



Review

Ruminant Grazing Lands in the Tropics: Silvopastoral Systems and *Tithonia diversifolia* as Tools with Potential to Promote Sustainability

Ana Maria Krüger ¹, Paulo de Mello Tavares Lima ^{1,2,*} , Vagner Ovani ¹ , Simón Pérez-Marquéz ^{1,3},
Helder Louvandini ¹ and Adibe Luiz Abdalla ¹

¹ Centro de Energia Nuclear na Agricultura, Universidade de São Paulo Av. Centenário, 303, Piracicaba 13400-970, SP, Brazil; anakruger@usp.br (A.M.K.); vagnerovani@usp.br (V.O.); simon.perez-marquez@rothamsted.ac.uk (S.P.-M.); louvandini@cena.usp.br (H.L.); abdalla@cena.usp.br (A.L.A.)

² Department of Animal Science, University of Wyoming, 1000 East University Avenue, Laramie, WY 82071, USA

³ Rothamsted Research, North Wyke, Okehampton EX20 2SB, UK

* Correspondence: pdemello@uwyo.edu

Abstract: Food security, sustainability of food production, and greenhouse gas (GHG) production of ruminant livestock are topics that generate scrutiny and debates worldwide. In a scenario of increasing human population and concerns with climate change, it is necessary to increase animal-derived food in sustainable operations. Grazing systems are crucial for ruminant production worldwide, and in the tropics, well-managed grasslands can provide sustainable intensification of this activity. In these regions, production often relies on grass monoculture managed extensively, a practice that commonly has led to the occurrence of degraded soils, limited animal productivity, and increased intensity of GHG emissions. Silvopastoralism is a practice that promotes several ecosystem services, showing potential to maintain soil quality while reducing the environmental impacts of ruminant production. These systems also have the potential to improve animal productive performance and reduce GHG emissions. The review was guided by a search in the Web of Science database using population terms and refined by document type (Article) and language (English OR Portuguese) following PRISMA protocol. Infographics were created using the Bibliometrix package in R software (version 4.3.2), and a specific topic on *Tithonia diversifolia* (Hemsl.) A. Gray was explored to demonstrate the importance of this forage resource for tropical silvopastoral systems and its potential contribution to food security. The *T. diversifolia* shrub is widely distributed in Latin America and tropical regions and presents several characteristics that make it a good option for silvopastoral systems. Focusing on the tropics, our objectives were to present one literature review addressing the role of grazing ruminant production towards the current climate change and food security challenges. Additionally, we aimed to explore the state of knowledge on silvopastoral systems and the use of *T. diversifolia*, presenting their potential to cope with this scenario of increased concerns with the sustainability of human activities.

Keywords: food security; grasslands; greenhouse gases; methane; shrubs



Citation: Krüger, A.M.; Lima, P.d.M.T.; Ovani, V.; Pérez-Marquéz, S.; Louvandini, H.; Abdalla, A.L. Ruminant Grazing Lands in the Tropics: Silvopastoral Systems and *Tithonia diversifolia* as Tools with Potential to Promote Sustainability. *Agronomy* **2024**, *14*, 1386. <https://doi.org/10.3390/agronomy14071386>

Academic Editors: Renata La Guardia Nave and Martin Gierus

Received: 16 April 2024

Revised: 23 May 2024

Accepted: 25 June 2024

Published: 27 June 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Grasslands are among the largest and most crucial ecosystems globally, serving as a primary pillar for human population development, particularly by supporting grazing ruminants' production [1]. Increased demand for animal food products, including milk and meat, is expected in the next few years due to a growing global population and changing dietary habits in many countries worldwide. Consequently, this trend will likely result in a greater demand for limited resources, such as land, fuel, water, and minerals [2]. Considering these factors, enhancing the production levels of animal food products is necessary to

meet these higher demands. However, at the same time, greenhouse gas (GHG) emissions from livestock production are identified as primary anthropogenic sources contributing to climate change. Enteric methane (CH_4) accounts for about 40% of emissions from the sector [3], with a global warming potential 25 times greater than that of CO_2 [4]. In addition to its environmental impacts, CH_4 production may be equivalent to a 2–12% loss of dietary gross energy [5], representing a limiting factor to animal productive performance. Therefore, achieving higher production should involve using sustainable practices, commonly referred to as sustainable intensification, to ensure that ruminant livestock production remains a feasible activity for both population and the planet [6,7].

The productivity (in terms of quality and quantity) of tropical grasses, the primary nutrient source for ruminants in the tropics, is often reduced during severe climatic conditions, leading to fluctuations in animal performance throughout the year. Various approaches to overcome this situation and maximize productivity in these grazing systems have been documented in the literature [1,8]. More recently, the use of trees or shrubs in grazing systems, an agroforestry practice named silvopastoralism, is gaining attention as a practice capable of providing benefits to forage production, animal performance, and the environment. Therefore, it represents a tool with the potential to enhance sustainability in tropical grazing systems [9,10].

Tithonia diversifolia (Hemsl.) A. Gray is a shrub species native to Central America but widespread in tropical and subtropical regions across the globe, showing promising potential for use in silvopastoral systems [11,12]. Compared to other tropical forages, *T. diversifolia* demonstrates greater crude protein (CP) and phosphorus content, maintaining a relatively stable nutritional value even during dry seasons while exhibiting tolerance to acidic soils and moderate to low water and fertilization demands [11,13,14]. Indigenous populations have traditionally used this plant for treating various diseases due to the presence of bioactive secondary compounds [15]. These compounds also influence ruminal fermentation, potentially reducing CH_4 emissions [11,16].

Given the characteristics of silvopastoral systems and *T. diversifolia*, as well as the relative novelty of both topics, it is evident that more comprehensive assessments of these in ruminant production, along with accurate analyses of the available data, are needed for the consolidation and elucidation of silvopastoralism and the use of this forage as a viable option for farmers in tropical regions to improve system productivity sustainably. Therefore, the objectives of the present review were to provide an overview of the role of grazing ruminant production towards the scenario of climate change and food security challenges as well as fundamental key aspects of the existing knowledge and potential impacts associated with the utilization of silvopastoralism in this context, with a specific focus on tropical systems and the utilization of the tropical shrub *T. diversifolia*.

2. Ruminant Production and Food Security—A Brief Overview

The food production sector is facing a challenging moment worldwide due to the increased number of people with inadequate access to nutritious diets, presenting insufficient daily intake of nutrients such as carbohydrates, proteins, and fats, leading to vitamin and mineral deficiencies. Simultaneously, issues like obesity and type two diabetes, which pose significant risks to human health, are increasingly prevalent in society [17,18]. Concurrently, there is a growing concern regarding the environmental impact of human activities. The livestock production sector is particularly noteworthy in this aspect, often cited as one of the primary contributors to these harmful effects on the environment [19,20]. Animal husbandry is identified as responsible for 14.5% of total anthropogenic GHG emissions in the atmosphere, with a very significant portion of these emissions, as mentioned earlier, being represented by the enteric CH_4 that arises from the fermentative process in the gastrointestinal tract of ruminants, serving as a byproduct from their feed digestion, especially structural carbohydrates [3,21]. However, products such as meat and milk obtained from ruminants can be part of well-balanced diets for humans, providing essential macro and micronutrients, including proteins, fatty acids, vitamins A, B12, calcium, iron, zinc, and

others, contributing significantly to promoting health in the population and reducing the occurrence of several illnesses [18,22,23].

The world's population growth, coupled with changes in the profile of societies such as increased average income and more widespread dissemination of the western lifestyle, leads to the estimation that the demand for animal food products will be 70% higher in 2050 relative to 2010 [2,24]. Ruminant production stands out in this scenario, as these animals can be reared on non-arable lands, consuming fibrous feedstuff to produce high-quality protein food, contributing significantly to achieving food security without relying on grains and other cereals that could be used in the human diet [6,25,26]. According to the Food and Agriculture Organization of the United Nations (FAO), food security entails "access to sufficient, safe, nutritious food to maintain a healthy and active life". Therefore, sustainable ruminant production should be crucial in meeting the increasing demand for food in human society while simultaneously achieving the targeted reductions of anthropogenic GHG emissions, frequently a focal point of discussion in climate-related scientific conferences [18,21,25,27].

Despite these facts, some scientists still propose a drastic reduction in the number of ruminants as a solution to avoid a climatic disaster [28]. However, the feasibility of this option must be analyzed in a broader context, considering the previously highlighted benefits that ruminant livestock production generates and the fact that livestock grasslands ecosystems support the livelihood of millions of people worldwide, providing economic goods and social-cultural services to these populations. Additionally, grasslands offer ecosystem services such as soil protection, maintenance of groundwaters quality, and climate regulation through carbon (C) sequestration, representing 25% of global soil sequestration potential [1,29–31].

Aiming for sustainability, ruminant production in pastoral systems should prioritize society's economic, environmental, social, and cultural demands. Productivity and the mitigation of GHG should serve as guiding principles to prevent ecological issues such as increased emissions and soil C loss, as well as shortage or unaffordable prices of animal food products [30]. Tropical grasslands are renowned as biodiversity hotspots, hosting several endangered species and serving as a pillar for environmental preservation [32]. As previously mentioned, tropical grasses are frequently susceptible to quality oscillations due to climatic factors, especially during dry seasons, when the plant may exhibit reduced CP content and increased structural carbohydrates, resulting in diminished productivity and increased GHG emissions per unit of generated product (i.e., GHG emissions' intensity) [33,34]. The use of silvopastoral systems by intercropping tropical forage grass species with native C_3 trees and shrubs is considered a management practice with potential to address the variation in forage productivity, as the presence of these trees and shrubs can enhance the system's resilience to extreme climate conditions, particularly during dry seasons, and contribute to increased biomass production, enhancing nutritive value of forages in such pastures, potentially resulting in a reduced intensity of CH_4 emissions [10,35].

3. Silvopastoralism in Ruminant Production

Due to increasing demand for animal products, especially in developing countries (which are mostly located in the world's tropical regions, particularly in Latin America), deforestation has occurred in order to expand pasture areas to support the growing requirements for higher animal production, since deforestation costs are usually lower than those associated with production intensification [36]. Approximately 70% of agricultural land is used for livestock production in the tropics. Pastures on these farms are commonly based on grass species cultivated in monocultural extensive systems, in which inadequate management practices often lead to issues like overgrazing and reduced soil productive potential, affecting animal performance and eventually leading to increased environmental impacts of the activity. Additionally, modest stocking rates are often observed, reducing productivity per land area [37,38]. Furthermore, the intensive use of inorganic fertilizers and biocides has diminished soil surface cover and destroyed crucial microbial communi-

ties responsible for soil ecosystem functions, contributing substantially to the deterioration of the physical, chemical, and biological properties of the soil [30]. Recognizing such factors has accelerated efforts towards sustainable intensification, which aims at increasing product generation per unit of area while simultaneously reverting soil degradation and enhancing ecosystem services [36].

