucial to predicting the outcomes of climate warming and land management practices upon SOC stocks [2]. However, a thorough understanding is challenging due to the complexities of biogeochemical reactions involved in SOC cycling [3].

Of all biotic and abiotic factors, temperature and water have the greatest influence upon biogeochemical processes in soil [4]. Although their roles in homogeneous media are fairly well understood, controversy arises when applying this understanding to soils because of their complexity and heterogeneity [5].

Microbial activity in soil is patchy and biogeochemical reactions proceed only where substrates and enzymes exist [6]. Soil variables measured from sampling without information on substrate accessibility can thus give rise to erroneous conclusions when used to calculate biogeochemical reactions [7]. One example is the observed reduced temperature response of aerobic microbial respiration, where the underlying mechanisms have been a point of contention for decades [8–11]. Changes in microbial physiology with temperature and substrate heterogeneity have both been postulated, but there is little consensus about their relative significance [3].

Previous studies on the temperature response of soil respiration have focused on biological processes; overlooking the fact that biogeochemical reactions are driven by bioavailability of oxygen (O_2) and substrate accessibility within the pore space. Resistance against O_2 dissolution in soil water and its subsequent movement limits its delivery to microbes, evident as the pervasiveness of anaerobicity in relatively dry soils [12]. While soil water was thought to keep microbes hydrated and carry dissolved substrates and enzymes away and towards microsites [13], experiments suggest that reduced substrate availability due to soil water decrease is far more important than dehydration [14]. Within soil pore space, gaseous O_2 first dissolves at the water–air interface before diffusing to aerobic microbes (reactive sites). Respiration generates localized O_2 concentration gradients between the water–air interface and reactive sites. These gradients vary with temperature and soil water content, as rising temperature reduces O_2 dissolution while a change in soil water content reshapes the water–air interface and the distance between the sites of O_2 dissolution and microsites. As a result, the influences of temperature and soil water content on microbial activity have been postulated to be coupled: a change in one factor is likely to alter the response of soil respiration to the other. However, most SOC models decouple the effects of temperature and soil water content using separate moisture and temperature functions to describe their respective influence [15,16]. There are no systematic studies of the potential errors associated with this approach; indirect evidence indicates that the temperature coefficient, Q_{10} (or activation energy, E_a) which characterizes the temperature sensitivity of soil respiration, varies with soil moisture [17]. When predicting the response of soil respiration to global warming, a small change in Q_{10} can lead to substantial differences [18].

The temperature function used in most SOC models is Q_{10} or the Arrhenius kinetic model [4]. By contrast, the moisture functions employed are diverse, including both empirical formulae and mechanistic models [15]. While the empirical functions are phenomenological, most mechanistic models are based on the influence of soil water on the diffusion of gaseous O_2 and aqueous substrates. This approach overlooks the fact that biogeochemical reactions alter local O_2 concentration gradients and hence the diffusion of dissolved O_2 . For soil, these models predict a fixed optimal moisture content where respiration is maximized. This is inconsistent with experimental results which show that the optimal soil water content for soil respiration varies with temperature [19–22], microbial activity [23] and even soil depth [24]. This implies that the effect of moisture on respiration is modulated by temperature [25], and that their combined influence is more complicated than described by the separated moisture and temperature functions used in current models [3].

As well as mediating O_2 dissolution and diffusion, soil water content also controls microbial access to substrates [13,26,27]. The consensus view arising from decades of study is that increasing soil water content facilitates substrate movement, increasing microbial access [4,28]. Some models account for this by introducing a moisture-dependent barrier between substrates and reactive sites [22,29]. While this is rational for substrates and enzymes, it does not apply to O_2 because O_2 must dissolve at water–air interface before becoming bioavailable for respiring microbes. Increasing water content of a dried soil initially increases O_2 dissolution, but once a threshold is reached, increasing soil water content further reduces O_2 supply because of the reduced area of water–air interface across which O_2 must dissolve, and the increased distance for dissolved O_2 to travel to reactive sites [30]. Dissolution of O_2 and its diffusion control biogeochemical reactions in soil [31], but they are difficult to model due to their complexity [32]. Consequently, most SOC models do not consider O_2 explicitly, probably based on an erroneous perception that O_2 in the topsoil is not a limiting factor [33]. Decades of studies have demonstrated anaerobic reactions persist even when soil is relatively dry, especially in the rhizosphere [31,34,35].

Given the importance of soil architecture in modulating water distribution and O_2 dissolution and diffusion, as well as the influence that biogeochemical reactions impose on local O_2 concentration gradients between water–air interface and reactive sites, we hypothesized that the reduced O_2 dissolution and increased microbial metabolism at raised temperatures attenuate the temperature response of soil respiration. We developed a volumetric-average method to incorporate microscale soil architecture to calculate soil respiration. Considering that soil architecture and its associated microscopic processes are complicated, to make the calculation analytically tractable, we adopted several rational simplifications. These include (i) gaseous O_2 concentration in a soil sample is uniform as O_2 diffuses four orders of magnitude faster in air than in liquid water [36]; (ii) biogeochemical reactions in a soil sample are in a quasi-steady state where the amount of O_2 dissolved at water–air interfaces is the same as the amount of O_2 reduced by aerobically respiring microbes, and that the mass of O_2 respired is the same as the mass of O_2 diffusing from water–air interfaces to all reactive sites; (iii) O_2 reduction by aerobic microbial activity at hydrated reactive sites in a soil sample is proportional to the dissolved O_2 concentration at reactive sites; (iv) the majority of soil microbes adopt a ‘waiting’ strategy to acquire substrates and O_2 [37]. Volumetrically averaging microscopic processes over the hydrated pore space in a soil sample yields an analytical model to calculate respiration; soil water content in the model is the result of the volumetric-average and nonlinearly coupled with temperature and other factors. Such coupling has been conjectured since the 1970s [38,39]: here we demonstrate its existence and use theory to infer that a change in one factor reshapes the response of soil respiration to the other.

2. Material and methods

2.1. Theoretical analysis

Figure 1 depicts the microscopic processes considered in the model. Gaseous O_2 first dissolves at the water–air interface before moving in water as dissolved O_2 ; soil organic matter

associated with the matrix is decomposed enzymatically by exoenzymes. At hydrated sites with the coexistence of substrates, aerobic microbes take up dissolved O₂ and substrates. The movement of dissolved O₂ and substrates are confined to the regions enclosed by the water–air interface and wetted pore walls. It is described by the following equations, with dissolved organic carbon and O₂ as the limiting substrates [4,40]:

$$\left. \begin{aligned} \frac{\partial c_O}{\partial t} &= \nabla D \nabla c_O - v_{\max} N \frac{c_D}{k_D + c_D} \frac{c_O}{k_O + c_O} \\ \text{and } \frac{\partial c_D}{\partial t} &= \nabla D_D \nabla c_D - (1 - \beta) v_{\max} N \frac{c_D}{k_D + c_D} \frac{c_O}{k_O + c_O} + s_{DOC} \end{aligned} \right\} \quad (2.1)$$

Variable nomenclature is given in appendix A. O₂ enters the system via dissolution at the water–air interface, which is described by a first-order kinetic process [41]

$$d_o = \alpha(C_{\text{eq}} - c_0), \quad (2.2)$$

where d_o is O₂ dissolution rate over a unit water–air interfacial area, α is the dissolution rate coefficient, c_0 is the dissolved O₂ concentration at the water–air interface and C_{eq} is the saturated dissolved O₂ concentration, calculated from the Henry's law [42].

Compared with the timeframe over which soil water content changes, pore-scale processes are rapid and at a quasi-steady state over daily temporal scale. The aim of this paper is not to solve equation (2.1), but to develop a method to volumetrically average it throughout a soil sample to calculate the total reaction rate. To make the calculation analytically tractable while maintaining the key processes illustrated in figure 1, we made some rational simplifications as explained above. For a soil water content of θ , the total area of wetted pore walls in a soil is represented by $A_{\text{ws}}(\theta)$, and the spatial variation of the wetted pore wall in the Cartesian coordinate system is described by function $s(x, y, z)$. If the number of aerobic microbes on the wetted wall located at (x, y, z) is $n(s)$, the total aerobic microbes on hydrated microsites in the soil is $\int_{A_{\text{ws}}} n(s) \cdot ds$, which reduces to $A_{\text{ws}}(\theta) \cdot n_0$ if the microbes are uniformly distributed in that $n(s) = n_0$. Similarly, the total water–air interfacial area is represented by $A_{\text{wa}}(\theta)$, and the spatial variation of the water–air interface in the Cartesian coordinate system is described by function $s'(x, y, z)$. If the dissolved O₂ concentration at the water–air interface located at (x, y, z) is $c_0(s')$, the O₂ dissolution rate in the soil is $\int_{A_{\text{wa}}} \alpha[C_{\text{eq}} - c_0(s')] \cdot ds'$. The Michaelis–Menten constant associated with O₂ in equation (2.1) regulates microbial growth, varying with microbial species and substrate quality [43,44]. Since we consider whole microbial communities, it is approximated by $Kc_0/(k_0 + c_0) \approx \kappa c_0$ [33].