The conscientious management of ruminants in agroforestry systems, particularly in silvopastoral systems which involve utilizing land for both forest products and animal production through the browsing of shrubs and trees and/or grazing of co-existing forage crops can significantly mitigate the ecological challenges posed by ruminant production systems [9]. Several tropical regions have implemented silvopastoral systems, reporting numerous benefits. To enhance our understanding of the current scenario regarding the utilization of these systems and the primary effects of their adoption, we conducted a brief systematic review following the Preferred Reporting Items for Systematic Review and Meta-Analyses (PRISMA) [39] guidelines. The Web of Science electronic database platforms were utilized to find papers using search population terms (i.e., TS = (silvopastoral OR silvipastoral OR silvopasture OR silvopastoralism)), refined by document type (i.e., Article) and language (i.e., “English” OR “Portuguese”). Intervention, comparison, and outcome terms were not used since the objective was to obtain an overview of the silvopastoral systems-related aspects being researched. The timeframe was extended until 5 March 2024.

This search yielded 1603 documents exported to BibTeX for evaluation through the Bibliometrix package [40] using R version 4.3.2 [41]. Over the last 8 years, we observed an exponential increase in published works on silvopastoral systems, reaching the highest number of documents published per year in 2022 (Figure 1C).

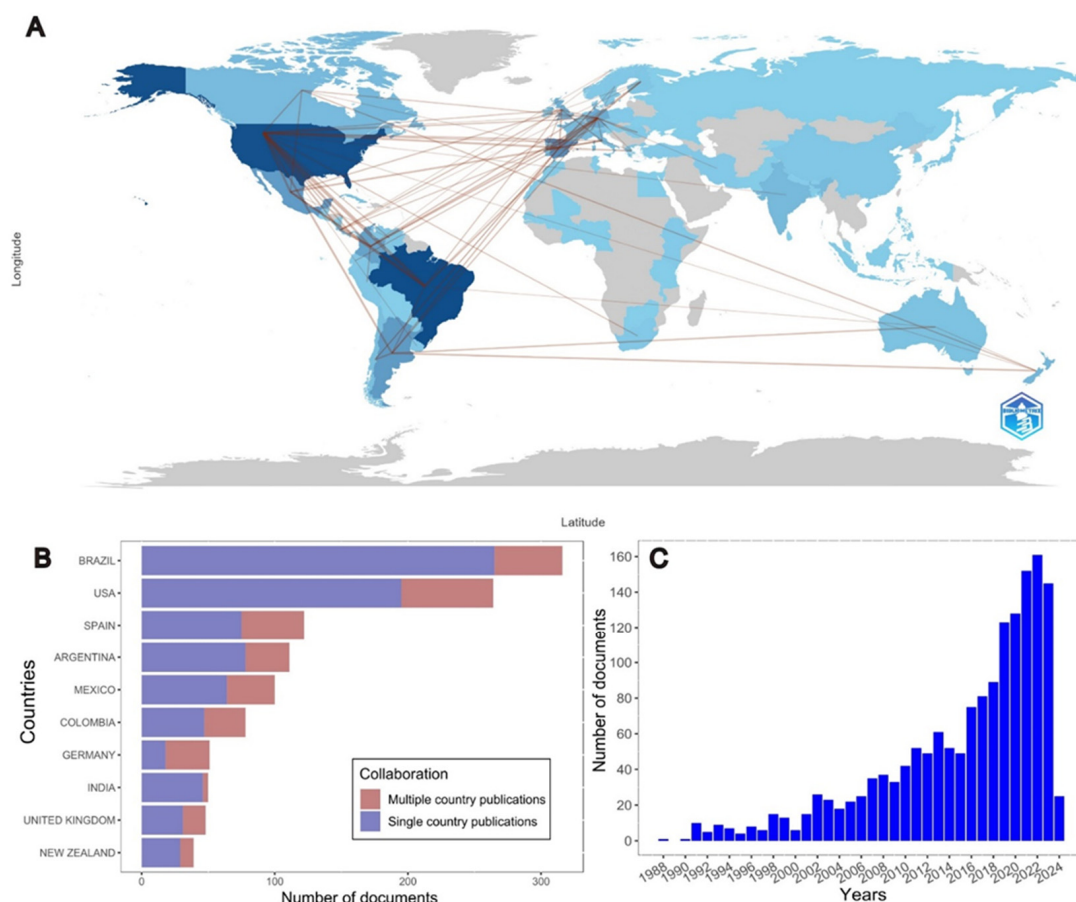


Figure 1. Country collaboration map (lines connecting countries indicate collaboration between themselves) (A), most relevant countries per corresponding authors (B), and annual total scientific production (C).

When evaluating the corresponding authors' data, it is evident that Brazil and the United States of America (USA) are the two countries that contribute the most to research in this area, respectively (Figure 1B), followed by Spain, Argentina, Mexico, and Colombia. This pattern influences the collaboration map, revealing a solid connection between Brazil, the USA, and other countries in South America, Central America, and Europe (Figure 1A).

The substantial number of studies on silvopastoral systems in South, Central, and North American countries was expected, given that many of these nations widely adopt ruminant production practices in pastures, including extensive tropical regions. For example, Brazil, with approximately 239 million hectares of agricultural land, and the USA, with about 405 million hectares, feature extensive permanent meadows and pastures covering around 173 million and 245 million hectares, respectively. Similarly, Argentina, Mexico, and Colombia, with agricultural land areas of around 117 million, 97 million, and 42 million hectares, respectively, each have substantial coverage of permanent meadows and pastures, approximately 74 million, 74 million, and 38 million hectares, respectively [42].

To comprehend the benefits of adopting silvopastoral systems, we analyzed the first 40 most frequently used keywords from these 1603 articles in a word cloud (Figure 2). Among the prominently featured words, in addition to “silvopastoral system”, are “forage production”, “management”, “pastures”, “carbon (C) sequestration”, and “nutritive value”, along with terms related to climate change, biodiversity, and ecosystem services.



Figure 2. Word cloud of the first 40 author's keywords.

Figure 3 was also derived from the author's keywords, with clusters formed on the X and Y axes. The X-axis signifies centrality, providing information about the importance of a theme, while the Y-axis symbolizes density, serving as a measure of the theme's development [43]. Consequently, four quadrants are formed: the motor themes (well-developed and crucial for structuring the research field), the niche themes (of limited importance for the field), the emerging or declining themes (weakly developed and marginal), and the basic themes (concerning general topics transversal to different research areas within the field) [43]. We observe that the motor themes for silvopastoral systems studies have been associated with C sequestration, and keywords focused on nutritive value, such as crude protein and digestibility. Additionally, other terms related to forage, such as grazing and leaf area index, are prominent. Meanwhile, basic and well-developed themes like soil fertility, greenhouse gases, animal welfare and behavior, shade, and forage production are highlighted in the fourth quadrant.

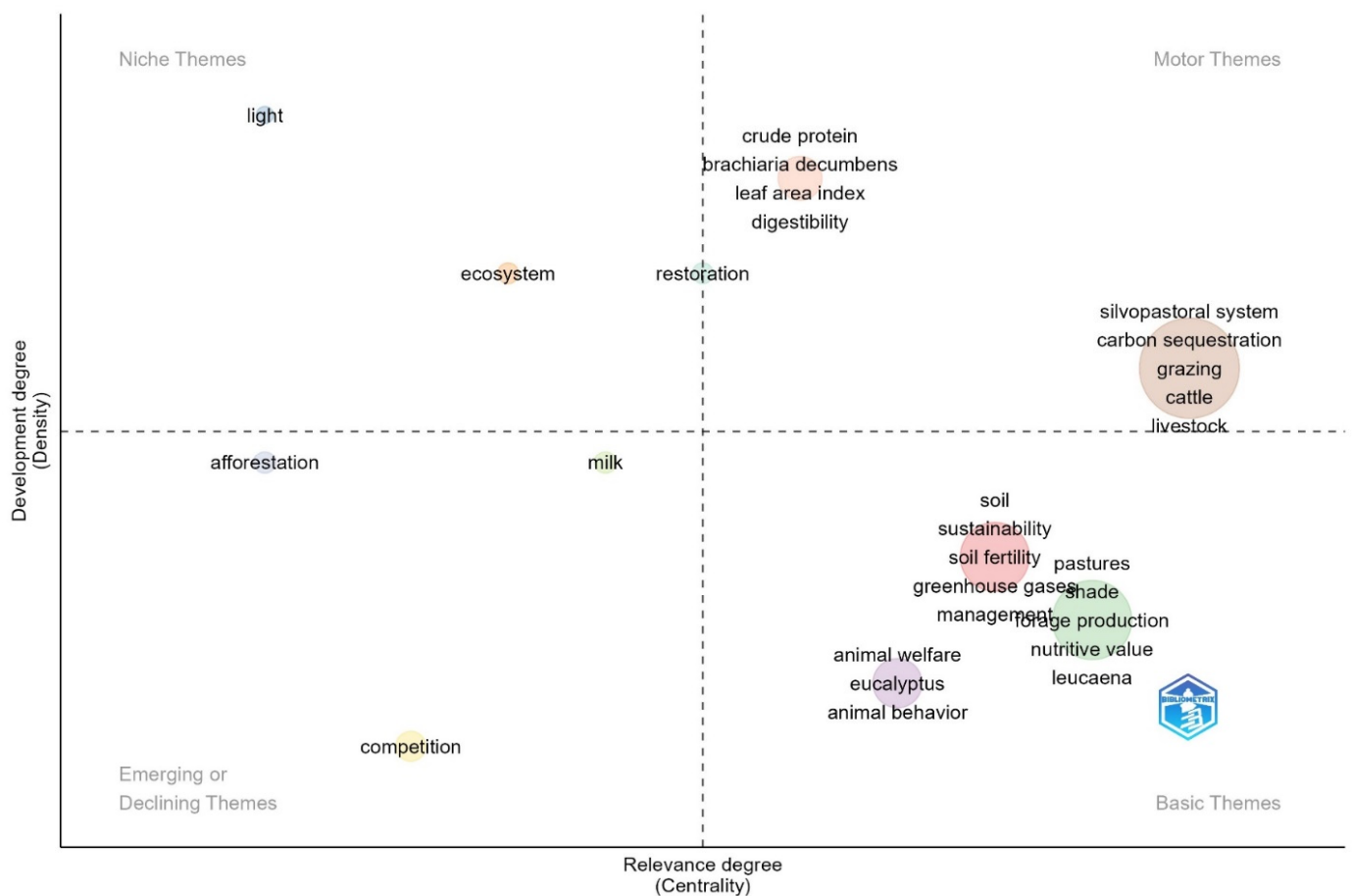


Figure 3. Thematic map showing clusters by the author's keywords.

The impact of silvopastoral systems management on forage production and nutritive value has been extensively studied [44–46], and the primary factor analyzed in these parameters has been shade and its effect on forage production and nutritive value [47–49]. Generally, the main impacts of excessive shade include increased in fiber content due to plant etiolation and enhanced lignification in the pursuit of vertical growth in competition for light [45,50]. However, benefits are observed in low to moderate shade, with forage production similar or superior to plants grown in full sun, presenting better nutritive value with enhanced crude protein content due to delayed maturity and reduced senescence rates [51,52].