At steady state, the rate of microbial reduction of O₂ in a soil sample is balanced by the O₂ dissolution rate at water–air interfaces. When soil water content is θ , we have

$$\int_{A_{\text{wa}}} \alpha[C_{\text{eq}} - c_0(s')] ds' = \int_{A_{\text{ws}}} n(s)(1 - \beta)v_{\max} \kappa c_0 \frac{c_D}{k_D + c_D} ds. \quad (2.3)$$

The dissolved O₂ concentration at the water–air interface in a soil sample is approximately constant as the gaseous O₂ concentration is constant. By contrast, substrate concentrations and the number of aerobically respiring microbes vary over the wetted pore wall. Equation (2.3) is hence calculated by

$$\alpha A_{\text{wa}}(C_{\text{eq}} - c_0) = n'(1 - \beta)v_{\max} \frac{c_D}{k_D + c_D} \kappa \int_{A_{\text{ws}}} c_0 ds, \quad (2.4)$$

where C_D and n' are the average substrate concentration and the average number of aerobic microbes over a unit area of wetted pore walls in the soil, respectively, when soil water content is θ . The diffusion of O₂ in water is slow and its concentration over the wetted pore walls, c_0 , varies. The integral in equation (2.4)

is approximated by

$$\int_{A_{\text{ws}}} c_0 ds = A_{\text{ws}} C_0, \quad (2.5)$$

where C_0 is the average dissolved O₂ concentration over the wetted pore wall. We thus have

$$\alpha A_{\text{wa}}(C_{\text{eq}} - c_0) = n'(1 - \beta)v_{\max} \kappa \frac{c_D}{k_D + c_D} A_{\text{ws}} C_0, \quad (2.6)$$

Diffusion of O₂ from the water–air interface to reactive sites depends on their spatial separation and local concentration gradients. If the average hydraulic distance between air–water interfaces and wetted pore walls is L , and the average cross-sectional area in soil water for O₂ to diffuse from water–air interface to the microsites is A_D , the overall diffusive flux is

$$Q = D \frac{c_0 - C_0}{L} A_D. \quad (2.7)$$

Approximating A_D by the mean of the water–air interfacial areas and wetted pore wall areas, i.e. $A_D = 0.5(A_{\text{ws}} + A_{\text{wa}})$, at steady state, the mass balance requires that the diffusive flux Q equates the total O₂ respiration rate, i.e.

$$D \frac{c_0 - C_0}{L} \frac{A_{\text{ws}} + A_{\text{wa}}}{2} = n'(1 - \beta) v_{\max} \kappa A_{\text{ws}} \frac{c_D}{k_D + c_D} C_0. \quad (2.8)$$

Solving for c_0 yields:

$$\left. \begin{aligned} c_0 &= \left(1 + k \frac{2L \cdot A_{\text{ws}}}{D(A_{\text{ws}} + A_{\text{wa}})} \right) C_0 \\ \text{and } k &= n'(1 - \beta)v_{\max} \kappa \frac{c_D}{k_D + c_D} \end{aligned} \right\} \quad (2.9)$$

Substituting equation (2.9) into equation (2.6) gives

$$C_0 = \frac{\alpha A_{\text{wa}}}{\alpha A_{\text{wa}} [1 + 2L \cdot (k/D) \cdot A_{\text{ws}}/(A_{\text{ws}} + A_{\text{wa}})] + k A_{\text{ws}}} C_{\text{eq}}. \quad (2.10)$$

The total respiration rate from the soil can be calculated as follows:

$$\left. \begin{aligned} \Omega &= k \cdot A_{\text{ws}} \cdot C_{\text{eq}} \cdot E_r \\ \text{and } E_r &= \left[1 + \frac{k}{\alpha} \left(\frac{2\alpha}{D} \frac{L \cdot A_{\text{ws}}}{(A_{\text{ws}} + A_{\text{wa}})} + \frac{A_{\text{ws}}}{A_{\text{wa}}} \right) \right]^{-1} \end{aligned} \right\}, \quad (2.11)$$

where $E_r < 1$ is a feedback factor emerging from the volumetric-average, describing the reduction in respiration due to O₂ dissolution and diffusion in soil water. Physically, k represents the potential demand of aerobic microbes over a unit area of wetted pore wall for O₂ when O₂ is not a limiting factor. It depends on temperature, substrate quality/quantity, the number of microbes and their metabolic rates. The hydraulic distance between the water–air interface and the microsites increases with soil water content, approximated by [41]

$$L = \lambda \frac{A_{\text{ws}}}{A_{\text{wa}}}, \quad (2.12)$$

where λ is constant depending upon soil architecture.

The effect of O₂ dissolution and diffusion on soil respiration is described by the O₂ dissolution rate coefficient and the molecular diffusion coefficient of dissolved O₂, respectively. When O₂ dissolution is significantly faster than microbial respiration, i.e. $k/\alpha \ll 1$, the feedback factor reduces to

$$E_r = \left[1 + \frac{2k\lambda}{D} \frac{A_{\text{ws}}}{(A_{\text{ws}} + A_{\text{wa}})} \frac{A_{\text{ws}}}{A_{\text{wa}}} \right]^{-1}, \quad (2.13)$$

with O₂ diffusion being the limiting factor. When dissolved O₂ diffusion is significantly faster than microbial respiration,

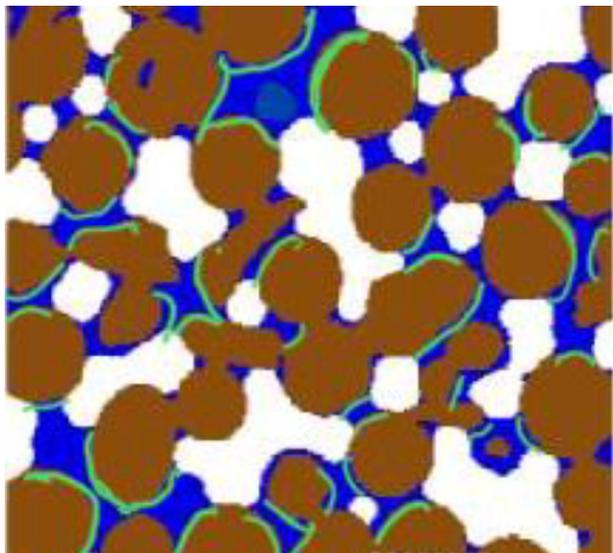


Figure 1. Schematic of microscopic processes controlling aerobic microbial respiration at the pore scale in soil. Brown represents the soil matrix, white regions represent air and blue represents water. Gaseous O_2 (red) dissolves at the water–air interface; dissolved O_2 (yellow) moves to aerobic microbes (green) adjacent to wetted pore walls where O_2 is respired.

i.e. $kL/D \ll 1$, the feedback factor reduces to

$$E_r = \left[1 + \frac{k A_{ws}}{\alpha A_{wa}} \right]^{-1}, \quad (2.14)$$

with O_2 dissolution being the limiting factor.

The influence of pore geometry and water distribution within the pore space is described by a combination of the water–air interfacial area, the wetted pore wall area and the hydraulic distance separating them. Temperature influences soil respiration in two ways. Physiologically, rising temperature increases microbial metabolic rate, described by the Arrhenius kinetic equation [5],

$$v_{\max} = v \cdot \exp \left[-\frac{E_a}{R(T+273)} \right]. \quad (2.15)$$

Physically, increasing temperature reduces O_2 dissolution but enhances its diffusion. At one atmospheric pressure, changes in saturated concentrations of dissolved O_2 and its molecular diffusion coefficient with temperature are described by [36],

$$\begin{aligned} C_{\text{eq}} &= 0.434 \exp(0.000064T^2 - 0.0114T + 1.161), \\ D &= 0.434 \exp[-4.41 + 773.8/(T+273) - 2564.4/(T+273)^2]. \end{aligned} \quad (2.16)$$

2.2. Water–air interfaces and wetted pore walls

As soil water content increases, the wetted pore wall area increases monotonically. However, the water–air interfacial area first increases and then declines. To investigate if such changes can be described by general formulae, we simulated water distribution in over 100 soil samples as described in our previous work [45,46]. For all soil samples, the changes in water–air interfacial area and the wetted pore wall area with soil saturation follow similar trends (electronic supplementary material, figure S1). Normalized by the volume of the soil sample, their changes with soil saturation, Θ , can be described by

$$\text{and} \quad \left. \begin{aligned} A_{wa} &= A_a(1 + \varepsilon - \Theta)^\sigma \Theta^\tau \\ A_{ws} &= A_w \Theta^\mu, \end{aligned} \right\} \quad (2.17)$$

where ε is a parameter to represent that when the soil surface is opened to the atmosphere, its water–air interfacial area is non-zero when the soil is fully saturated; others are soil parameters.

2.3. The model

The rate of O_2 dissolution is much faster than its diffusion in water as commonly assumed in hydrogen fuel cells [41]. Since microbial reduction of O_2 is slower than electrochemical reduction in fuel cells, all analysis in what follows is based on equation (2.13). For a soil sample, combining equation (2.13) and equation (2.17) gives

$$\left. \begin{aligned} \Omega(\Theta, T) &= k \cdot A_w \Theta^\mu \cdot C_{\text{eq}} \cdot E_r, \\ E_r &= \frac{1}{1 + 2 \cdot k \cdot \lambda \cdot \Theta^{\mu-\tau}(1 + \varepsilon - \Theta)^{-\sigma} [D \cdot A + D \cdot A^2 \cdot \Theta^{\tau-\mu}(1 + \varepsilon - \Theta)^\sigma]^{-1}}, \\ A &= A_a/A_w \end{aligned} \right\} \quad (2.18)$$

and $k = n'(1 - \beta) \cdot \kappa \cdot v \cdot \exp \left(-\frac{E_a}{R(T+273)} \right) \frac{C_D}{k_D + C_D}.$

2.4. Implementation

Equation (2.18) is used to calculate soil respiration rates. The influence of temperature and substrates is described by the parameter k . Apart from k , all other parameters are related to soil architecture. Mathematically, each parameter in equation (2.18) can take an arbitrary value, but in validating the model against experimental data, we take all soil structure parameters as a set, calculating it by mining a soil image dataset consisting of more than 100 X-ray images we have accumulated over the past decade for soils with various textures ranging from clay loam to sandy soils [45–49]. Using a method we developed previously [50], we calculated liquid water distribution within the pore geometry at different water contents, as well as the associated water–air interfaces and wetted pore walls for each soil image. For each soil image, we substituted its water–air interfacial areas and wetted pore walls into equations (2.17) and (2.18), and then adjusted parameter k , which can be either constant or varies with pore size (see the Results and discussion section for details), until the calculated respiration rates from equation (2.18) matched experimental data.

3. Results and discussion

3.1. Comparison with experimental data

Our first example is an incubation experiment that measured soil respiration rates under different soil saturations using a repacked sandy soil [51]. All respiration rates were normalized by the maximum respiration at the optimal soil water content. Figure 2a compares the measured and modelled results using the soil architecture shown in the electronic supplementary material, figure S1A. They agreed well, indicating the model captures the key processes underpinning respiration at different soil saturations.

Our second example is a field experiment designed to investigate the response of soil respiration to changes in soil moisture [52]. Field soil is more heterogeneous than sieved and repacked soil. Its respiration often exhibits the ‘Birch’ phenomenon [53], suggesting that microbes in small pores respire less than those in large pores due to pore-scale variation in substrates and microbial composition [54,55]. We modelled this by allowing k to increase with pore size. Since water progressively fills small to large pores as soil

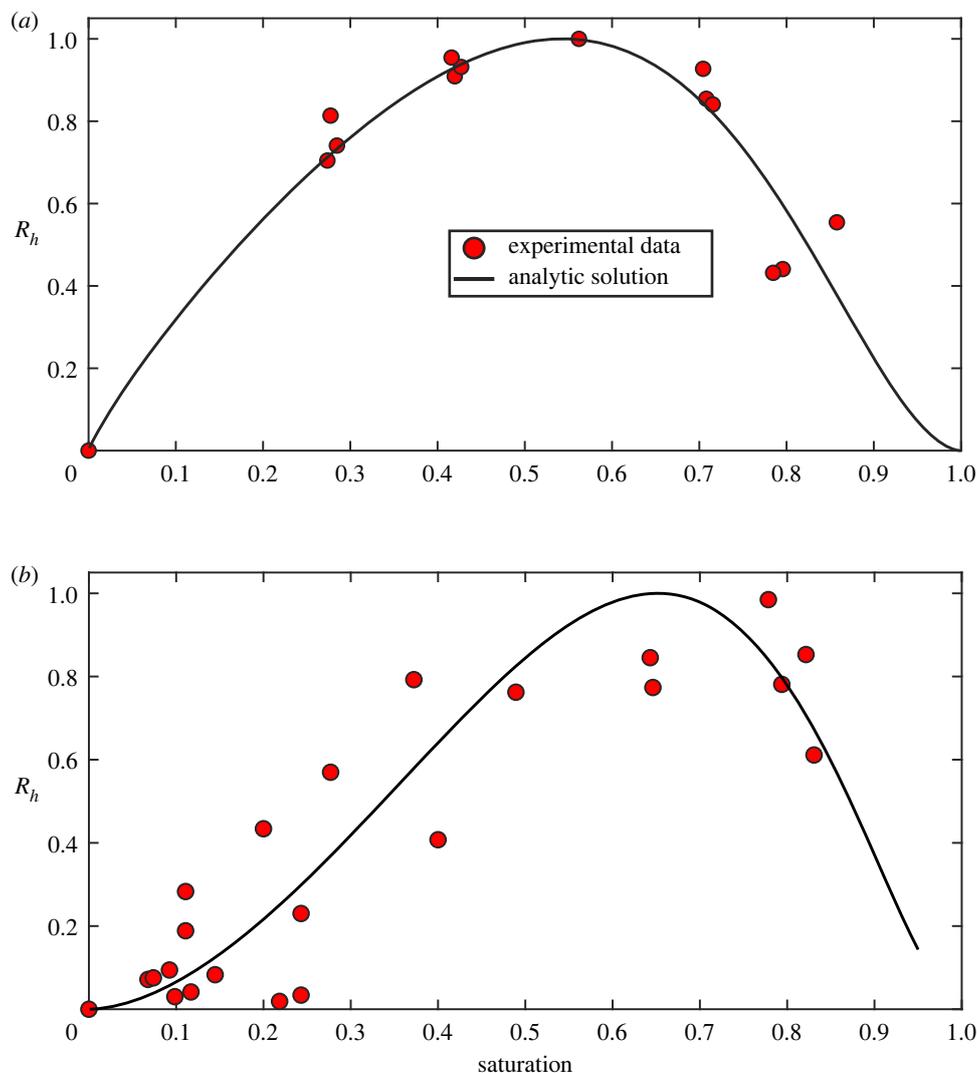


Figure 2. Comparison of experimental data (symbols) and model results (solid lines) using exemplar soil architectures under different soil saturations from an incubation experiment using a well-mixed sandy soil where the potential demand of aerobic microbes for O_2 was relatively uniform (a); for a field experiment using a loamy soil where microbes and substrates are sparser in small pores than in large pores (b).

water increases, we described this pore-scale substrate and microbial heterogeneity by allowing k to increase linearly with soil water content. Figure 2b shows the comparison. The model reproduces the change in respiration from dry to wet soil, revealing that O_2 dissolution and diffusion in soil water regulates the response of aerobic heterotrophic soil respiration to soil water change.

Our final example was chosen to demonstrate that the model captures the nonlinear coupling between soil water and temperature in their effect on respiration using an incubation experiment that involved both moisture and temperature gradients [56]. The influence of soil water content was measured experimentally by maintaining a constant temperature, while the effect of temperature was measured by keeping the soil moisture content constant. We first calculated the soil structure parameters by calibrating the model against respiration rates measured at different saturations at 15°C and then used these parameters and equation (2.15) to predict respiration rate variation when temperature was increased from 5 to 30°C . The molecular diffusion coefficient of dissolved O_2 and the saturated O_2 concentration at different temperatures were calculated from equation (2.16). The potential demand of the reactive sites for O_2 at temperature T is k_T , calculated as follows

based on its value at 15°C (T_{15}) calibrated for obtaining the soil structure parameters:

$$k_T = k_{15} \exp\left[\frac{E_a}{R} \left(\frac{1}{T_{15}} - \frac{1}{T}\right)\right], \quad (3.1)$$

where k_{15} is the value of k at 15°C , and T_{15} and T are absolute temperature (K).