Studies have reported that silvopastoral systems may enhance C sequestration in soil [53–55], which may be attributed to increased abundance of microbial species, improved soil nutrient cycling and stability, enhanced watershed function, more abundant biodiversity and wildlife habitat while simultaneously achieving higher levels of healthy food production [30]. Additionally, most tropical grass species use the C₄ photosynthetic pathway, resulting in higher rate of lignin deposition and reduced digestibility and voluntary feed intake, especially during periods of scarcity, such as dry seasons [34,56]. In silvopastoral systems, animals can use foliage, pods, and even fruits from trees or shrubs as feed complementation, helping overcome feed shortages in critical periods [10,57].

Moreover, pastures in such systems exhibit increased productivity. They can influence grazing area microclimate parameters, providing reduced temperature, and increased humidity, benefiting animal behavior, and allowing for extended grazing periods due to more favorable environmental conditions [58,59]. In a silvopastoral system with *Andropogon gayanus* grass pasture cultivated alongside native trees in the northeast region of Brazil, Zambrano et al. [60] observed that Anglo-Nubian goats dedicated more time to grazing than animals in an *A. gayanus* monocultural system. These authors also noted increased

forage biomass production and reduced environmental temperature in the silvopastoral system compared to the monoculture. Another benefit of silvopastoral system for animal production is the increased shade area in pastures due to the presence of trees, mitigating heat stress in grazing animals, especially in regions with hot climates, as frequently observed in tropical and subtropical countries of Latin America [48].

Using an in vitro fermentation system, Ovani et al. [61] assessed the inclusion of *Chloroleucon acacioides* tree fruits in tropical grass-based diet substrates. These authors observed greater estimated microbial biomass production and short-chain fatty acids synthesis, associated with increased organic matter degradability in treatments containing *C. acacioides* fruits compared to the control treatment consisting of 100% tropical grass hay. Furthermore, the authors highlighted the potential for incorporating this tree species, native to the Brazilian Amazon, into tropical grass pastures, as its fruits can serve as a nutrient source for animals during periods when forage quality and biomass production are reduced, such as in dry seasons.

A widespread beef cattle management in many parts of the world is to raise animals on a continuously grazing system, and then, for finishing stages, the animals are taken to feedlot systems and fed on grain-based diets so they can increase their weight faster and be ready for the market. Usually, this practice is associated with lower GHG emissions per unit of generated product since lower overall time is required from rearing to slaughter. However, this kind of statement does not consider all the emissions generated by feedlot operations, including those of grain crop production or machinery utilized in such management, a factor that usually underestimates GHG emissions from feedlot systems [30]. Undoubtedly, standard grain production practices can be changed and more regenerative, reducing its overall emissions and impacts. However, in addition to the C sequestration potential that well-managed grazing systems present, grazing is a natural behavior for cattle. Consequently, pastoral systems provide an opportunity for increased welfare conditions for these animals [29,62]. In addition to the ecosystem services that conventional grazing systems can provide [36], in silvopastoral, trees and shrubs by-products can be a source of phenolic compounds that interfere both positively or negatively with feed intake and digestibility [10], and several of these plants, especially those containing tannins, offer a range of benefits, including the increased flow of dietary amino acids to the small intestine, control of gastrointestinal nematodes infections, reduction of bacterial loads in feces, decreased occurrence of frothy bloat in animals consuming legume forages, and mitigation of enteric CH₄ production during ruminal fermentation, characterizing one of the most prominent benefits of silvopastoralism considering all the concerns about ruminant production in a climate change scenario [34,63–66]. Albores-Moreno et al. [67] used an in vitro system to evaluate the impacts of tree foliage consumed by cattle in Mexico on ruminal fermentation parameters. These authors reported reductions in CH₄ production of up to 31% when diets were supplemented with 300 g/kg DM of foliage in substrates in relation to the control treatment, containing only tropical grass forages; in this study, the authors highlighted that the presence of condensed tannins could be one of the possible explanations for such reductions.

Soltan et al. [68] fed Santa Inês sheep with a tropical-grass-based diet supplemented with *Leucaena leucocephala*, a tannin-containing legume tree/shrub used in silvopastoral systems in Latin America [69]; these authors observed reduced CH₄ emissions when compared to the control diet, with no supplementation. In addition to a direct reduction of CH₄ production in the rumen, dietary tannins may also lead to reduced emissions of another important GHG: some tannins may form complexes with dietary protein, increasing their flux and absorption in the small intestine and also shifting the excretion of nitrogen (arising out of these proteins) from urine to feces, which can be an advantage in terms of nitrous oxide (N₂O) emissions, since fecal nitrogen is in its organic form and less prone to volatilization, while urinary nitrogen is mostly urea, which can be easily converted into N₂O [70,71].

Silvopastoral systems using legume trees or shrubs can benefit from the ability of these plants to do biological atmospheric nitrogen fixation. This process leads to increased inputs of this element into the soil, reducing the need for nitrogen fertilization [72]. Nitrogen fertilization is a chief source of N₂O emissions from agriculture since microbial processes that nitrogen goes through in the soil, especially nitrification and denitrification, lead to the production of this gas. In addition to direct emissions from the application of fertilizers, it is also necessary to consider emissions deriving out of nitrogen that lixiviates from agricultural fields that may also lead to N₂O emissions [73]. Therefore, the presence of legumes in silvopastoral systems may provide this additional benefit concerning GHG emissions and production sustainability.

4. *Tithonia diversifolia* and Sustainable Ruminant Production

As previously observed in this review and highlighted by other authors [74–76], many tree and shrub species can be used in silvopastoral systems, including both cultivated and native plants. One such shrub species is *T. diversifolia*, known as titonia, botón del oro, Mexican or wild sunflower. Belonging to the Asteraceae family, *T. diversifolia* originates from Mexico but has now spread widely across the humid and sub-humid tropics in Central and South America, Asia, and Africa [77,78]. It typically grows between 1.5 to 4.0 m tall, presenting leaves with serrated edges and peduncles ranging from 5 to 20 cm long, with yellow inflorescence [79]. Among the forage options for tropical silvopastoral systems, *T. diversifolia* presented characteristics such as high CP compared to tropical grasses, good adaptability to harsh environmental conditions, high biomass production, and have led to increased volatile fatty acids production and microbial protein synthesis on in vitro trials, characteristics that made this plant stand out as a promising option for such systems [14,77,80,81].

In the African continent, a study was conducted to verify the biomass production of *T. diversifolia* under different pruning practices, with the idea of using residues from pruning as natural fertilizer for the soil [82]. The authors observed that adopting a cutting height of 50 cm above soil on a bi-monthly frequency could lead to an annual DM production as high as 7.2 t ha^{−1}, which makes evident how productive and effective this forage can be in providing available biomass for grazing ruminants. This author highlighted that productivity numbers may vary according to region.

Also, given the fast decomposition of *T. diversifolia* plant material and its ability to mobilize soil phosphorus, it is a good option for green fertilizer. The high productivity of this plant, combined with its adaptability to various environmental conditions, makes it easy to grow and spread. Consequently, it is often considered an invasive species in both agricultural and non-agricultural lands, being commonly observed in marginal areas along roads or crop fields, not requiring great soil fertility and demonstrating good tolerance to acidic soils and short periods of drought, the later due to its longer roots compared to grass forages, allowing it to explore deeper soil profiles in search for water and nutrients [38,83–85].

A significant number of studies in the literature using different *T. diversifolia* sources reported CP levels varying around 200 g/kg DM, illustrating the agronomic potential of *T. diversifolia* (Table 1).

Table 1. Nutritional composition of *Tithonia diversifolia* in different studies. Except for dry matter (DM), all values are presented as g/kg on DM basis.

References	DM	CP ¹	NDF	ADF	Obs.
Argüello-Rangel et al. [86]	190	252	337	145	Whole plant Leaves
Calsavara et al. [14]	200	165	476	333	
Calsavara et al. [14]	195	225	410	261	
Chin and Hue [87]	146	239	384	n/a	
Durango et al. [88]	212	185	462	343	
Guatusmal-Gelpud et al. [89]	n/a	267	331	150	

Table 1. Cont.

References	DM	CP ¹	NDF	ADF	Obs.
Lezcano et al. [90]	101	219	n/a	n/a	Rainy season
Lezcano et al. [90]	127	190	n/a	n/a	Dry season
Londoño et al. [91]	185	273	268	169	No fertilization
Mahecha and Rosales [79]	172	242	253	304	
Mahecha et al. [92]	n/a	223	359	181	
Naranjo and Cuartas [93]	191	241	386	345	
Van Sao et al. [77]	146	239	384	n/a	
Verdecia et al. [94]	198	289	436	276	Rainy season
Verdecia et al. [94]	182	275	404	241	Dry season

¹ CP—crude protein; NDF—neutral detergent fiber; ADF—acid detergent fiber; n/a—information not available on papers; Obs.—Observation, reflecting extra information on forage samples when made available by referred authors.

Krüger et al. [12] also observed CP levels around 200 g/kg DM during the dry season in southeastern Brazil. Moreover, Pérez-Márquez et al. [81], working with an in vitro fermentation system with inclusion levels of *T. diversifolia* on a 60:40 forage:concentrate ratio substrates, observed higher iso-valerate, iso-butyrate, as well as microbial biomass production in the first 24 h of incubation. Both iso-valerate and iso-butyrate are branched-chain fatty acids that originate from the degradation of branched-chain amino acids and rumen microbes utilize them for microbial protein synthesis [95]. Cellulolytic bacteria might benefit from using these fatty acids for their growth, potentially leading to increased fiber degradability [96]. In addition, increased branched-chain fatty acid production can be indicative of good protein degradability [34], which, if combined with the potential positive impact on microbial protein synthesis, makes it evident that *T. diversifolia* can be an excellent feeding resource to increase protein supply for animal in grazing tropical production systems, especially in the dry season. In this period, tropical grasses may show CP levels lower than 70 g/kg DM, which can be critical for the ruminal ecosystem [97,98]. Panadero and Montaña [38] also emphasized the potential of this plant for recovering degraded soil areas, a scenario often observed in the tropics.

In terms of CH₄ production, a review of the literature reveals how the inclusion of *T. diversifolia* affects this variable (Table 2). Despite the lack of a consistent pattern across studies, there are several examples where the inclusion of *T. diversifolia* has led to reduced CH₄ production. In most of these cases, authors attributed the reduction to a direct action on methanogenic microorganisms due to the presence of polyphenols in *T. diversifolia* (such as tannins) or to a reduction in the acetate:propionate (A:P) ratio [16,76,99,100]. The lack of consistency among studies demonstrates that the effect of *T. diversifolia* on CH₄ production is closely associated with the quality and type of substrate in which *T. diversifolia* is included. As observed by Akanmu et al. [100], the inclusion of *T. diversifolia* was more pronounced in fibrous substrates.