Moriyama *et al.* [56] estimated the apparent activation energy based on respiration rates measured across a range of temperatures, effectively a bulk estimate representing the collective impact of all factors which influence the temperature response of soil respiration. In our model, E_a is intrinsic, determined by microbial composition and molecular structures of the substrates. However, because substrates and microbial composition vary with pore size [55,57], even the intrinsic E_a itself is by necessity an average value, representing the average of the intrinsic E_a of the substrates and microbes on different hydrated reactive sites. In this example, we considered the average intrinsic E_a as an unknown and calibrated it to obtain the results which matched the measured respiration rates at different temperatures.

For comparing respiration rates, we normalized all variables in the model and multiplied the dimensionless

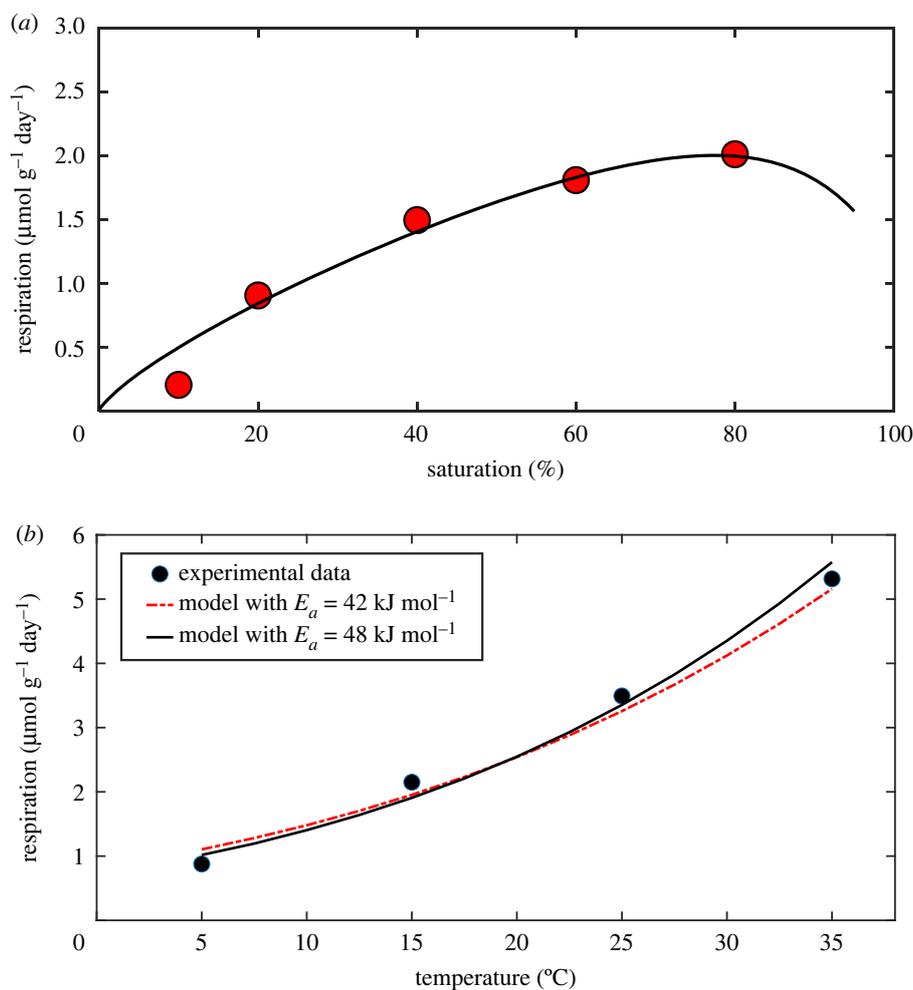


Figure 3. (a) Comparison of measured (symbols) and calculated respiration rates (solid line) using calibrated soil structure and potential O_2 demand when soil saturation is increased from 7 to 80% at 15°C. (b) The calibrated model was used to predict respiration rates for temperatures between 5 and 35°C at a constant water saturation of 60%. The increase in microbial metabolic activity with temperature is described by the Arrhenius kinetic model with an intrinsic activation energy (E_a) of 48 kJ mol^{-1} . For comparison, we also calculated respiration rates using the bulk E_a of 42 kJ mol^{-1} measured from the experimental data [56].

respiration rates calculated using these normalized variables by a single scalar to match the experimental data. As an illustration, figure 3a compares the measured and calculated respiration rates under different soil water saturations at 15°C; figure 3b compares the measured and predicted respiration rates when the soil was 60% saturated as temperature increases from 5 to 35°C using an intrinsic E_a of 48 kJ mol^{-1} . They agree well, indicating that the combined influence of soil water and temperature on soil respiration is nonlinear and is captured well by our model. For comparison, we also calculated the temperature response of respiration rates directly using the bulk E_a (45 kJ mol^{-1}) reported in Moriyama *et al.* [56] and plot the results in figure 5b. The predictions using the intrinsic $E_a = 48 \text{ kJ mol}^{-1}$ are slightly better. Because of the limited experimental data, in terms of matching the measured data, the difference between the two activation energies is not significant, but it corroborates that intrinsic E_a is higher than bulk E_a [5].

3.2. Attenuated temperature response of respiration

Nonlinear coupling of the influence of soil water content and temperature in equation (2.18) indicates that the temperature response of respiration is regulated by soil water content, and the strength of their coupling is modulated by E_a . This differs

from previous studies which attributed the temperature response of respiration to microbial physiology and substrate quality [58–65]. Equation (2.18) separates soil structure and its associated physical processes from other factors in their role in mediating the temperature response of soil respiration. This is important but has been overlooked. For example, increasing temperature from 5 to 35°C reduces the saturated O_2 concentration in water from approximately 14 to 7 mg l^{-1} [36]; ignoring this fall would overestimate the thermal adaptation of soil microbes. The effect of these physical factors is described by the feedback factor (equation (2.16)), whose magnitude varies with soil water content, E_a and temperature (electronic supplementary material, figures S2).

The attenuated temperature response of soil respiration is regulated by both E_a and soil water content. Figure 4 shows the change in the feedback factor with temperature at different soil saturations (figure 4a) and E_a (figure 4b), respectively, calculated using soil parameters for the example in figure 3. To allow comparison, the feedback factor calculated for each saturation (or E_a), was normalized by the value of the feedback factor at 5°C. Depending on soil saturation (or E_a), increasing temperature from 5 to 35°C could dampen respiration rates by approximately 60% due to reduced O_2 dissolution and increased hydraulic resistance against O_2 flux from the water–air interface to the reactive sites as temperature rises.