Despite Terry et al. [80] observing that the *T. diversifolia* inclusion led to increased CH₄ production, this increased methanogenesis was accompanied by higher acetate production, which can lead to improved animal performance in production systems. Elevated acetate production can be essential, especially to dairy production systems, as this fatty acid is an important precursor of milk fat, which in turn is an indicator of milk quality [101,102], allowing farmers to potentially have additional incomes from their product. On the other hand, Rivera et al. [103] observed that when including around 150 g/kg (fresh material) of *T. diversifolia* in a grass-based diet of cows, the presence of this shrub reduced their CH₄ emissions when expressed on a daily basis, per unit of DM intake, and per unit of degraded DM intake as well. The authors attributed this reduction to several factors, such as the decreased fiber content of *T. diversifolia*, accompanied by its increased CP, digestibility, and the presence of plant secondary compounds, reinforcing the multiple positive aspects of this plant as a feeding resource for ruminants.

Table 2. In vitro methane (CH₄) production, fiber content, and results found in the literature of *Tithonia diversifolia* (TD) in association with different substrates.

Treatments	CH ₄ Production		Unit	NDF (%)	ADF (%)	A:P Ratio		Authors' Discussion	Reference
	TRT ¹	CON				TRT	CON		
10% TD with <i>Lolium perenne</i>	29.3	30.5	mL/gDM	53.8	27.1	2.00	2.06	No differences in CH ₄ and A:P ratio. Decreased CH ₄ due to the presence of tannins.	[103]
20% TD with <i>Lolium perenne</i>	25.9			52.1	28.9	1.95			
33% TD with <i>Pennisetum purpureum</i>	1.5	2.4	mmol/g	69.1	52.1	2.16	2.06	No differences in CH ₄ and A:P ratio.	[104]
75% TD with <i>Pennisetum purpureum</i>	8.6	18.9	mL/gDOM	59.1	49.6	3.27	2.63	Decreased CH ₄ . Similar acetate and decreased propionate.	[105]
75% TD with <i>Cynodon dactylon</i>	4	7.3	mL/gDOM	61.3	48.3	3.41	3.04	No differences in CH ₄ . Similar propionate and increased acetate.	
TD extract with Commercial Concentrate (TMR)	25.3	42.9	mL/kgDOM	30.1	21.4	1.39	1.99	Decreased CH ₄ due to the presence of tannins. Similar propionate and decreased acetate.	[100]
TD extract with lucerne hay	18.2	36.8		40.6	32.1	1.71	2.16	Decreased CH ₄ due to the presence of tannins. Similar propionate and decreased acetate.	
TD extract with <i>Eragostis curvula</i>	5.8	47.7		78.4	49.2	1.63	2.48	Decreased CH ₄ due to the presence of tannins. Increased propionate and decreased acetate.	
6.9% TD with sugarcane and concentrate	0.7	0.5	mL/gIDM	29.4	-	0.90	0.71	TD inclusion produced more CH ₄ due to increased A:P ratio. Increased acetate and decreased propionate.	[80]
15.2% TD with sugarcane and concentrate	1.2			30.7	-	1.09			
29.2% TD with sugarcane and concentrate	3.3			34.5	-	1.55			
25% TD with <i>Urochloa brizantha</i>	~22.9	26.2	mg/gIDM	~55.5	~38.3	2.37	3.56	Decreased CH ₄ due to decreased A:P ratio. Decreased acetate and increased propionate.	[76]
30% TD with <i>Cynodon nlemfuensis</i>	0.9	6.5	mL/100 mL	-	-	-	-	Decreased CH ₄ due to the presence of tannins.	[16]
30% TD with <i>Cynodon nlemfuensis</i>	9.2	65.2	uL/gDM	33.4	29.5	-	-	Decreased CH ₄ due to the presence of tannins.	[106]
30% de TD with <i>Cynodon nlemfuensis</i>	47.2			35.3	30.4	-	-	Decreased CH ₄ .	
100% TD	15.7	43.4	mL/gDDM	39	27.2	-	-	Decreased CH ₄ due to the presence of tannins.	[99]
5% TD with <i>Cechrus clandestinum</i>	34.8	43.4		-	-	4.01	4.52	Decreased CH ₄ due to the presence of tannins. Decreased acetate and increased propionate.	
3% TD with <i>Cechrus clandestinum</i> , concentrate and fat	41	60.3		-	-	3.97	4.80	Decreased CH ₄ due to the presence of tannins. Similar acetate and increased propionate.	

¹ TRT—Treatment group; CON—Controls; NDF—neutral detergent fiber; ADF—acid detergent fiber; A:P—acetate:propionate ratio; DM—dry matter; IDM—incubated dry matter; DDM—degraded dry matter; DOM—degraded organic matter.

Additionally, *T. diversifolia* is a source of a wide range of secondary compounds [15]. In a study to characterize the phytochemical composition of this forage, Olayinka et al. [107] prepared aqueous and ethanol extracts with stems, leaves, and root of this plant. In both cases, extracts tested positive for alkaloids, flavonoids, saponins, terpenoids, tannins, and other phenolic compounds. Tagne et al. [15] analyzed more than 160 scientific articles, and identified more than 100 secondary metabolites isolated from different *T. diversifolia* extracts. Thanks to that extensive diversity of compounds, several properties, activities, and

effects of interest for human medicine, such as anti-inflammatory activity, anti-protozoal effect, repellent against insects, antidiabetic effect, antibacterial and antifungal activities, antiviral, antioxidant, antiproliferative (i.e., against cancer cells), and even effects against gastrointestinal disorders, have been listed by the authors and attributed to the use of this plant.

For ruminant nutrition, a group of secondary compounds that for decades has been eliciting interest from the scientific community are the tannins, due to the beneficial effects of these extensively studied molecules [34,71,108] for the metabolism of ruminants as described in the previous section of this paper. Concerning the tannins found in *T. diversifolia*, Delgado et al. [16] reported moderate concentrations of these compounds. They observed reduced CH₄ concentrations in total in vitro gas production, along with a decreased protozoa population compared to other plants tested in their experiment. Such effects were attributed to the presence of tannins in *T. diversifolia*. Additionally, other authors in different studies who observed the reduction of CH₄ production due to the inclusion of *T. diversifolia* also pointed out that these results were due to the presence of tannins (Table 2). However, it is consolidated that the gold standard method to evaluate the biological effects of a certain tannin source on the metabolism of ruminants is doing in vitro or even in vivo trials using a tannin-neutralizing agent such as polyethylene glycol [109,110]. Therefore, studies using *T. diversifolia* with this experimental design are still warranted in order to provide a more accurate understanding about the tannins of this plant.

Nitrous oxide (N₂O) emissions data from operations using *T. diversifolia* in the diet of ruminants are still scarce in the literature. However, several researchers have assessed the impact of *T. diversifolia* inclusion on the animal's nitrogen balance (Table 3), which directly impacts the amount of N excreted and the subsequent conversion of N into N₂O since N balance and N excretion means (i.e., urine or feces) have significant influence on the potential of N₂O emissions from soils [70,71].

As observed for CH₄ production, it seems evident that the plant's influence on N balance is also dependent on associated diets' characteristics (Table 3). Associations with fibrous diets show more pronounced results than those with concentrated ones, as noted by Yousuf et al. [111], Ribeiro et al. [11], and Chacón Góngora [112], who used *T. diversifolia* in concentrated diets and found no significant differences in N retention compared to diets without *T. diversifolia*, while Ramírez-Rivera et al. [113], Castañeda Serrano et al. [114], Fajemisin et al. [115], and Durango et al. [88], associating *T. diversifolia* with exclusively forage diets, reported an increase in N retention compared to diets without *T. diversifolia*. Recently, Rivera et al. [116] evaluated soil N₂O emissions from grazing sites using cross-bred dairy cows in the Colombian Amazon by employing static closed chambers and reported that the silvopastoral system using *T. diversifolia* have led to lower N₂O emissions than the conventional grazing systems, which was composed by partially degraded *Brachiaria humidicola* areas. Therefore, silvopastoral systems with *T. diversifolia* arise as a sustainable production alternative for pasture-based systems, playing a crucial role in minimizing environmental footprint and promoting ecosystem services, which are becoming progressively vital and sought after in the current landscape of climate change.

Table 3. Nitrogen (N) balance of diets including *Tithonia diversifolia* (TD) in the literature.

Treatments	DMI ¹ g/Day	NI g/Day	NF % NI	NU % NI	NR % NI	Authors' Discussion	Reference
0% TD extract + Cassava + concentrate	378	6.98	39	39.8	21.2	80% TD resulted in decreased fecal N excretion and higher urinary N excretion. N retention was similar in all treatments except at 80%.	[111]
20% TD extract + Cassava + concentrate	374	6.92	31.4	46.7	22		
40% TD extract + Cassava + concentrate	371	6.87	29.7	46.4	23.9		
80% TD extract + Cassava + concentrate	318	5.88	25.9	56.3	17.9		
0% TD + <i>Dichanthium aristatum</i>	410	9.78	24.9	12.9	62.1	TD inclusion resulted in decreased N excretion in feces and urine. N retention was higher with TD inclusion.	[114]
25% TD + <i>Dichanthium aristatum</i>	704	68.93	6.8	7.3	85.9		

Table 3. Cont.

Treatments	DMI ¹ g/Day	NI g/Day	NF % NI	NU % NI	NR % NI	Authors' Discussion	Reference
0% TD + <i>Brachiaria decumbens</i>	652	63.23	48	35	16	TD inclusion resulted in higher N excretion in feces and urine. N retention was higher with TD inclusion.	[88]
35% TD + <i>Brachiaria decumbens</i>	840	113.48	34	26	39		
0% TD + <i>Panicum maximum</i>	312	7.59	31.5	14.5	54	No difference in fecal N excretion. Inclusions of 20% and 30% resulted in decreased urine N excretion. N retention was reduced by 20% and 30% TD diets.	[117]
10% TD + <i>Panicum maximum</i>	311	6.84	31.9	14.8	53.4		
20% TD + <i>Panicum maximum</i>	306	5.83	37	11.3	51.6		
30% TD + <i>Panicum maximum</i>	305	5.76	42	6.3	51.7		
TD 0% + sugarcane + concentrate	1860	563	34.5	11	54.6	No significant effects of TD inclusion on N excretion and nitrogen balance.	[11]
TD 6.5% + sugarcane + concentrate	1890	564.1	35.6	10	54.4		
TD 15.4% + sugarcane + concentrate	1870	557.2	35.5	11.4	53.1		
0% TD + <i>Pennisetum purpureum</i> + sugarcane	1050	13.05	50.73	39.2	9	TD inclusion increased N excretion in feces and urine. Only at 20% inclusion was there positive N retention. Other inclusions were not significant.	[113]
20% TD + <i>Pennisetum purpureum</i> + sugarcane	1510	19.93	47.22	30	23.3		
35% TD + <i>Pennisetum purpureum</i> + sugarcane	1550	25.51	47.75	34.1	18.7		
50% TD + <i>Pennisetum purpureum</i> + sugarcane	1520	30.11	47.23	37.6	15.6		
0% TD + <i>Brachiaria</i> + concentrate	1471	66.4	25.1	55.4	19.5	No effect of TD inclusion on fecal N excretion. Urinary N excretion was Decreased with 12% TD inclusion.	[112]
6% TD + <i>Brachiaria</i> + concentrate	1432	66.4	24.9	54.4	20.7		
12% TD + <i>Brachiaria</i> + concentrate	1452	66	27.1	43.7	29.2		
0% TD + <i>Panicum maximum</i>	1580	30.5	-	-	76	TD inclusions led to increased N retention.	[115]
25% TD + <i>Panicum maximum</i>	1970	56.6	-	-	71.9		
50% TD + <i>Panicum maximum</i>	2070	64.4	-	-	75		
75% TD + <i>Panicum maximum</i>	2130	72.8	-	-	76.7		

¹ DMI—dry matter intake; NI—N intake; NF—N in feces; NU—N in urine; NR—N retention; For easier comparison across studies, N values in feces, urine, and retained were expressed as a percentage of N intake.

5. Conclusions

With the mounting pressure from a scenario marked by a growing human population, higher demand for animal-derived food production, and increasing concerns about climate change, sustainable food production seems an inevitable requirement for humanity in the next few years. Tropical grasslands, abundant in Latin America, could be crucial in addressing these challenges. They offer rich biodiversity and have management practices that can improve food production, particularly protein, while reducing adverse impacts and promoting sustainability of the production system. Silvopastoral systems, while not yet widely adopted, seem to be one of the most promising practices, as evidenced by the literature gathered in this review, showing that these systems can preserve and recover natural resources such as soil and groundwater, while providing benefits to animals such as abundant nutrient sources, improved welfare, and offering cultural and ecosystem services for communities and populations reliant on these systems. Our research also showed that *T. diversifolia* is a shrub excellently suited for tropical silvopastoral systems, boasting significant potential for exploration. Its high-quality nutritional composition, agronomic adaptability to various tropical conditions, and potential to enhance animal performance while reducing GHG emissions intensity all underscore its importance. The information compiled in our review makes it clear that silvopastoral systems, as well as *T. diversifolia* should be mandatory topics in future discussions on sustainable ruminant production grazing systems in tropical environments. However, more thorough studies are still warranted to accurately characterize its impacts on animal performance and metabolism, while the scientific community should dedicate especial attention to its secondary bioactive metabolites and their direct impacts on GHG production, information not well enough detailed and clarified in the literature, making it an area of expertise to be further explored.

Author Contributions: A.M.K.—conceptualization, literature research, and writing of the original draft; P.d.M.T.L., V.O., S.P.-M. and H.L.—literature research, manuscript review, and editing; A.L.A., conceptualization, supervision, manuscript review, and editing. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES—Finance code 001), and Fundação de Amparo à Pesquisa do Estado de São Paulo (grant no. 2016/26035-3).

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. Boval, M.; Dixon, R.M. The Importance of Grasslands for Animal Production and Other Functions: A Review on Management and Methodological Progress in the Tropics. *Animal* **2012**, *6*, 748–762. [\[CrossRef\]](#) [\[PubMed\]](#)
2. Flachowsky, G.; Meyer, U.; Südekum, K.-H. Land Use for Edible Protein of Animal Origin—A Review. *Animals* **2017**, *7*, 25. [\[CrossRef\]](#) [\[PubMed\]](#)
3. Gerber, P.; Steinfeld, H.; Henderson, B.; Mottet, A.; Opio, C.; Dijkman, J.; Falcucci, A.; Tempio, G. *Tackling Climate Change through Livestock—A Global Assessment of Emissions and Mitigation Opportunities*; Food and Agriculture Organization of the United Nations—FAO: Rome, Italy, 2013.
4. Benchaar, C.; Greathead, H. Essential Oils and Opportunities to Mitigate Enteric Methane Emissions from Ruminants. *Anim. Feed Sci. Technol.* **2011**, *166*–*167*, 338–355. [\[CrossRef\]](#)
5. Belanche, A.; Hristov, A.N.; van Lingen, H.J.; Denman, S.E.; Kebreab, E.; Schwarm, A.; Kreuzer, M.; Niu, M.; Eugène, M.; Niderkorn, V.; et al. Prediction of Enteric Methane Emissions by Sheep Using an Intercontinental Database. *J. Clean. Prod.* **2023**, *384*, 135523. [\[CrossRef\]](#)
6. Eisler, M.C.; Lee, M.R.F.; Tarlton, J.F.; Martin, G.B.; Beddington, J.; Dungait, J.A.J.; Greathead, H.; Liu, J.; Mathew, S.; Miller, H.; et al. Agriculture: Steps to Sustainable Livestock. *Nature* **2014**, *507*, 32–34. [\[CrossRef\]](#) [\[PubMed\]](#)
7. Habermann, E.; Dias de Oliveira, E.A.; Contin, D.R.; Delvecchio, G.; Viciado, D.O.; de Moraes, M.A.; de Mello Prado, R.; de Pinho Costa, K.A.; Braga, M.R.; Martinez, C.A. Warming and Water Deficit Impact Leaf Photosynthesis and Decrease Forage Quality and Digestibility of a C4 Tropical Grass. *Physiol. Plant.* **2019**, *165*, 383–402. [\[CrossRef\]](#) [\[PubMed\]](#)
8. Pirhofer-Walzl, K.; Rasmussen, J.; Høgh-Jensen, H.; Eriksen, J.; Sørensen, K.; Rasmussen, J. Nitrogen Transfer from Forage Legumes to Nine Neighbouring Plants in a Multi-Species Grassland. *Plant Soil* **2012**, *350*, 71–84. [\[CrossRef\]](#)
9. Allen, V.G.; Batello, C.; Berretta, E.J.; Hodgson, J.; Kothmann, M.; Li, X.; McIvor, J.; Milne, J.; Morris, C.; Peeters, A.; et al. An International Terminology for Grazing Lands and Grazing Animals. *Grass Forage Sci.* **2011**, *66*, 2–28. [\[CrossRef\]](#)
10. Jose, S.; Dollinger, J. Silvopasture: A Sustainable Livestock Production System. *Agrofor. Syst.* **2019**, *93*, 1–9. [\[CrossRef\]](#)
11. Ribeiro, R.S.; Terry, S.A.; Sacramento, J.P.; Silveira, S.R.E.; Bento, C.B.P.; da Silva, E.F.; Mantovani, H.C.; Gama, M.A.S.D.; Pereira, L.G.R.; Tomich, T.R.; et al. Tithonia Diversifolia as a Supplementary Feed for Dairy Cows. *PLoS ONE* **2016**, *11*, e0165751. [\[CrossRef\]](#)
12. Krüger, A.; Márquez, S.; Bizzuti, B.; Ovani, V.; Takahashi, L.; Santos, A.; Lima, P.; Maurício, R.; Abdalla, A. Tithonia Diversifolia and Tropical Grasses Intercropping as a Sustainable Alternative for Ruminants. In Proceedings of the International Symposium on Sustainable Animal Production and Health, Vienna, Austria, 28 June–2 July 2021.
13. Nziguheba, G.; Merckx, R.; Palm, C.A. Mutuo Combining Tithonia Diversifolia and Fertilizers for Maize Production in a Phosphorus Deficient Soil in Kenya. *Agrofor. Syst.* **2002**, *55*, 165–174. [\[CrossRef\]](#)
14. Calsavara, L.H.F.; Ribeiro, R.S.; Silveira, S.R.; Delarota, G.; Freitas, D.S.; Sacramento, J.P.; Paciullo, D.S.C.; Maurício, R.M. Potential of Tithonia Diversifolia as Source of Forage for Ruminants. *Livest. Res. Rural Dev.* **2016**, *28*, 1–9.
15. Mabou Tagne, A.; Marino, F.; Cosentino, M. *Tithonia diversifolia* (Hemsl.) A. Gray as a Medicinal Plant: A Comprehensive Review of Its Ethnopharmacology, Phytochemistry, Pharmacotoxicology and Clinical Relevance. *J. Ethnopharmacol.* **2018**, *220*, 94–116. [\[CrossRef\]](#) [\[PubMed\]](#)
16. Delgado, D.C.; Galindo, J.; González, R.; González, N.; Scull, I.; Dihigo, L.; Cairo, J.; Aldama, A.I.; Moreira, O. Feeding of Tropical Trees and Shrub Foliages as a Strategy to Reduce Ruminal Methanogenesis: Studies Conducted in Cuba. *Trop. Anim. Health Prod.* **2012**, *44*, 1097–1104. [\[CrossRef\]](#) [\[PubMed\]](#)
17. Rock, C.L.; Flatt, S.W.; Pakiz, B.; Taylor, K.S.; Leone, A.F.; Brelje, K.; Heath, D.D.; Quintana, E.L.; Sherwood, N.E. Weight Loss, Glycemic Control, and Cardiovascular Disease Risk Factors in Response to Differential Diet Composition in a Weight Loss Program in Type 2 Diabetes: A Randomized Controlled Trial. *Diabetes Care* **2014**, *37*, 1573–1580. [\[CrossRef\]](#)
18. Mottet, A.; de Haan, C.; Falcucci, A.; Tempio, G.; Opio, C.; Gerber, P. Livestock: On Our Plates or Eating at Our Table? A New Analysis of the Feed/Food Debate. *Glob. Food Sec.* **2017**, *14*, 1–8. [\[CrossRef\]](#)
19. Matthews, C.; Crispie, F.; Lewis, E.; Reid, M.; O'Toole, P.W.; Cotter, P.D. The Rumen Microbiome: A Crucial Consideration When Optimising Milk and Meat Production and Nitrogen Utilisation Efficiency. *Gut Microbes* **2019**, *10*, 115–132. [\[CrossRef\]](#)