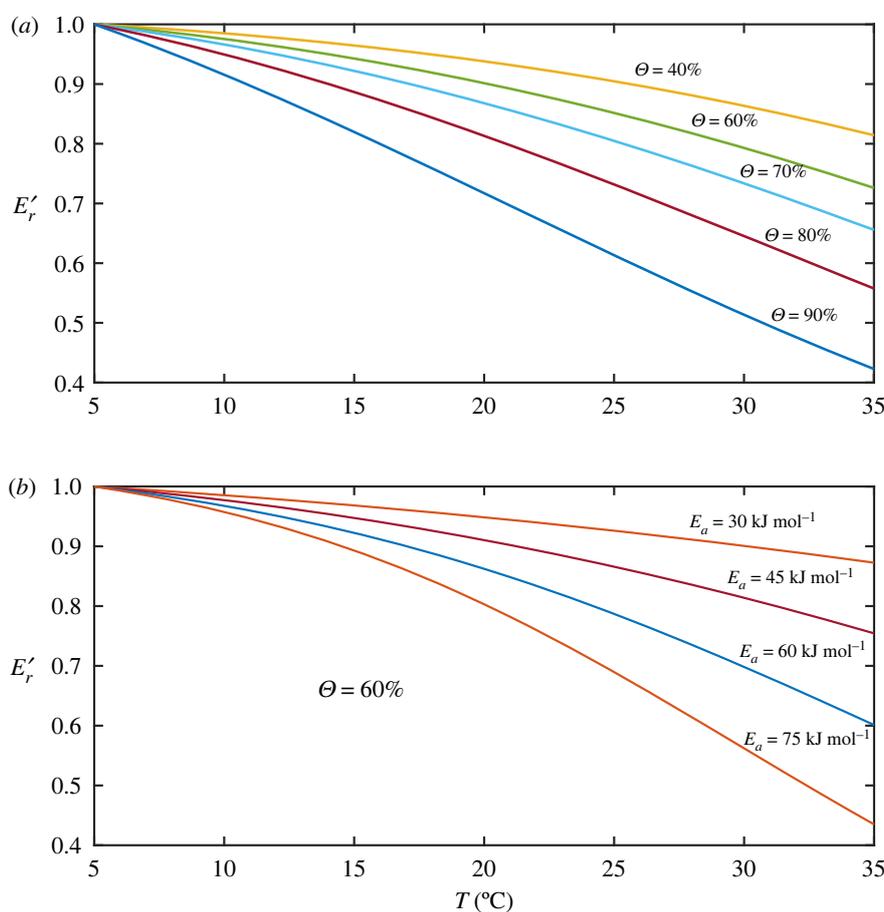


Figure 4. The feedback factor describing the attenuation of respiration rates decreases with the increase in soil saturation (a) and activation energy E_a (b). E'_r is the ratio between the feedback factor at temperature T to the feedback factor at 5°C .

This is consistent with experimental results that reducing O_2 supply substantially reduced respiration [52]; it is also corroborated by the results of a whole soil profile experiment where E_a in the subsoil was greater than that in the topsoil [66,67]. While biological factors such as differences in SOC quality and microbial community composition between the top- and sub-soils are also likely to be important [68,69], O_2 dissolution and diffusion are critical factors because the subsoil is more saturated, and the preferential consumption of O_2 by roots and microbes in the topsoil limits O_2 diffusion to reactive sites in the subsoil. Not accounting explicitly for the reduced O_2 dissolution and diffusion in the subsoil would underestimate the intrinsic temperature sensitivity of soil respiration, giving rise to a reduced E_a or Q_{10} .

3.3. Non-unique optimal soil moisture for aerobic respiration

The analytical model derived from the volumetric-average reveals that the influence of soil water content and temperature on soil respiration is more complicated than described by separate moisture and temperature functions [70]. For each temperature, however, there is still an optimal soil water content at which respiration rate peaks when other factors are fixed. Normalizing respiration rates at different soil water contents by this maximum gives a curve which is equivalent to the moisture function used in most SOC models [15]. Taking the soil parameters calibrated for figure 3 as an example, we calculated the moisture function

at different temperatures with other factors kept constant (figure 5a). It is evident that for the same soil, the optimal soil saturation varies with temperature. As temperature increases from 2 to 30°C for the example shown in figure 5a, the optimal soil saturation decreases, from approximately 80 to 55% ; this range covers the soil saturation deemed optimal (60%) for aerobic microbes used in most incubation experiments [9,56,59].

In soils lacking liable carbon substrates, the respiration rate measured in incubation experiments typically increases linearly as soil water increases rather than following the trend shown in figures 2 and 3 [72]. Even when soils are approximately saturated, a significant amount of CO_2 continues to emit from the soils [56,71]. The likely reason is that the soil surface remains open to the atmosphere. When soil is fully saturated with water, O_2 continues to dissolve at the soil surface and move into the soil. Thus, depending on the quantity and quality of SOC within the soil and the soil surface areas, the change in respiration with soil water content can be either approximately linear, nonlinear or bell-shaped; all these responses are captured by our model. As an illustrative example, figure 5b compares how the soil surface opening affects the response of respiration to soil water content when microbial activity is low and other parameters are the same.

3.4. Substrate and microbial heterogeneity

Aerobic respiration measured during short-term experiments exhibits an exponential increase as temperature rises [10,38],

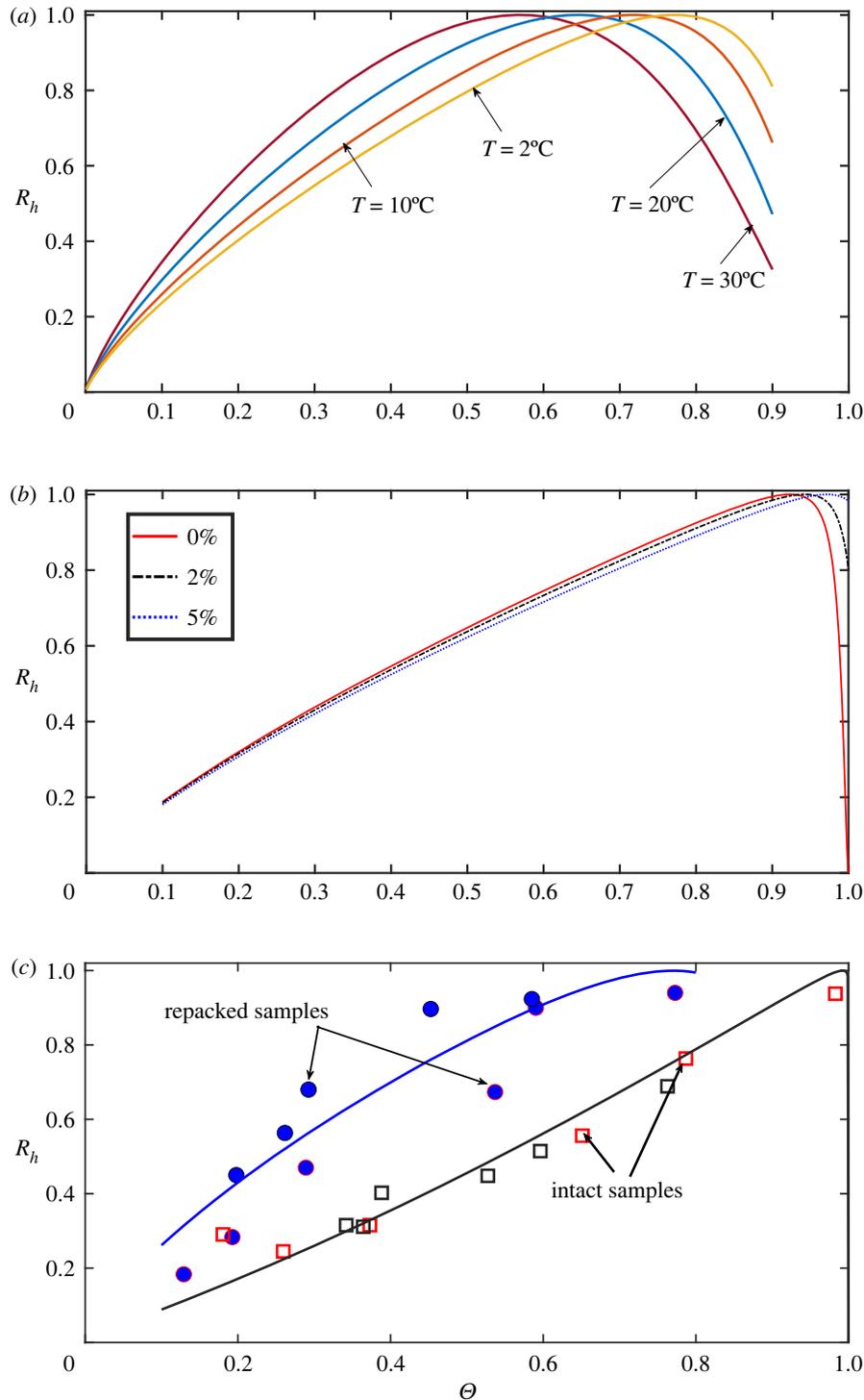


Figure 5. (a) The optimal saturation for maximal aerobic heterotrophic respiration (R_h) traditionally used in moisture functions is not unique, but is temperature (T) dependent because of its nonlinear coupling with soil moisture. (b) Maintaining the soil surface open in incubation experiments alters the response of R_h to soil water when soil is close to saturation; the effects increase from non-open (0%) to having 5% of pores in direct contact with the atmosphere. (c) Accounting for microbial and substrate heterogeneities explains the variation of experimentally measured saturation–respiration relationships [71]; open squares are data measured from a field-structured loamy soil; solid circles are data measured after sieving-repacking; solid lines are results calculated from the model.

while the change in respiration with soil moisture is inconsistent, ranging from a linear increase [73,74], concave increase [51], convex increase [71], to convex increase followed by a plateau before declining [52]. Repacking a sieved soil can also dramatically change the moisture response of respiration compared with undisturbed soil [71]. In the latter, physical constraints are likely to prevent microbes from entering small pores, and there is evidence that substrate quality in small pores is less energetically favourable than those in

large pores [57]. These heterogeneities are a possible cause of the broader variation in the moisture response of soil respiration. Representing all these heterogeneities in a single analytical model is a formidable task, but their effects can still be quantitatively accounted for by allowing the density of microbial numbers on wetted pore walls and substrate concentrations to vary with pore size [54]. For example, when soil water content is low, only small pores are filled by water in which microbes are less active as most microbes in

larger pores are in dormancy [75]. As soil water content increases and larger pores become progressively refilled by water, dormant microbes become more active and substrate availability increases contemporaneously. Allowing the density of microbes over the wetted pore walls and substrate concentration to increase with pore size in the model can represent these pore-scale heterogeneities to produce a diverse set of saturation–respiration relationships. Figure 5c shows how including such pore-scale heterogeneity reshapes the moisture response of respiration rates, in comparison with an experiment which measured respiration from intact and repacked soil cores [71]. We acknowledge that other factors are also likely to play a role in these diverse respiration–saturation relationships as those in figure 5c, but here we highlight the importance of microscopic soil architecture and physical processes which are typically overlooked in most data analysis and SOC models. Under certain circumstances, they might overwhelm biotic factors and physiological change in microbes in mediating the moisture and temperature response of microbial activity [13].

4. Conclusion

We develop a volumetric-average method, with soil architecture and microscopic physical processes represented explicitly, to calculate aerobic respiration analytically from soil samples. Soil water content in the model is the result of the volumetric-average, and it is nonlinearly coupled with temperature and other factors. Comparison with experimental data shows the model reproduces respiration rates measured from soils with both water content and temperature gradients. Incorporating microbial and substrate heterogeneities into the model can explain the diverse moisture– and temperature–respiration relationships. The model demonstrates that, alongside thermal adaptation, substrate heterogeneity and carbon use efficiency of microbes, O_2 dissolution and diffusion in soil water attenuate the temperature response of soil respiration. Overlooking these mechanisms in data analysis risks incorrectly ascribing their influence to biological factors, thereby overestimating the role of microbes and substrate heterogeneity in regulating the temperature response of soil respiration. The next generation of SOC models should therefore consider soil architecture and microscopic physical processes.

Data accessibility. Soil images used to calculate the results presented in this paper are available from https://figshare.com/articles/data_set/Soil_respiration/20198717.

Authors' contributions. X.Z.: conceptualization, data curation, formal analysis, investigation, methodology, software, validation, writing—original draft and writing—review and editing; P.A.W.: investigation, methodology and writing—review and editing; A.S.G.: investigation, methodology and writing—review and editing; A.L.N.: investigation, methodology and writing—review and editing; K.C.: methodology and writing—review and editing; W.R.W.: conceptualization, funding acquisition, methodology, project administration, supervision and writing—review and editing; S.J.M.: conceptualization, methodology and writing—review and editing; T.H.I.: conceptualization, funding acquisition, methodology, project administration, supervision and writing—review and editing; K.S.: conceptualization, funding acquisition, methodology, project administration and supervision.

All authors gave final approval for publication and agreed to be held accountable for the work performed therein.

Conflict of interest declaration. We declare that we have no competing interests.

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Appendix A

See table 1.

Table 1. Nomenclature.

A_{ws}	specific water–pore wall interfacial area (cm^2)
A_{wa}	specific water–air interfacial area (cm^2)
A_w	specific water–pore wall interfacial area when soil is saturated (cm^2)
A_a	parameter in the specific water–air interfacial area (cm^{-1})
C_{eq}	saturated dissolved O_2 concentration at water–air interface (mg l^{-1})
C_o	average dissolved O_2 concentration at wetted pore wall (mg l^{-1})
c_o	dissolved O_2 concentration (mg l^{-1})
c_D	dissolved organic carbon concentration (mg l^{-1})
D	diffusion coefficient of dissolved O_2 ($\text{cm}^2 \text{s}^{-1}$)
D_D	diffusion coefficient of dissolved organic carbon ($\text{cm}^2 \text{s}^{-1}$)
E_a	activation energy (kJ mol^{-1})
E_r	feedback factor
k_D	Michaelis–Menten constant for dissolved organic carbon (mg l^{-1})
k_o	Michaelis–Menten constant for dissolved O_2 (mg l^{-1})
L	average distance between water–air interface and wetted pore wall (cm)
N	number of aerobic microbes in a unit volume of water
n	number of the aerobic microbes associated with a unit area of wetted pore wall
R	gas constant (J mol^{-1})
R_h	heterotrophic respiration ($\mu\text{mol g}^{-1} \text{d}^{-1}$)
s_{DOC}	dissolution rate of polymerized carbon to a unit volume ($\text{mg l}^{-1} \text{s}^{-1}$)
T	temperature (K)
u_{max}	pro-exponential constant (mg s^{-1})
v_{max}	maximum microbial consumption rate of O_2 (mg s^{-1})
Θ	saturation
α	dissolution rate of gaseous O_2 at water–air interface (cm s^{-1})
μ	parameter characterizing wetted pore wall areas
τ	parameter characterizing water–air areas
σ	parameter characterizing water–air areas
λ	thickness of the thin water layer inhabited by microbes (cm)
κ	constant control microbial uptake of O_2 (l mg^{-1})