20. McLaren, S.; Berardy, A.; Henderson, A.; Holden, N.; Huppertz, T.; Jolliet, O.; De Camillis, C.; Renouf, M.; Rugani, B.; Saarinen, M.; et al. *Integration of Environment and Nutrition in Life Cycle Assessment of Food Items: Opportunities and Challenges*; Food and Agriculture Organization of the United Nations–FAO: Rome, Italy, 2021; ISBN 978-92-5-135532-9.
21. Beauchemin, K.A.; Ungerfeld, E.M.; Eckard, R.J.; Wang, M. Review: Fifty Years of Research on Rumen Methanogenesis: Lessons Learned and Future Challenges for Mitigation. *Animal* **2020**, *14*, s2–s16. [\[CrossRef\]](#) [\[PubMed\]](#)
22. Wyness, L. The Role of Red Meat in the Diet: Nutrition and Health Benefits. *Proc. Nutr. Soc.* **2016**, *75*, 227–232. [\[CrossRef\]](#)
23. Givens, D.I. Review: Dairy Foods, Red Meat and Processed Meat in the Diet: Implications for Health at Key Life Stages. *Animal* **2018**, *12*, 1709–1721. [\[CrossRef\]](#)
24. Cerri, C.C.; Moreira, C.S.; Alves, P.A.; Raucci, G.S.; de Almeida Castigioni, B.; Mello, F.F.C.; Cerri, D.G.P.; Cerri, C.E.P. Assessing the Carbon Footprint of Beef Cattle in Brazil: A Case Study with 22 Farms in the State of Mato Grosso. *J. Clean. Prod.* **2016**, *112*, 2593–2600. [\[CrossRef\]](#)
25. McAuliffe, G.A.; Takahashi, T.; Orr, R.J.; Harris, P.; Lee, M.R.F. Distributions of Emissions Intensity for Individual Beef Cattle Reared on Pasture-Based Production Systems. *J. Clean. Prod.* **2018**, *171*, 1672–1680. [\[CrossRef\]](#)
26. Sakita, G.Z.; Lima, P.M.T.; Abdalla Filho, A.L.; Bompadre, T.F.V.; Ovani, V.S.; Bizzuti, B.E.; da Costa, W.D.S.; do Prado Paim, T.; Campioni, T.S.; de Oliva Neto, P.; et al. Treating Tropical Grass with Fibrolytic Enzymes from the Fungus *Trichoderma Reesei*: Effects on Animal Performance, Digestibility and Enteric Methane Emissions of Growing Lambs. *Anim. Feed Sci. Technol.* **2022**, *286*, 115253. [\[CrossRef\]](#)
27. Loeb, J. COP26: Is Change Afoot in Livestock Farming? *Vet. Rec.* **2021**, *189*, 348–351. [\[CrossRef\]](#)
28. Eisen, M.B.; Brown, P.O. Rapid Global Phaseout of Animal Agriculture Has the Potential to Stabilize Greenhouse Gas Levels for 30 Years and Offset 68 Percent of CO₂ Emissions This Century. *PLoS Clim.* **2022**, *1*, e0000010. [\[CrossRef\]](#)
29. de Oliveira Silva, R.; Barioni, L.G.; Hall, J.A.J.; Folegatti Matsuura, M.; Zanett Albertini, T.; Fernandes, F.A.; Moran, D. Increasing Beef Production Could Lower Greenhouse Gas Emissions in Brazil If Decoupled from Deforestation. *Nat. Clim. Chang.* **2016**, *6*, 493–497. [\[CrossRef\]](#)
30. Teague, W.R.; Apfelbaum, S.; Lal, R.; Kreuter, U.P.; Rowntree, J.; Davies, C.A.; Conser, R.; Rasmussen, M.; Hatfield, J.; Wang, T.; et al. The Role of Ruminants in Reducing Agriculture’s Carbon Footprint in North America. *J. Soil Water Conserv.* **2016**, *71*, 156–164. [\[CrossRef\]](#)
31. Boval, M.; Angeon, V.; Rudel, T. Tropical Grasslands: A Pivotal Place for a More Multi-Functional Agriculture. *Ambio* **2017**, *46*, 48–56. [\[CrossRef\]](#) [\[PubMed\]](#)
32. Bond, W.J.; Parr, C.L. Beyond the Forest Edge: Ecology, Diversity and Conservation of the Grassy Biomes. *Biol. Conserv.* **2010**, *143*, 2395–2404. [\[CrossRef\]](#)
33. Bezabih, M.; Pellikaan, W.F.; Tolera, A.; Khan, N.A.; Hendriks, W.H. Chemical Composition and in Vitro Total Gas and Methane Production of Forage Species from the Mid Rift Valley Grasslands of Ethiopia. *Grass Forage Sci.* **2014**, *69*, 635–643. [\[CrossRef\]](#)
34. Lima, P.M.T.; Moreira, G.D.; Sakita, G.Z.; Natel, A.S.; Mattos, W.T.; Gimenes, F.M.A.; Gerdes, L.; McManus, C.; Abdalla, A.L.; Louvandini, H. Nutritional Evaluation of the Legume Macrotyloma Axillare Using In Vitro and in Vivo Bioassays in Sheep. *J. Anim. Physiol. Anim. Nutr.* **2018**, *102*, e669–e676. [\[CrossRef\]](#) [\[PubMed\]](#)
35. Rao, I.M.; Peters, M.; Castro, A.; Schultze-Kraft, R.; White, D.; Fisher, M.; Miles, J.W.; Lascano Aguilar, C.E.; Blümmel, M.; Bungenstab, D.J.; et al. LivestockPlus—The Sustainable Intensification of Forage-Based Agricultural Systems to Improve Livelihoods and Ecosystem Services in the Tropics. *Trop. Grassl. Forrajes Trop.* **2015**, *3*, 59. [\[CrossRef\]](#)
36. Teutscheroová, N.; Vázquez, E.; Sotelo, M.; Villegas, D.; Velásquez, N.; Baquero, D.; Pulleman, M.; Arango, J. Intensive Short-Duration Rotational Grazing Is Associated with Improved Soil Quality within One Year after Establishment in Colombia. *Appl. Soil Ecol.* **2021**, *159*, 103835. [\[CrossRef\]](#)
37. Rao, I.; Miles, J.; Wenzl, J.; Lowl-Gaume, A.; Cardoso, J.; Polania, J.; Rincón, J.; Hoyos, V.; Frossard, E.; Wagatsuma, T.; et al. Mechanisms of Adaptation of Brachiaria Grasses to Abiotic Stress Factors in the Tropics. In Proceedings of the III International Symposium on Forage Breeding, Bonito, Brazil, 7–11 November 2011.
38. Navas Panadero, A.; Montaña, V. Comportamiento de *Tithonia Diversifolia*, Bajo Condiciones de Bosque Húmedo Tropical. *Rev. Investig. Vet. del Perú* **2019**, *30*, 721–732. [\[CrossRef\]](#)
39. Page, M.J.; McKenzie, J.E.; Bossuyt, P.M.; Boutron, I.; Hoffmann, T.C.; Mulrow, C.D.; Shamseer, L.; Tetzlaff, J.M.; Akl, E.A.; Brennan, S.E.; et al. The PRISMA 2020 Statement: An Updated Guideline for Reporting Systematic Reviews. *BMJ* **2021**, *372*, n71. [\[CrossRef\]](#)
40. Aria, M.; Cuccurullo, C. Bibliometrix: An R-Tool for Comprehensive Science Mapping Analysis. *J. Informetr.* **2017**, *11*, 959–975. [\[CrossRef\]](#)
41. R Core Team R: A Language and Environment for Statistical Computing. Available online: <https://www.r-project.org/> (accessed on 15 February 2024).
42. FAOSTAT. Land Use and Agricultural Area. Available online: <https://www.fao.org/faostat/en/#country> (accessed on 14 February 2024).
43. Aria, M.; Alterisio, A.; Scandurra, A.; Pinelli, C.; D’Aniello, B. The Scholar’s Best Friend: Research Trends in Dog Cognitive and Behavioral Studies. *Anim. Cogn.* **2021**, *24*, 541–553. [\[CrossRef\]](#) [\[PubMed\]](#)
44. Sousa, L.F.; Maurício, R.M.; Moreira, G.R.; Gonçalves, L.C.; Borges, I.; Pereira, L.G.R. Nutritional Evaluation of “Braquiaraão” Grass in Association with “Aroeira” Trees in a Silvopastoral System. *Agrofor. Syst.* **2010**, *79*, 189–199. [\[CrossRef\]](#)

45. de Castro Santos, D.; Júnior, R.G.; Vilela, L.; Pulrolnik, K.; Bufon, V.B.; de Souza França, A.F. Forage Dry Mass Accumulation and Structural Characteristics of Piaã Grass in Silvopastoral Systems in the Brazilian Savannah. *Agric. Ecosyst. Environ.* **2016**, *233*, 16–24. [\[CrossRef\]](#)
46. Gomes, F.J.; Cavalli, J.; Pedreira, B.C.; Pedreira, C.G.S.; Holschuch, S.G.; Pereira, D.H. Forage Nutritive Value of Marandu Palisade Grass under Clipping in a Silvopastoral System. *Agrofor. Syst.* **2022**, *96*, 79–88. [\[CrossRef\]](#)
47. Paciullo, D.S.C.; De Carvalho, C.A.B.; Aroeira, L.J.M.; Morenz, M.J.F.; Lopes, F.C.F.; Rossiello, R.O.P. Morphophysiology and Nutritive Value of Signalgrass under Natural Shading and Full Sunlight. *Pesqui. Agropecuária Bras.* **2007**, *42*, 573–579. [\[CrossRef\]](#)
48. Ainsworth, J.A.W.; Moe, S.R.; Skarpe, C. Pasture Shade and Farm Management Effects on Cow Productivity in the Tropics. *Agric. Ecosyst. Environ.* **2012**, *155*, 105–110. [\[CrossRef\]](#)
49. Casanova-Lugo, F.; Villanueva-López, G.; Alcudia-Aguilar, A.; Nahed-Toral, J.; Medrano-Pérez, O.R.; Jiménez-Ferrer, G.; Alayón-Gamboa, J.A.; Aryal, D.R. Effect of Tree Shade on the Yield of Brachiaria Brizantha Grass in Tropical Livestock Production Systems in Mexico. *Rangel. Ecol. Manag.* **2022**, *80*, 31–38. [\[CrossRef\]](#)
50. Paciullo, D.S.C.; Campos, N.R.; Gomide, C.A.M.; Castro, C.R.T.D.; Tavela, R.C.; Rossiello, R.O.P. Crescimento de Capim-Braquiária Influenciado Pelo Grau de Sombreamento e Pela Estação Do Ano. *Pesqui. Agropecuária Bras.* **2008**, *43*, 917–923. [\[CrossRef\]](#)
51. Abraham, E.M.; Kyriazopoulos, A.P.; Parissi, Z.M.; Kostopoulou, P.; Karatassiou, M.; Anjalandidou, K.; Katsouta, C. Growth, Dry Matter Production, Phenotypic Plasticity, and Nutritive Value of Three Natural Populations of Dactylis Glomerata L. under Various Shading Treatments. *Agrofor. Syst.* **2014**, *88*, 287–299. [\[CrossRef\]](#)
52. Gomes, F.J.; Pedreira, B.C.; Santos, P.M.; Bosi, C.; Lulu, J.; Pedreira, C.G.S. Microclimate Effects on Canopy Characteristics of Shaded Palisadegrass Pastures in a Silvopastoral System in the Amazon Biome of Central Brazil. *Eur. J. Agron.* **2020**, *115*, 126029. [\[CrossRef\]](#)
53. Resende, L.D.O.; Müller, M.D.; Kohmann, M.M.; Pinto, L.F.G.; Cullen Junior, L.; de Zen, S.; Rego, L.F.G. Silvopastoral Management of Beef Cattle Production for Neutralizing the Environmental Impact of Enteric Methane Emission. *Agrofor. Syst.* **2020**, *94*, 893–903. [\[CrossRef\]](#)
54. Filho, J.F.L.; de Oliveira, H.M.R.; de Souza Barros, V.M.; dos Santos, A.C.; de Oliveira, T.S. From Forest to Pastures and Silvopastoral Systems: Soil Carbon and Nitrogen Stocks Changes in Northeast Amazônia. *Sci. Total Environ.* **2024**, *908*, 168251. [\[CrossRef\]](#) [\[PubMed\]](#)
55. Suárez, J.C.; Segura, M.; Andrade, H.J. Agroforestry Systems Affect Soil Organic Carbon Stocks and Fractions in Deforested Landscapes of Amazonia. *Agrofor. Syst.* **2024**, *98*, 1139–1151. [\[CrossRef\]](#)
56. Archimède, H.; Eugène, M.; Marie Magdeleine, C.; Boval, M.; Martin, C.; Morgavi, D.P.; Lecomte, P.; Doreau, M. Comparison of Methane Production between C3 and C4 Grasses and Legumes. *Anim. Feed Sci. Technol.* **2011**, *166–167*, 59–64. [\[CrossRef\]](#)
57. Melesse, A.; Steingass, H.; Schollenberger, M.; Holstein, J.; Rodehutsord, M. Nutrient Compositions and in Vitro Methane Production Profiles of Leaves and Whole Pods of Twelve Tropical Multipurpose Tree Species Cultivated in Ethiopia. *Agrofor. Syst.* **2019**, *93*, 135–147. [\[CrossRef\]](#)
58. Bahamonde, H.A.; Peri, P.L.; Alvarez, R.; Barneix, A.; Moretto, A.; Martínez Pastur, G. Litter Decomposition and Nutrients Dynamics in Nothofagus Antarctica Forests under Silvopastoral Use in Southern Patagonia. *Agrofor. Syst.* **2012**, *84*, 345–360. [\[CrossRef\]](#)
59. Sousa, L.F.; Maurício, R.M.; Paciullo, D.S.C.; Silveira, S.R.; Ribeiro, R.S.; Calsavara, L.H.; Moreira, G.R. Forage Intake, Feeding Behavior and Bio-Climatological Indices of Pasture Grass, under the Influence of Trees, in a Silvopastoral System. *Trop. Grassl. Forrajes Trop.* **2015**, *3*, 129. [\[CrossRef\]](#)
60. Zambrano, E.R.A.; Edvan, R.L.; Oliveira, M.E.; Da Costa Araujo, D.L.; Costa, J.V.; Da Silva, I.R.; Barros, D.M.A.; De Sousa Reis, G.; Dias-Silva, T.P. Characterization of Pasture of Andropogon Grass and Behavior of Grazing Goats in a Silvopastoral System. *Agrofor. Syst.* **2021**, *95*, 1155–1165. [\[CrossRef\]](#)
61. Ovani, V.S.; Pérez-Márquez, S.; da Silva, B.T.N.; Louvandini, H.; Abdalla, A.L.; de Azevedo Olival, A. Potential of Chloroleucon Acacioides Trees as an Alternative Feed Supplement for Grazing Ruminants in a Tropical Silvopastoral System. *J. Agric. Food Res.* **2023**, *11*, 100524. [\[CrossRef\]](#)
62. Kilgour, R.J. In Pursuit of “Normal”: A Review of the Behaviour of Cattle at Pasture. *Appl. Anim. Behav. Sci.* **2012**, *138*, 1–11. [\[CrossRef\]](#)
63. Wang, Y.; Jin, L.; Ominski, K.H.; He, M.; Xu, Z.; Krause, D.O.; Acharya, S.N.; Wittenberg, K.M.; Liu, X.L.; Stanford, K.; et al. Screening of Condensed Tannins from Canadian Prairie Forages for Anti-Escherichia Coli O157:H7 with an Emphasis on Purple Prairie Clover (*Dalea purpurea* Vent). *J. Food Prot.* **2013**, *76*, 560–567. [\[CrossRef\]](#) [\[PubMed\]](#)
64. Hoste, H.; Torres-Acosta, J.F.J.; Sandoval-Castro, C.A.; Mueller-Harvey, I.; Sotiraki, S.; Louvandini, H.; Thamsborg, S.M.; Terrill, T.H. Tannin Containing Legumes as a Model for Nutraceuticals against Digestive Parasites in Livestock. *Vet. Parasitol.* **2015**, *212*, 5–17. [\[CrossRef\]](#) [\[PubMed\]](#)
65. MacAdam, J.; Villalba, J. Beneficial Effects of Temperate Forage Legumes That Contain Condensed Tannins. *Agriculture* **2015**, *5*, 475–491. [\[CrossRef\]](#)
66. Tedeschi, L.O.; Muir, J.P.; Naumann, H.D.; Norris, A.B.; Ramírez-Restrepo, C.A.; Mertens-Talcott, S.U. Nutritional Aspects of Ecologically Relevant Phytochemicals in Ruminant Production. *Front. Vet. Sci.* **2021**, *8*, 155. [\[CrossRef\]](#)