References

- Lehmann J, Kleber M. 2015 The contentious nature of soil organic matter. *Nature* **528**, 60–68. (doi:10.1038/nature16069)
- Button ES, Pett-Ridge J, Murphy DV, Kuzyakov Y, Chadwick DR, Jones DL. 2022 Deep-C storage: biological, chemical and physical strategies to enhance carbon stocks in agricultural subsoils. *Soil Biol. Biochem.* **170**, 108697. (doi:10.1016/j.soilbio.2022.108697)
- Carey JC *et al.* 2016 Temperature response of soil respiration largely unaltered with experimental warming. *Proc. Natl Acad. Sci. USA* **113**, 13 797–13 802. (doi:10.1073/pnas.1605365113)
- Davidson EA, Samanta S, Caramori SS, Savage K. 2012 The dual Arrhenius and Michaelis-Menten kinetics model for decomposition of soil organic matter at hourly to seasonal time scales. *Glob. Change Biol.* **18**, 371–384. (doi:10.1111/j.1365-2486.2011.02546.x)
- Davidson EA, Janssens IA. 2006 Temperature sensitivity of soil carbon decomposition and feedbacks to climate change. *Nature* **440**, 165–173. (doi:10.1038/nature04514)
- Young IM, Crawford JW. 2004 Interactions and self-organization in the soil-microbe complex. *Science* **304**, 1634–1637. (doi:10.1126/science.1097394)
- Ebrahimi A, Or D. 2016 Microbial community dynamics in soil aggregates shape biogeochemical gas fluxes from soil profiles – upscaling an aggregate biophysical model. *Glob. Change Biol.* **22**, 3141–3156. (doi:10.1111/gcb.13345)
- Hartley IP, Hopkins DW, Garnett MH, Sommerkorn M, Wookey PA. 2008 Soil microbial respiration in Arctic soil does not acclimate to temperature. *Ecol. Lett.* **11**, 1092–1100. (doi:10.1111/j.1461-0248.2008.01223.x)
- Dacal M, Bradford MA, Plaza C, Maestre FT, Garcia-Palacios P. 2019 Soil microbial respiration adapts to ambient temperature in global drylands. *Nat. Ecol. Evol.* **3**, 232. (doi:10.1038/s41559-018-0770-5)
- Fang CM, Smith P, Moncrieff JB, Smith JU. 2005 Similar response of labile and resistant soil organic matter pools to changes in temperature. *Nature* **433**, 57–59. (doi:10.1038/nature03138)
- Nottingham AT, Meir P, Velasquez E, Turner BL. 2020 Soil carbon loss by experimental warming in a tropical forest. *Nature* **584**, 234. (doi:10.1038/s41586-020-2566-4)
- Keiluweit M, Bougoure JJ, Nico PS, Pett-Ridge J, Weber PK, Kleber M. 2015 Mineral protection of soil carbon counteracted by root exudates. *Nat. Clim. Chang.* **5**, 588–595. (doi:10.1038/nclimate2580)
- Schimel JP. 2018 Life in dry soils: effects of drought on soil microbial communities and processes. In *Annual review of ecology, evolution, and systematics*, vol. 49 (ed. DJ Futuyma), pp. 409–432. Palo Alto, CA: Annual Reviews.
- Butcher KR, Nasto MK, Norton JM, Stark JM. 2020 Physical mechanisms for soil moisture effects on microbial carbon-use efficiency in a sandy loam soil in the western United States. *Soil Biol. Biochem.* **150**, 10. (doi:10.1016/j.soilbio.2020.107969)
- Moyano FE, Manzoni S, Chenu C. 2013 Responses of soil heterotrophic respiration to moisture availability: an exploration of processes and models. *Soil Biol. Biochem.* **59**, 72–85. (doi:10.1016/j.soilbio.2013.01.002)
- Bauer J, Herbst M, Huisman JA, Weiermuller L, Vereecken H. 2008 Sensitivity of simulated soil heterotrophic respiration to temperature and moisture reduction functions. *Geoderma* **145**, 17–27. (doi:10.1016/j.geoderma.2008.01.026)
- Meyer N, Welp G, Amelung W. 2018 The temperature sensitivity (Q10) of soil respiration: controlling factors and spatial prediction at regional scale based on environmental soil classes. *Glob. Biogeochem. Cycle* **32**, 306–323. (doi:10.1002/2017GB005644)
- Zhou T, Shi P, Hui D, Luo Y. 2009 Global pattern of temperature sensitivity of soil heterotrophic respiration (Q10) and its implications for carbon-climate feedback. *J. Geophys. Res. Biogeosci.* **114**, G02016. (doi:10.1029/2008JG000850)
- Balogh J, Pinter K, Fóti S, Cserhalmi D, Papp M, Nagy Z. 2011 Dependence of soil respiration on soil moisture, clay content, soil organic matter, and CO₂ uptake in dry grasslands. *Soil Biol. Biochem.* **43**, 1006–1013. (doi:10.1016/j.soilbio.2011.01.017)
- Quan Q, Tian DS, Luo Y, Zhang F, Crowther TW, Zhu K, Chen HYH, Zhou Q, Niu S. 2019 Water scaling of ecosystem carbon cycle feedback to climate warming. *Sci. Adv.* **5**, 7. (doi:10.1126/sciadv.aav1131)
- Rey A, Petsikos C, Jarvis PG, Grace J. 2005 Effect of temperature and moisture on rates of carbon mineralization in a Mediterranean oak forest soil under controlled and field conditions. *Eur. J. Soil Sci.* **56**, 589–599. (doi:10.1111/j.1365-2389.2004.00699.x)
- Moyano FE, Vasilyeva N, Menichetti L. 2018 Diffusion limitations and Michaelis-Menten kinetics as drivers of combined temperature and moisture effects on carbon fluxes of mineral soils. *Biogeosciences* **15**, 5031–5045. (doi:10.5194/bg-15-5031-2018)
- Fer M, Kodesova R, Kalkusova B, Klement A, Nikodem A. 2020 An empirical model for describing the influence of water content and concentration of sulfamethoxazole (antibiotic) in soil on the total net CO₂ efflux. *J. Hydrol. Hydromech.* **68**, 351–358. (doi:10.2478/johh-2020-0031)
- Liu YC, Lawrence CR, Winnick MJ, Hsu HT, Maher K, Druhan JL. 2019 Modeling transient soil moisture limitations on microbial carbon respiration. *J. Geophys. Res. Biogeosci.* **124**, 2222–2247. (doi:10.1029/2018jg004628)
- Reichstein M *et al.* 2003 Modeling temporal and large-scale spatial variability of soil respiration from soil water availability, temperature and vegetation productivity indices. *Glob. Biogeochem. Cycle* **17**, 15. (doi:10.1029/2003gb002035)
- Dungait JAJ, Hopkins DW, Gregory AS, Whitmore AP. 2012 Soil organic matter turnover is governed by accessibility not recalcitrance. *Glob. Change Biol.* **18**, 1781–1796. (doi:10.1111/j.1365-2486.2012.02665.x)
- Lehmann J *et al.* 2020 Persistence of soil organic carbon caused by functional complexity. *Nat. Geosci.* **13**, 529–534. (doi:10.1038/s41561-020-0612-3)
- Sihi D, Davidson EA, Savage KE, Liang D. 2020 Simultaneous numerical representation of soil microsite production and consumption of carbon dioxide, methane, and nitrous oxide using probability distribution functions. *Glob. Change Biol.* **26**, 200–218. (doi:10.1111/gcb.14855)
- Manzoni S, Moyano F, Katterer T, Schimel J. 2016 Modeling coupled enzymatic and solute transport controls on decomposition in drying soils. *Soil Biol. Biochem.* **95**, 275–287. (doi:10.1016/j.soilbio.2016.01.006)
- Totsche KU *et al.* 2018 Microaggregates in soils. *J. Plant Nutr. Soil Sci.* **181**, 104–136. (doi:10.1002/jpln.201600451)
- Harris E *et al.* 2021 Denitrifying pathways dominate nitrous oxide emissions from managed grassland during drought and rewetting. *Sci. Adv.* **7**, eabb7118. (doi:10.1126/sciadv.abb7118)
- Leuther F, Kohne JM, Metreveli G, Vogel HJ. 2020 Transport and retention of sulfidized silver nanoparticles in porous media: the role of air-water interfaces, flow velocity, and natural organic matter. *Water Resour. Res.* **56**, 19. (doi:10.1029/2020wr027074)
- Fang C, Moncrieff JB. 1999 A model for soil CO₂ production and transport 1: model development. *Agric. For. Meteorol.* **95**, 225–236. (doi:10.1016/s0168-1923(99)00036-2)
- Keiluweit M, Wanzek T, Kleber M, Nico P, Fendorf S. 2017 Anaerobic microsites have an unaccounted role in soil carbon stabilization. *Nat. Commun.* **8**, 10. (doi:10.1038/s41467-017-01406-6)
- Angert A, Yakir D, Rodeghiero M, Preisler Y, Davidson EA, Weiner T. 2015 Using O₂ to study the relationships between soil CO₂ efflux and soil respiration. *Biogeosciences* **12**, 2089–2099. (doi:10.5194/bg-12-2089-2015)
- Han P, Bartels DM. 1996 Temperature dependence of oxygen diffusion in H₂O and D₂O. *J. Phys. Chem.* **100**, 5597–5602. (doi:10.1021/jp952903y)
- Nunan N, Schmidt H, Raynaud X. 2020 The ecology of heterogeneity: soil bacterial communities and C dynamics. *Phil. Trans. R. Soc. B* **375**, 20190249. (doi:10.1098/rstb.2019.0249)
- Davidson EA, Belk E, Boone RD. 1998 Soil water content and temperature as independent or confounded factors controlling soil respiration in a temperate mixed hardwood forest. *Glob. Change Biol.* **4**, 217–227. (doi:10.1046/j.1365-2486.1998.00128.x)
- Bunnell FL, Tait DEN, Flanagan PW, Van Clever K. 1977 Microbial respiration and substrate weight