67. Albores-Moreno, S.; Alayón-Gamboa, J.A.; Miranda-Romero, L.A.; Alarcón-Zúñiga, B.; Jiménez-Ferrer, G.; Ku-Vera, J.C.; Piñeiro-Vázquez, A.T. Effect of Tree Foliage Supplementation of Tropical Grass Diet on in Vitro Digestibility and Fermentation, Microbial Biomass Synthesis and Enteric Methane Production in Ruminants. *Trop. Anim. Health Prod.* **2019**, *51*, 893–904. [\[CrossRef\]](#)
68. Soltan, Y.A.; Morsy, A.S.; Sallam, S.M.A.; Lucas, R.C.; Louvandini, H.; Kreuzer, M.; Abdalla, A.L. Contribution of Condensed Tannins and Mimosine to the Methane Mitigation Caused by Feeding *Leucaena leucocephala*. *Arch. Anim. Nutr.* **2013**, *67*, 169–184. [\[CrossRef\]](#)
69. Pachas, N.A.; Radrizzani, A.; Murgueitio, E.; Uribe, F.; Zapata Cadavid, Á.; Chará, J.; Ruiz, T.E.; Escalante, E.; Mauricio, R.M.; Ramírez-Avilés, L. Establishment and Management of *Leucaena* in Latin America. *Trop. Grassl. Forrajes Trop.* **2019**, *7*, 127–132. [\[CrossRef\]](#)
70. Eckard, R.J.; Grainger, C.; de Klein, C.A.M. Options for the Abatement of Methane and Nitrous Oxide from Ruminant Production: A Review. *Livest. Sci.* **2010**, *130*, 47–56. [\[CrossRef\]](#)
71. Mueller-Harvey, I.; Bee, G.; Dohme-Meier, F.; Hoste, H.; Karonen, M.; Kölliker, R.; Lüscher, A.; Niderkorn, V.; Pellikaan, W.F.; Salminen, J.-P.; et al. Benefits of Condensed Tannins in Forage Legumes Fed to Ruminants: Importance of Structure, Concentration, and Diet Composition. *Crop Sci.* **2019**, *59*, 861–885. [\[CrossRef\]](#)
72. Schipanski, M.E.; Drinkwater, L.E. Nitrogen Fixation in Annual and Perennial Legume-Grass Mixtures across a Fertility Gradient. *Plant Soil* **2012**, *357*, 147–159. [\[CrossRef\]](#)
73. Delgado, J.A.; Groffman, P.M.; Nearing, M.A.; Goddard, T.; Reicosky, D.; Lal, R.; Kitchen, N.R.; Rice, C.W.; Towery, D.; Salon, P. Conservation Practices to Mitigate and Adapt to Climate Change. *J. Soil Water Conserv.* **2011**, *66*, 118A–129A. [\[CrossRef\]](#)
74. Mauricio, R.; Ribeiro, R.; Paciullo, D.; Cangussú, M.; Murgueitio, E.; Chará, J.; Estrada, M. *Silvopastoral Systems in Latin America for Biodiversity Environmental and Socioeconomic Improvements*; Elsevier: Amsterdam, Netherlands, 2019; ISBN 9780128110508.
75. Olival, A.D.A.; Souza, S.E.X.F.D.; Leles, G.M. Effects of Native Tree Species on Forage Quality and Availability in the Mato-Grossense Amazon. *Rev. Ibero-Americana Ciências Ambient.* **2021**, *12*, 146–156. [\[CrossRef\]](#)
76. Rivera, J.; Chará, J.; Arango, J.; Barahona, R. Effect of Different Genotypes of *Tithonia diversifolia* on Fermentation of Feed Mixtures with *Urochloa Brizantha* Cv. Marandú. *Crop Pasture Sci.* **2021**, *72*, 850–859. [\[CrossRef\]](#)
77. Van Sao, N.; Mui, N.T.; Van Binh, D. Biomass Production of *Tithonia diversifolia* (Wild Sunflower), Soil Improvement on Sloping Land and Use as High Protein Foliage for Feeding Goats. *Livest. Res. Rural Dev.* **2010**, *22*, 1–7.
78. García, I.R. Potencialidades de *Tithonia diversifolia* (Hemsl.) Gray En La Alimentación Animal. *Livest. Res. Rural Dev.* **2017**, *29*, 1–11.
79. Mahecha, L.; Rosales, M. Valor Nutricional Del Follaje de Botón de Oro (*Tithonia diversifolia* [Hemsl] Gray) En La Producción Animal En El Trópico. *Livest. Res. Rural Dev.* **2005**, *17*, 1–9.
80. Terry, S.A.; Ribeiro, R.S.; Freitas, D.S.; Delarota, G.D.; Pereira, L.G.R.; Tomich, T.R.; Maurício, R.M.; Chaves, A.V. Effects of *Tithonia diversifolia* on in Vitro Methane Production and Ruminant Fermentation Characteristics. *Anim. Prod. Sci.* **2016**, *56*, 437. [\[CrossRef\]](#)
81. Pérez-Márquez, S.; Ovani, V.S.; Lima, P.D.M.T.; Lana, Á.M.Q.; Louvandini, H.; Abdalla, A.L.; Maurício, R.M. *Tithonia diversifolia* Improves In Vitro Rumen Microbial Synthesis of Sheep Diets without Changes in Total Gas and Methane Production. *Agronomy* **2023**, *13*, 2768. [\[CrossRef\]](#)
82. Partey, S.T. Effect of Pruning Frequency and Pruning Height on the Biomass Production of *Tithonia diversifolia* (Hemsl) A. Gray. *Agrofor. Syst.* **2011**, *83*, 181–187. [\[CrossRef\]](#)
83. Chagas-Paula, D.A.; Oliveira, R.B.; Rocha, B.A.; Da Costa, F.B. Ethnobotany, Chemistry, and Biological Activities of the Genus *Tithonia* (Asteraceae). *Chem. Biodivers.* **2012**, *9*, 210–235. [\[CrossRef\]](#)
84. Mauricio, R.M. Feeding Ruminants Using *Tithonia diversifolia* as Forage. *J. Dairy, Vet. Anim. Res.* **2017**, *5*, 00146. [\[CrossRef\]](#)
85. Ovani, V.; Pérez-Márquez, S.; Evangelista, V.; Louvandini, H.; Abdalla, A. Root and Biomass Production of *Tithonia diversifolia* in Acidic Soil Levels. In Proceedings of the 57o Reunião da Sociedade Brasileira de Zootecnia—Tropical Animal Science and Practice to Feed the Planet, Campinas, Brazil, 25–29 July 2022.
86. Argüello-Rangel, J.; Mahecha-Ledesma, L.; Angulo-Arizala, J. Perfil Nutricional y Productivo de Especies Arbustivas En Trópico Bajo, Antioquia (Colombia). *Cienc. Tecnol. Agropecu.* **2020**, *21*, 1–20. [\[CrossRef\]](#)
87. Chin, N.; Hue, K. Supplementing *Tithonia diversifolia* with Guinea Grass or Tree Foliages: Effects on Feed Intake and Live Weight Gain of Growing Goats. *Livest. Res. Rural Dev.* **2012**, *24*, 188.
88. Durango, S.G.; Barahona, R.; Bolívar, D.; Chirinda, N.; Arango, J. Feeding Strategies to Increase Nitrogen Retention and Improve Rumen Fermentation and Rumen Microbial Population in Beef Steers Fed with Tropical Forages. *Sustainability* **2021**, *13*, 10312. [\[CrossRef\]](#)
89. Guatusmal-Gelpud, C.; Escobar-Pachajoa, L.D.; Meneses-Buitrago, D.H.; Cardona-Iglesias, J.L.; Castro-Rincón, E. Producción y Calidad de *Tithonia diversifolia* y *Sambucus Nigra* En Trópico Altoandino Colombiano. *Agron. Mesoam.* **2020**, *31*, 193–208. [\[CrossRef\]](#)
90. Lezcano, Y.; Soca, M.; Ojeda, F.; Roque, E.; Fontes, D.; Montejo, I.; Santana, H.; Martínez, J.; Cubillas, N. Caracterización Bromatológica de *Tithonia diversifolia* (Hemsl) A Gray En Dosetapas de Dos Etapas de Su Ciclo Fisiológico. *Pastos y Forrajes* **2012**, *35*, 275–282.
91. Londoño, J.M.B.; Carabalí, A.G.; Londoño, M.A.B. Yield, Agronomic Parameters and Nutritional Quality of *Tithonia diversifolia* in Response to Different Fertilization Levels. *Rev. Mex. Ciencias Pecu.* **2019**, *10*, 789–800. [\[CrossRef\]](#)