- loss—I: a general model of the influences of abiotic variables. *Soil Biol. Biochem.* **9**, 33–40. (doi:10.1016/0038-0717(77)90058-X)
40. Yan ZF, Liu CX, Todd-Brown KE, Liu Y, Bond-Lamberty B, Bailey VL. 2016 Pore-scale investigation on the response of heterotrophic respiration to moisture conditions in heterogeneous soils. *Biogeochemistry* **131**, 121–134. (doi:10.1007/s10533-016-0270-0)
41. Zhang XX, Gao Y. 2016 Impact of liquid water on oxygen reaction in cathode catalyst layer of proton exchange membrane fuel cell: a simple and physically sound model. *J. Power Sourc.* **318**, 251–263. (doi:10.1016/j.jpowsour.2016.04.024)
42. Borer B, Tecron R, Or D. 2018 Spatial organization of bacterial populations in response to oxygen and carbon counter-gradients in pore networks. *Nat. Commun.* **9**, 11. (doi:10.1038/s41467-018-03187-y)
43. Yan ZF, Bond-Lamberty B, Todd-Brown KE, Bailey VL, Li SL, Liu CQ, Liu C. 2018 A moisture function of soil heterotrophic respiration that incorporates microscale processes. *Nat. Commun.* **9**, 10. (doi:10.1038/s41467-018-04971-6)
44. Ebrahimi A, Or D. 2015 Hydration and diffusion processes shape microbial community organization and function in model soil aggregates. *Water Resour. Res.* **51**, 9804–9827. (doi:10.1002/2015wr017565)
45. Bacq-Labreuil A, Crawford J, Mooney SJ, Neal AL, Akkari E, Mcauliffe C, Zhang X, Redmile-Gordon M, Ritz K. 2018 Effects of cropping systems upon the three-dimensional architecture of soil systems are modulated by texture. *Geoderma* **332**, 73–83. (doi:10.1016/j.geoderma.2018.07.002)
46. Zhang X *et al.* 2021 Relationship between soil carbon sequestration and the ability of soil aggregates to transport dissolved oxygen. *Geoderma* **403**, 115370. (doi:10.1016/j.geoderma.2021.115370)
47. Bacq-Labreuil A, Neal AL, Neal AL, Crawford J, Mooney SJ, Akkari E, Zhang X, Clark I, Ritz K. 2021 Significant structural evolution of a long-term fallow soil in response to agricultural management practices requires at least 10 years after conversion. *Eur. J. Soil Sci.* **72**, 829–841. (doi:10.1111/ejss.13037)
48. Neal AL, Bacq-Labreuil A, Zhang X, Clark IM, Coleman K, Mooney SJ, Ritz K, Crawford JW. 2020 Soil as an extended composite phenotype of the microbial metagenome. *Sci. Rep.* **10**, 10649. (doi:10.1038/s41598-020-67631-0)
49. Wang F, Zhang X, Neal AL, Crawford JW, Mooney SJ, Bacq-Labreuil A. 2022 Evolution of the transport properties of soil aggregates and their relationship with soil organic carbon following land use changes. *Soil Tillage Res.* **215**, 105226. (doi:10.1016/j.still.2021.105226)
50. Hu WL, Huang N, Zhang XX. 2014 Impact of saturation on mass transfer rate between mobile and immobile waters in solute transport within aggregated soils. *J. Hydrol.* **519**, 3557–3565. (doi:10.1016/j.jhydrol.2014.10.057)
51. Skopp J, Jawsom MD, Doran JW. 1990 Steady-state aerobic microbial activity as a function of soil water content. *Soil Sci. Soc. Am. J.* **54**, 1619–1625. (doi:10.2136/sssaj1990.03615995005400060018x)
52. Sierra CA, Malghani S, Loescher HW. 2017 Interactions among temperature, moisture, and oxygen concentrations in controlling decomposition rates in a boreal forest soil. *Biogeosciences* **14**, 703–710. (doi:10.5194/bg-14-703-2017)
53. Birch HF. 1958 The effect of soil drying on humus decomposition and nitrogen availability. *Plant Soil* **10**, 9–31. (doi:10.1007/BF01343734)
54. Kravchenko AN, Guber AK, Razavi BS, Koestel J, Quigley MY, Robertson GP, Kuzyakov Y. 2019 Microbial spatial footprint as a driver of soil carbon stabilization. *Nat. Commun.* **10**, 3121. (doi:10.1038/s41467-019-11057-4)
55. de Vries FT *et al.* 2018 Soil bacterial networks are less stable under drought than fungal networks. *Nat. Commun.* **9**, 3033. (doi:10.1038/s41467-018-05516-7)
56. Moriyama A, Yonemura S, Kawashima S, Du MY, Tang YH. 2013 Environmental indicators for estimating the potential soil respiration rate in alpine zone. *Ecol. Indic.* **32**, 245–252. (doi:10.1016/j.ecolind.2013.03.032)
57. Bailey VL, Smith AP, Tfaily M, Fansler SJ, Bond-Lamberty B. 2017 Differences in soluble organic carbon chemistry in pore waters sampled from different pore size domains. *Soil Biol. Biochem.* **107**, 133–143. (doi:10.1016/j.soilbio.2016.11.025)
58. Bradford MA. 2013 Thermal adaptation of decomposer communities in warming soils. *Front. Microbiol.* **4**, 16. (doi:10.3389/fmicb.2013.00333)
59. Bradford MA, Davies CA, Frey SD, Maddox TR, Melillo JM, Mohan JE, Reynolds JF, Treseder KK, Wallenstein MD. 2008 Thermal adaptation of soil microbial respiration to elevated temperature. *Ecol. Lett.* **11**, 1316–1327. (doi:10.1111/j.1461-0248.2008.01251.x)
60. Bradford MA, Watts BW, Davies CA. 2010 Thermal adaptation of heterotrophic soil respiration in laboratory microcosms. *Glob. Change Biol.* **16**, 1576–1588. (doi:10.1111/j.1365-2486.2009.02040.x)
61. Crowther TW, Bradford MA. 2013 Thermal acclimation in widespread heterotrophic soil microbes. *Ecol. Lett.* **16**, 469–477. (doi:10.1111/ele.12069)
62. Hemingway JD, Rothman DH, Grant KE, Rosengard SZ, Eglinton TI, Derry LA, Galy VV. 2019 Mineral protection regulates long-term global preservation of natural organic carbon. *Nature* **570**, 228. (doi:10.1038/s41586-019-1280-6)
63. Hartley IP, Heinemeyer A, Ineson P. 2007 Effects of three years of soil warming and shading on the rate of soil respiration: substrate availability and not thermal acclimation mediates observed response. *Glob. Change Biol.* **13**, 1761–1770. (doi:10.1111/j.1365-2486.2007.01373.x)
64. Hartley IP, Hopkins DW, Garnett MH, Sommerkorn M, Wookey PA. 2009 No evidence for compensatory thermal adaptation of soil microbial respiration in the study of Bradford *et al.* (2008). *Ecol. Lett.* **12**, E12–E14. (doi:10.1111/j.1461-0248.2009.01300.x)
65. Karhu K *et al.* 2014 Temperature sensitivity of soil respiration rates enhanced by microbial community response. *Nature* **513**, 81–84. (doi:10.1038/nature13604)
66. Pries CEH, Castanha C, Porras RC, Torn MS. 2017 The whole-soil carbon flux in response to warming. *Science* **355**, 1420–1422. (doi:10.1126/science.aal1319)
67. Xiao J, Yu F, Zhu W, Xu C, Zhang K, Luo Y, Tiedje JM, Zhou J, Cheng L. 2018 Comment on ‘The whole-soil carbon flux in response to warming’. *Science* **359**, eaao0218. (doi:10.1126/science.aao0218)
68. Dove NC, Torn MS, Hart SC, Tas N. 2021 Metabolic capabilities mute positive response to direct and indirect impacts of warming throughout the soil profile. *Nat. Commun.* **12**, 2089. (doi:10.1038/s41467-021-22408-5)
69. Eilers KG, Debenport S, Anderson S, Fierer N. 2012 Digging deeper to find unique microbial communities: the strong effect of depth on the structure of bacterial and archaeal communities in soil. *Soil Biol. Biochem.* **50**, 58–65. (doi:10.1016/j.soilbio.2012.03.011)
70. Xu X, Schimel JP, Thornton PE, Song X, Yuan F, Goswami S. 2014 Substrate and environmental controls on microbial assimilation of soil organic carbon: a framework for Earth system models. *Ecol. Lett.* **17**, 547–555. (doi:10.1111/ele.12254)
71. Herbst M, Tappe W, Kummer S, Vereecken H. 2016 The impact of sieving on heterotrophic respiration response to water content in loamy and sandy topsoils. *Geoderma* **272**, 73–82. (doi:10.1016/j.geoderma.2016.03.002)
72. Cook FJ, Orchard VA. 2008 Relationships between soil respiration and soil moisture. *Soil Biol. Biochem.* **40**, 1013–1018. (doi:10.1016/j.soilbio.2007.12.012)
73. Orchard VA, Cook FJ. 1983 Relationship between soil respiration and soil-moisture. *Soil Biol. Biochem.* **15**, 447–453. (doi:10.1016/0038-0717(83)90010-x)
74. Fierer N, Allen AS, Schimel JP, Holden PA. 2003 Controls on microbial CO₂ production: a comparison of surface and subsurface soil horizons. *Glob. Change Biol.* **9**, 1322–1332. (doi:10.1046/j.1365-2486.2003.00663.x)
75. Wang GS, Jagadamma S, Mayes MA, Schadt CW, Megan Steinweg J, Gu L, Post WM. 2015 Microbial dormancy improves development and experimental validation of ecosystem model. *ISME J.* **9**, 226–237. (doi:10.1038/ismej.2014.120)