92. Mahecha, L.; Londoño, J.D.; Angulo, J. Agronomic and Nutritional Assessment of an Intensive Silvopastoral System: *Tithonia diversifolia*, *Sambucus nigra*, *Cynodon nlemfuensis*, and *Urochloa plantaginea*. *Proc. Natl. Acad. Sci. India Sect. B Biol. Sci.* **2022**, *92*, 37–47. [\[CrossRef\]](#)
93. Naranjo, J.; Cuartas, C. Nutritional Characterization and Ruminant Degradation Kinetics of Some Forages with Potential for Ruminants Supplementation in the Highland Tropics of Colombia. *CES Med. Zootec* **2005**, *6*, 9–19.
94. Verdecia, D.; Ramirez, J.; Leonard, I.; Alvarez, Y.; Bazán, Y.; Bodas, R.; Andrés, S.; Alvarez, J.; Giraldez, F.; Lopez, S. Nutritive Value of the *Tithonia diversifolia* in a Location of Valle Del Cauto. *Rev. electrón. Vet* **2011**, *12*, 1–13.
95. Andries, J.I.; Buyse, F.X.; De Brabander, D.L.; Cottyn, B.G. Isoacids in Ruminant Nutrition: Their Role in Ruminant and Intermediary Metabolism and Possible Influences on Performances—A Review. *Anim. Feed Sci. Technol.* **1987**, *18*, 169–180. [\[CrossRef\]](#)
96. Zebeli, Q.; Terrill, S.J.; Mazzolari, A.; Dunn, S.M.; Yang, W.Z.; Ametaj, B.N. Intraruminal Administration of *Megasphaera elsdenii* Modulated Rumen Fermentation Profile in Mid-Lactation Dairy Cows. *J. Dairy Res.* **2012**, *79*, 16–25. [\[CrossRef\]](#)
97. Sampaio, C.B.; Detmann, E.; Paulino, M.F.; Valadares Filho, S.C.; de Souza, M.A.; Lazzarini, I.; Rodrigues Paulino, P.V.; de Queiroz, A.C. Intake and Digestibility in Cattle Fed Low-Quality Tropical Forage and Supplemented with Nitrogenous Compounds. *Trop. Anim. Health Prod.* **2010**, *42*, 1471–1479. [\[CrossRef\]](#) [\[PubMed\]](#)
98. Tambara, A.A.C.; Härter, C.J.; Rabelo, C.H.S.; Kozloski, G.V. Effects of Supplementation on Production of Beef Cattle Grazing Tropical Pastures in Brazil during the Wet and Dry Seasons: A Meta-Analysis. *Rev. Bras. Zootec.* **2021**, *50*, 1–22. [\[CrossRef\]](#)
99. Cardona Iglesias, J.L.; Mahecha Ledesma, L.; Angulo Arizala, J. Efecto Sobre La Fermentación in Vitro de Mezclas de *Tithonia Diversifolia*, *Cenchrus Clandestinum* y Grasas Poliinsaturadas. *Agron. Mesoam.* **2017**, *28*, 405. [\[CrossRef\]](#)
100. Akanmu, A.M.; Hassen, A.; Adejoro, F.A. Gas Production, Digestibility and Efficacy of Stored or Fresh Plant Extracts to Reduce Methane Production on Different Substrates. *Animals* **2020**, *10*, 146. [\[CrossRef\]](#)
101. Folley, S.J.; French, T.H. Acetate as a Possible Precursor of Ruminant Milk Fat, Particularly the Short-Chain Fatty Acids. *Nature* **1949**, *163*, 174–175. [\[CrossRef\]](#)
102. Lin, M.; Jiang, M.; Yang, T.; Tan, D.; Hu, G.; Zhao, G.; Zhan, K. Acetate-Induced Milk Fat Synthesis Is Associated with Activation of the MTOR Signaling Pathway in Bovine Mammary Epithelial Cells. *Animals* **2022**, *12*, 2616. [\[CrossRef\]](#)
103. Rivera, J.E.; Villegas, G.; Chará, J.; Durango, S.G.; Romera, M.A.; Verchot, L. Effect of *Tithonia diversifolia* (Hemsl.) A. Gray intake on in vivo methane (CH₄) emission and milk production in dual purpose cows in the Colombian Amazonian piedmont. *Transl. Anim. Sci.* **2022**, *6*, 1–12. [\[CrossRef\]](#)
104. Herrera, J.P.N.; Preston, T.R.; Edmundo, J.; Guerrero, A.; Riascos, R. Producción de Metano En Una Incubación in Vitro de *Cenchrus Clandestinus* y *Lolium Hybridum* Suplementado Con *Tithonia diversifolia* En El Trópico Alto Del Departamento Del Putumayo, Colombia. *Livest. Res. Rural Dev.* **2020**, *32*, 1–6.
105. Holguín, V.A.; Cuchillo-Hilario, M.; Mazabel, J.; Quintero, S.; Mora-Delgado, J. Efecto de La Mezcla Ensilada de *Penisetum Purpureum* y *Tithonia Diversifolia* Sobre La Fermentación Ruminal in Vitro y Su Emisión de Metano en el Sistema RUSITEC. *Rev. Mex. Ciencias Pecu.* **2020**, *11*, 19–37. [\[CrossRef\]](#)
106. Pérez-Márquez, S.; Ribeiro, R.S.; Abdalla, A.L.; Lana, A.M.Q.; Maurício, R.M. Does *Tithonia diversifolia* Influences in Vitro Gas Production Parameters and Nutritive Value of Graminous *Cynodon* Spp and *Penisetum purpureum* When Associated? In *Proceedings of the X Congreso Internacional Sobre Sistemas Silvopastoriles: Por una Producción Sostenible*; CIPAV Editorial Asunción: Asunción, Paraguay, 2019; pp. 493–495.
107. Olayinka, B.; Raiyemo, D.; Etejere, E. Phytochemical and Proximate Composition of *Tithonia diversifolia* (Hemsl.) A Gray. *Ann. Food Sci. Technol.* **2015**, *16*, 195–200.
108. Moreira, G.D.; Lima, P.D.M.T.; Borges, B.O.; Primavesi, O.; Longo, C.; McManus, C.; Abdalla, A.; Louvandini, H. Tropical Tanniniferous Legumes Used as an Option to Mitigate Sheep Enteric Methane Emission. *Trop. Anim. Health Prod.* **2013**, *45*, 879–882. [\[CrossRef\]](#) [\[PubMed\]](#)
109. Goel, G.; Makkar, H.P.S. Methane Mitigation from Ruminants Using Tannins and Saponins. *Trop. Anim. Health Prod.* **2012**, *44*, 729–739. [\[CrossRef\]](#) [\[PubMed\]](#)
110. Lima, P.D.M.T.; Filho, A.L.A.; Issakowicz, J.; Ieda, E.H.; Corrêa, P.S.; de Mattos, W.T.; Gerdes, L.; McManus, C.; Abdalla, A.L.; Louvandini, H. Methane Emission, Ruminant Fermentation Parameters and Fatty Acid Profile of Meat in Santa Inês Lambs Fed the Legume Macrotiloma. *Anim. Prod. Sci.* **2020**, *60*, 665. [\[CrossRef\]](#)
111. Yousuf, B.M.; Adefunmilayo Adeloye, A.; Kehinde Okukpe, M.; Dauda Adeyemi, K.; Julius Ogundun, N. Influence of Dietary Sunflower (*Tithonia diversifolia*) Leaf Extracts on Performance Characteristics of Goats Fed Cassava Peeling Wastes-Based Diet. *J. Agric. Technol.* **2014**, *10*, 59–65.
112. Chacón Góngora, P.A. Uso de *Tithonia Diversifolia* Como Forraje Alternativo Para La Reducción de Emisiones de Óxido Nitroso En Excretas de Vacas. CaTIE: Turrialba, Costa Rica, 2018.
113. Ramírez-Rivera, U.; Sanginés-García, J.R.; Escobedo-Mex, J.G.; Cen-Chuc, F.; Rivera-Lorca, J.A.; Lara-Lara, P.E. Effect of Diet Inclusion of *Tithonia diversifolia* on Feed Intake, Digestibility and Nitrogen Balance in Tropical Sheep. *Agrofor. Syst.* **2010**, *80*, 295–302. [\[CrossRef\]](#)
114. Castañeda Serrano, R.D.; Piñeros Varón, R.; Vélez Giraldo, A. Foliage of Tropical Arboreal Species in Feeding Ovines (*Ovis aries*): Intake, Digestibility and Balance Nitrogen. *Boletín Científico Cent. Museos Mus. Hist. Nat.* **2018**, *22*, 58–68. [\[CrossRef\]](#)

115. Fajemisin, A.N.; Salihu, T.; Fadiyimu, A.; Alokun, A. Dietary Effect of Substituting Panicum Maximum with Tithonia Diversifolia Forage on Performance of Yankasa Sheep. In Proceedings of the XXII International Grassland Congress, Sidney, Australia, 15–19 September 2019; pp. 563–564.
116. Rivera, J.E.; Villegas, G.; Chará, J.; Durango, S.; Romero, M.; Verchot, L. Silvopastoral Systems with *Tithonia diversifolia* (Hemsl.) A. Gray Reduce N₂O–N and CH₄ Emissions from Cattle Manure Deposited on Grasslands in the Amazon Piedmont. *Agrofor. Syst.* **2023**, *98*, 1091–1104. [[CrossRef](#)]
117. Odedire, J.; Oloidi, F. Feeding Wild Sunflower (*Tithonia diversifolia* Hemsl., A. Gray) to West African Dwarf Goats as a Dry Season Forage Supplement. *World J. Agric. Res.* **2014**, *2*, 280–284. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.